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Federal Contaminated Sites Action Plan (FCSAP): Ecological Risk Assessment Guidance Module 3: Standardization of Wildlife Receptor Characteristics

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Module 3: Uniformisation des caractéristiques des récepteurs fauniques

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Acknowledgments

1. BACKGROUND

The Federal Contaminated Sites Action Plan (FCSAP) was developed to support federal departments, agencies and consolidated crown corporations to reduce the risks to human health and the environment, as well as to reduce the financial liabilities associated with federal contaminated sites. Under FCSAP, ecological risk assessments (ERAs) are commonly used as a site management tool at federal contaminated sites. The FCSAP Ecological Risk Assessment Focus Group is developing guidance for ERA supplemental to the existing CCME guidance

(1996, 1997). The FCSAP ERA guidance consists of a comprehensive main ERA guidance document (Environment Canada [EC] 2012) and several specific technical guidance modules.

This document is a technical guidance module on standardization of wildlife receptor characteristics used to quantify contaminant exposure via oral intake (see Section 3.3.3 of the main guidance document – EC 2012). In this document, wildlife refers to birds and mammals, as well as reptiles and amphibians. This

Important Information:

This module is **not** intended to standardize the process of selection of receptors in ERA. Rather, it should be used as a resource *after* the appropriate receptors of concern (ROCs) have already been selected. In all cases, the selection of ROCs in the problem formulation phase of the ERA should be based on site-specific considerations. In addition, while some receptor characteristics could be expected to vary little among sites (e.g., adult body weight), others may vary considerably among sites (e.g., diet and foraging range size). For these latter characteristics, the information provided in this module should be viewed as a default that, depending on the ERA, may need to be tailored on a site-specific basis. For sites where precision in estimates of wildlife risks is important for guiding risk management decisions, dietary composition and foraging range should be developed on a site-specific basis.

module identifies 27 wildlife species commonly used in ERAs throughout Canada which use both terrestrial and aquatic environments, and provides information on specific receptor characteristics that can be applied in ERAs on federal contaminated sites. Providing standardized receptor characteristics for ERAs to federal custodians and their consultants is intended to improve national consistency in the management of federal contaminated sites.

1.1. Receptor Characteristics in ERA

In ERA, a receptor of concern (ROC)¹ can be any non-human individual, population, community, habitat or ecosystem that is potentially exposed to contaminants of concern (COCs). COCs are

¹ The term Valued Ecosystem Component (VEC) has the same or similar meaning, but is not generally used by ERA practitioners.

those contaminants that have been selected for evaluation in the ERA². The level of biological organization at which an ROC is defined varies. In the case of lower trophic levels, the community is often identified as the ROC (e.g., invertebrate community, grassland community). In the case of higher trophic levels, the ROC is usually defined at the species level (individual organism or population – e.g., mink, eagle). In the latter case, a species may be selected for direct assessment of that species (i.e., assuming that the species chosen is of particular interest to risk managers) or for use as a representative (or surrogate) for similar organisms. For wildlife receptors, the focus of this Technical Module, a surrogate ROC can be used in the ERA to represent risks to a type of receptor with particular characteristics (e.g., a shrew may be used as a surrogate ROC for insectivorous mammal). In most cases, the receptor types are selected on the basis of functional feeding groups (e.g., small omnivorous mammals, piscivorous birds) rather than on taxonomic linkages.

In selecting a specific surrogate ROC, the risk assessor considers the degree to which the ROC may be assumed to be representative and/or protective of other similar receptors on the basis of contaminant sensitivity and similarities in characteristics such as diet and foraging range (see Section 2.2.5 of the main guidance document – EC 2012). More than one surrogate ROC may be selected, particularly where a particular ROC has a higher relative sensitivity to the COCs (see Section 2.2.5.1 of the main guidance document – EC 2012) or is rare or endangered, or has some similar status (e.g., consult the *Species at Risk Act (SARA)* and provincial lists). Once the surrogate wildlife ROCs are chosen, receptor-specific characteristics such as body weight, feeding behaviour, ingestion rates (food, water and soil/sediment), and habitat range are used to estimate the total (i.e., multi-media) dose for each COC (see Section 3 of the main guidance document – EC 2012). The total dose estimate is then compared to a Toxicity Reference Value (TRV; see Technical Module 2 on Selection or Development of Site-Specific Toxicity Reference Values) or to dose-response data when characterizing risks (see Section 5.3.2 of the main guidance document – EC 2012).

1.2. Scope of Module

This module provides standardized receptor characteristics for wildlife species (as defined in **Section 1** and the glossary) commonly used in ERAs throughout Canada, which use both aquatic and terrestrial environments; these characteristics are used to quantify COC exposure via oral intake.

In contrast to wildlife ROCs, other receptor groups, particularly lower trophic levels (e.g., plants, soil invertebrates), are usually assessed by evaluating the concentration of contaminants in external exposure media (e.g., water, soil, sediment), rather than total dietary dose, thus details of receptor characteristics are often not relevant for estimating exposure for these lower trophic levels. While there may be exceptions (e.g., an ERA where a particular plant community is an

² In some jurisdictions, terms such as COPC (Contaminants of Potential Concern) or PCOC (Potential Contaminant of Concern) refer to the initial list of substances considered, whereas the term Contaminants

of Concern (COC) is used to refer to the final list after the selection process conducted as part of the problem formulation. In other jurisdictions the term COC is not used at all and the final list is referred to as the list of COPCs. In this guidance document, the term COC refers to the final list of substances retained

for the risk assessment at the end of problem formulation.

important receptor), cases where detailed receptor characteristics for these lower trophic level organisms are important would be rare, and in those cases the receptor characteristics should probably be defined on a site-specific basis. These lower trophic level receptor groups are not addressed in this document.

In the past, most ERAs addressing "wildlife" have focused only on birds and mammals. Amphibians and reptiles were rarely assessed for COC exposure *via* oral intake due to a general lack of toxicological data. While there are still broad gaps and uncertainties, there is a growing database of

US EPA Wildlife Exposure Handbook (1993):

This document is intended to provide standard values for wildlife receptors across Canada, and as such, should be consulted prior to the US EPA Wildlife Exposure Handbook (1993). However, these two resources should be considered complementary as: (1) this document references US EPA (1993) allometric equations or receptor-specific information where no Canadian information was found; and (2) the US EPA (1993) main document and appendices contain information not provided herein.

toxicological information specific to amphibians and reptiles (Sparling et al. 2010). Consequently, these taxa are also included in this Technical Module.³ The objective of this module is to provide default standard values for wildlife receptor characteristics for use by the ERA practitioner, particularly for preliminary assessments where habitat information may not yet be available. However, since many wildlife species can exist in a variety of habitats, some specific characteristics such as diet and foraging range may vary substantially from habitat to habitat (and across Canada), as well as seasonally (see **Section 2.1.9** for a discussion of the seasonal aspects of receptor-specific characteristics relevant to exposure estimation). Therefore, it may be important for the ERA practitioner to get site-specific information for some receptor characteristics, depending on the needs of a specific ERA. To recognize this variability in receptor characteristics, ranges of values for some characteristics are provided where appropriate, along with a default standard value. The ERA practitioner should always provide appropriate rationale when choosing the values of the specific characteristics (regardless of whether default or other values are chosen).

1.3. Receptor Characteristics Covered in this Module

A number of receptor-specific characteristics have the potential to affect the degree to which ROCs are exposed to the contaminants of potential concern (COCs) on the site (see Section 2.2.5 of the main guidance document – EC 2012). The characteristics covered in this module include the following:

- Habitat type (e.g., forest, river, ocean);
- Foraging range;
- Body weight (BW);

³ TRVs for amphibians and reptiles are limited; however, total dose estimates can be evaluated along spatial gradients or compared between on-site and reference conditions.

- Receptor type (i.e., feeding guild, such as carnivore, herbivore, or omnivore) and foraging behaviour (e.g., hunting, grazing);
- Food type and dietary proportions;
- Feeding rate (amount of food typically consumed per defined time);
- Rate of drinking water ingestion (amount of water typically consumed per defined time); and
- Rate of incidental soil/sediment ingestion (amount of soil/sediment typically consumed per defined time).

As noted in **Section 1.2**, the focus of this module is on receptor characteristics relevant for determining exposure *via* oral intake, which is the only exposure route addressed for wildlife ROCs in the majority of ERAs. Characteristics or behaviours that may be important for quantifying other potential exposure routes (e.g., dermal absorption or inhalation) are discussed in **Section 2.1.9**, but have not been included in the receptor-specific tables (**Appendix A**). Additionally, contaminant bioavailability (although specific to contaminant type rather than receptor), is also discussed below (**Section 2.1.9**) since it is an important parameter for estimating total dose.

Receptor characteristics provided in this Technical Module apply to adult organisms – considerations relevant to juvenile life stages are discussed in **Section 2.1**.

1.4. Receptors Covered in this Module

The wildlife receptors included in this module were selected based on input obtained from: (1) ERA practitioners across Canada (from private practice), including practitioners with experience conducting risk assessments in the northern territories; and (2) information obtained from the Centre d'expertise en analyse environnementale Quebec (CEAEQ) regarding their list of wildlife receptors (Paramètres d'exposition chez les mammifères, oiseaux, CEAEQ 2006a, 2006b).

In many cases, mammalian and avian receptor species chosen by different practitioners were the same. In other cases, different species of the same receptor group were chosen. Where there was little agreement among consultants on the particular species (e.g., for shrews, mice, hawks and falcons), a species with: (1) the widest distribution across Canada; and (2) presence in a wide diversity of habitats was chosen. The primary intent of receptor selection was not necessarily to obtain receptors from each feeding guild and habitat type, but rather to identify the receptors most commonly used. Nevertheless, receptors were chosen to fill as many feeding guilds (e.g., omnivore, carnivore, herbivore) as possible, to encompass a variety of terrestrial and aquatic habitats, and to ensure relevance to most ERAs. Importantly, as noted in **Section 1.2**, one amphibian and one reptile species were included to facilitate the use of these receptors at federal sites, as appropriate. The aquatic and terrestrial receptors chosen for this module are categorized into types according to most relevant feeding guild (**Tables 1 and 2**).

 Table 1. Selected wildlife ROCs for aquatic ecosystems

		Surroga	ate ROCs
Aquatic Receptor Group	Aquatic Receptor Type	Marine	Freshwater
Mammals	Herbivore		muskrat; moose
	Insectivore		
	Piscivore/Carnivore	river otter	American mink, river otter
	Omnivore		
Birds	Herbivore	mallard duck	mallard duck
	Insectivore/Invertivore	spotted sandpipers, lesser scaup	spotted sandpipers, lesser scaup
	Piscivore/Carnivore	bald eagle, great-blue heron, common loon	common merganser, bald eagle, great-blue heron, common loon
	Omnivore	mallard duck	mallard duck

 Table 2. Selected wildlife ROCs for terrestrial ecosystems

		Surrogate ROCs
Terrestrial Receptor Group	Terrestrial Receptor Type	Terrestrial
Mammals	Herbivore	meadow vole, white-tailed deer, snowshoe hare
	Insectivore	common shrew
	Carnivore	red fox, short-tailed weasel
	Omnivore	black bear, deer mouse
Birds	Herbivore	ruffed grouse, spruce grouse
	Insectivore	barn swallow
	Carnivore	red-tailed hawk, peregrine falcon
	Omnivore	American robin
Amphibians	Carnivore	wood frog
Reptiles	Carnivore	common garter snake

2. GUIDANCE

This section describes each receptor characteristic and introduces the receptor-specific tables (**Appendix A**). Detailed guidance on how each of the receptor characteristics is used in ERA is provided in Section 3 of the main guidance document (EC 2012).

Information on ROCs was obtained from a thorough review of existing literature, including inhouse reference guides, on-line journals, government reports, articles and academic dissertations. Where available, information from Canadian studies was used to provide information on preferred habitat, foraging range size, diet and body weight. For foraging range, the minimum and maximum ranges encountered in the literature were included. The minimum foraging range, which could represent range in the breeding or wintering seasons, was also highlighted in order to be as conservative as possible. For body weight, the minimum and maximum weights were given, while the average weight was most often provided in the literature as being representative of most individuals. Diet proportions were based on a careful review of relevant literature and professional judgment. Ingestion rates were based on either allometric scaling or species-specific estimates. Literature on species-specific food ingestion rates for wildlife was preferred where details regarding age and gender of the animals were provided.

2.1. Review of Receptor-Specific Characteristics

As the majority of ERAs use adult organisms, the receptor characteristics provided in the receptor-specific tables and discussed below are for adult ROCs. In some instances, an assessment of juveniles (and hence juvenile-specific characteristics) may be appropriate, particularly when available TRVs are based solely on juvenile endpoints such as growth. However, receptor characteristics may vary greatly between juveniles and adults (and among adults) of the same species (e.g., body weight, diet) and exposure to COCs may be quite different. Receptor-specific characteristics for juveniles are not provided in this module (some information on juveniles, such as body weight, is provided in CEAEQ 2006a & 2006b, US EPA 1993 and OSWER 2005, Attachment 4-3, Table 20).

Additionally, many of the receptor characteristics discussed below vary seasonally (see **Section 2.1.9**), and by size, reproductive status and habitat. Although default numbers are provided for receptor characteristics where appropriate, ranges are also provided for situations where the default value may need to be adjusted.

2.1.1. Habitat Type

Information regarding habitat types is useful when identifying appropriate ROCs for a site, and will provide an indication of probable diet for the ROCs under consideration. Additionally, site use patterns by receptors will vary according to available habitat types (i.e., preferred habitat will be subject to the highest use). The configuration of high-use habitat types relative to the pattern of contamination will affect ROC exposure and may determine whether a ROC spends more time on-site or off-site, depending on the relative suitability of the habitat (see Section 3.4 of the main document for a discussion on spatially explicit exposure models). The ASTM-International Standard Guide for Estimating Wildlife Exposure Using Measures of Habitat Quality (Designation E2385-11) provides a useful discussion on when habitat modifications of exposure

estimates would reduce uncertainty (particularly where both habitat and contaminant distribution are heterogeneous), and may be more useful than modifications based solely on foraging range.

2.1.2. Foraging Range

The term "foraging range" is used in this module preferentially to the term "home range". In many cases, these two terms can be synonymous; however, foraging range is specific to the area typically explored by an animal while it is feeding (ASTM E2385-11), while the home range would typically also include activities such as denning. In the case of migratory birds, the home range can be extensive (spanning continents), while the foraging range is specific to local seasonal habitat (i.e., for breeding). For this module, the foraging range was considered more relevant to oral dose calculations than home range.

The foraging range of each receptor is usually used in ERA relative to the <u>size of the site</u> (or relevant portion of the site) under assessment. The ratio of site size to foraging range is often used as a modifying factor to total dose calculations. In some cases, a conservative screening assessment may assume that a receptor spends all of its time on-site, but more realistic assessments should apportion exposure between on-site and the surrounding 'off-site' areas, and may also include modifications of exposure estimates based on relative quality of habitat (**Section 2.1.1**, above and Section 3.4 of the main document), where warranted.⁴

Estimation of off-site contributions should be based on data (or a well-informed decision) rather than on a default assumption that off-site contributions are negligible. This is particularly important for large mammals or other receptors which may spend only a very small portion of time foraging on a site.

The default foraging range provided for each ROC is a conservative estimate based on an up-to-date literature review. However, foraging ranges can vary in size for a number of reasons, such as habitat quality in the general area, seasonal availability of foods, gender and rearing of young. Ranges are provided where site-specific considerations are important (e.g., if habitat quality is low, range size may be larger)⁵.

Although some wildlife (birds in particular) are migratory and may spend only a few months or weeks using a site before leaving for wintering or breeding grounds, migratory receptors should be considered as ROCs in many cases (see Section 2.2.5 of the main guidance document – EC 2012). Exposure estimation for migratory species requires careful consideration of foraging range (see Section 2.1.9 below). In all cases, rationale (based on best professional judgment) for the selection of a foraging range should be provided.

⁴ Where ROCs may have a large foraging range compared to small areas of high COC concentrations, acute exposure (and thus TRVs based on acute exposure) may be relevant.

⁵ The ERA practitioner will need to exercise best professional judgement when choosing a home range size, depending on the specific needs of the risk assessment.

2.1.3. Body Weight

Body weight values are usually used in allometric equations⁶ to determine ingestion rates. In the receptor-specific tables, body weight data were derived from up-to-date literature on each species (as recommended in the main guidance document – EC 2012). The US EPA Wildlife Exposure Factors Handbook (US EPA 1993) also has body weight values for a variety of species, which can be used as a default for ROCs not considered in this module.

2.1.4. Feeding Guild and Foraging Behaviour

General information on feeding guild and foraging behaviour is useful when selecting ROCs for an ERA. Wildlife belonging to different feeding guilds (e.g., omnivore, carnivore, piscivore) may have different exposure to COCs as different dietary items may contain higher or lower COC concentrations. Additionally, foraging behaviour may influence the ROC's exposure to contaminated soil or sediment. For example, a dabbling duck such as a mallard forages near the sediment/water interface and is more exposed to sediment-related contamination than a piscivorous waterfowl such as the common loon.

2.1.5. Food Types and Dietary Proportions

Most receptors consume more than one type of food. Although a conservative screening assessment may use the most contaminated food type to calculate total dose from food, more realistic assessments should consider dietary proportions for any receptor that consumes more than one type of food. Dietary proportions provided in receptor-specific tables were based on an up-to-date literature review; as mentioned earlier, these should often be modified based on site-specific considerations. For many receptors there is seasonal variation in diet (see **Section 2.1.9** below).

2.1.6. Ingestion Rate for Food

Ingestion rates for food are typically characterized as mg or kg/kg body weight/day. Food ingestion rates can vary by size and gender, and by seasonal changes in ambient temperature, activity levels, reproductive activities, and the type of diet consumed. Additionally, captive animals may have higher or lower ingestion rates than free-living animals, depending on the diet provided. However, detailed information on changes in food ingestion rates based on the above factors is not readily available. Typically, the most commonly used resources for food ingestion rates in ERA include US EPA (1993) and Sample et al. (1996). Where possible, species-specific ingestion rates were provided otherwise values derived from the allometric equations provided by the US EPA (1993; equations derived from Nagy 1987) have been included in the receptor-specific tables as a default. Some species-specific ingestion rates for food are on a wet weight

⁶ Allometric equations are considered acceptable for use in calculating wildlife food and water ingestion rates (as used by the US EPA [1993]). In contrast, allometric scaling of TRVs is generally not considered good practice (see Allard et al. 2010 and also Technical Module 2 on Selection or Development of Site-Specific Toxicity Reference Values).

⁷ An exhaustive literature search was not conducted, but searches were conducted in Biosis Previews, JSTOR (www.jstor.org), and using the search engine Google Scholar®.

basis (but not all), while allometric equations are on a dry weight basis⁸ (see footnote 4 regarding appropriate use of allometric scaling). The US EPA (1993; Tables 4-1 and 4-2) provides the water composition of some plant and animal wildlife foods, which could be used as an initial default where site-specific values are unavailable for conversion between wet and dry weights.

2.1.7. Ingestion Rate for Water

Ingestion rates for water are typically characterized as L/kg body weight/day and are used in total dose calculations. Water ingestion rates can vary by body weight, physiological adaptations, diet, temperature and activity levels (US EPA 1993). Similar to the lack of recent literature for wildlife food ingestion rates, recently published studies quantifying wildlife water ingestion rates were not identified. Therefore, values derived from the allometric equations provided by the US EPA (1993; equations derived from Calder and Braun 1983) have been included in the receptor-specific tables as a default (see footnote 4 regarding the appropriate use of allometric scaling).

2.1.8. Incidental Ingestion Rates for Soil and Sediment

Mammals and birds may incidentally ingest COCs in soil or sediment ingestion (e.g., during grooming, when consuming soil-covered plant roots or when dabbling for invertebrates). Information on this factor is typically lacking, but some soil ingestion rates are provided in US EPA (1993) and Beyer et al. (1994). The soil ingestion rates are provided as a percentage of soil in the diet on a dry weight basis. As these rates are based on scat analyses, they include soil ingested along with food, as well as soil incidentally ingested during grooming or digging. If for a certain receptor a soil or sediment ingestion rate is not provided in Appendix A and other reliable estimates are not available a default rate of 2% soil in the diet on a dry weight basis may be assumed for most receptors.

2.1.9. Seasonal Aspects of Receptor-Specific Characteristics Relevant to Exposure Estimation

Many wildlife characteristics relevant to exposure estimation (e.g., body weight, foraging range, food type and dietary proportions) may vary substantially from season to season. The values chosen for these characteristics have the potential to substantially alter the outcome of an ERA.

When considering values for receptor-specific characteristics used for exposure estimation, it is relevant to consider both: (1) the time scale over which the characteristic varies; and (2) the toxicological basis of the TRV used for each COC. Importantly, exposure estimates should not be modified by assigning a proportion of the year for which a ROC occupies a site (e.g., receptors

⁸ Wet weight food ingestion rates must be paired with wet weight contaminant concentrations, while dry weight food ingestion rates must be paired with dry weight contaminant concentrations. Alternatively, dry or wet weight food ingestion values can be converted using literature-based or site-specific specific moisture content of the food types ingested by the ROC.

⁹ If food tissue items collected for risk assessment purposes are unwashed (see Section 3.3.3 of the main guidance document), then there is some double-counting of ingested soil volumes.

which migrate or hibernate). Below we specifically consider migration, hibernation and food types and dietary proportions, as they relate to exposure estimation and TRV derivation.

Migration - When a migratory species is chosen as a ROC for a site, the exposure to COCs may be less than for non-migratory wildlife, depending on the length of time the ROC is present onsite. However, as a default, migration should not be used to dilute assumed exposure unless the ERA practitioner knows that the effects endpoints (e.g., TRVs) for a particular COC occur on a very long timescale. Most studies of chronic effects are on shorter timeframes than 6 months, and most available TRVs are derived as intake per unit body weight per day. Therefore, in most cases, there is no basis for diluting exposure based on migratory patterns.

Hibernation/torpor – As with migratory species, a wildlife receptor that hibernates (e.g., frog) or goes into torpor (e.g., bear) may have lower exposure to COCs, depending on the length of time spent in hibernation and whether it wakes to feed periodically. However, as with migration, hibernation should not be used to dilute assumed exposure, for the same reasons – unless effects are known to occur only over very long time frames, it should be assumed that a ROC may receive harmful exposure during its active season.

Food types and Dietary Proportions – Many wildlife species have a seasonal component to their diet. Some foods (such as berries or spawning salmon) are only available seasonally. In general, dietary proportions which reflect an average diet over the course of the year and include seasonal items (such as those presented in the receptor-specific tables in Appendix A) are appropriate for assessing risks. However, a season-specific diet assessment may be appropriate when: (1) there are considerable seasonal variations in diet; (2) COC concentrations in dietary items also vary considerably; and (3) effects may occur over a relatively short time scale (e.g., a few weeks to months). In such a case, it may be important for the ERA practitioner to refine seasonal estimates not only for the diet items but also for other receptor-specific characteristics, such as the seasonal foraging range¹¹.

2.1.10. Other Factors Relevant to Exposure Estimation

There are other factors that may affect total dose but that are often primarily contaminant-specific (but also highly dependent on the receptor). Three such factors include dose from dermal exposure, dose from inhalation exposure, and bioavailability of COCs in the gut of wildlife.

Dermal exposure (direct contact with soil and sediment) of wildlife should be considered when relevant, for COCs that can be absorbed readily through this pathway. Dermal exposure can also be a relevant exposure pathway for amphibians and reptiles. Detailed guidance on how to assess dermal exposure is limited (SAB-CS 2008, Suter 1996). Approaches for this pathway should be taken on a site-specific basis with appropriate rationale and consultation (see Section 2.2.6 of the main guidance document – EC 2012).

¹⁰ Hibernation and migration have not been included in the module as receptor-specific characteristics (**Section 2.1**), but given the potential implications to exposure estimation, have been included here for discussion.

¹¹ Although in most cases a chronic TRV would apply (most studies of chronic effects are on shorter timeframes than 6 months), the use of an acute TRV should be considered where a dietary component is very time-limited.

Inhalation exposure through wind-blown dust or inhalation of vapours can be a relevant pathway for some mammals, birds, reptiles and amphibians. In practice, this pathway has not been commonly assessed, but may be required in some jurisdictions in future, and should be considered where the conceptual model indicates potential widespread exposure. For example, a site with high concentrations of volatile compounds and good small mammal habitat may warrant consideration of vapour inhalation. Inhalation toxicity data are currently lacking for most contaminants, but some jurisdictions are developing guidance and screening values for soil and vapour. In addition, because small mammals generally construct their burrows to allow for air flow, characterizing exposure may be challenging (see Section 2.2.6 of the main guidance document – EC 2012). Detailed guidance on how to assess inhalation exposure to wildlife is limited. Approaches for this pathway should be taken on a site-specific basis with appropriate rationale and consultation.

Finally, total dose estimates can be adjusted to account for incomplete uptake or absorption of COCs in the gut of wildlife (i.e., incomplete bioavailability). Logically, most TRVs are developed from studies conducted using readily bioavailable forms of contaminant (e.g., soluble metal salts). Unless accounted for, the degree to which the COC form found at the site is less bioavailable than the form used in the TRV development will translate directly into over-estimated risks. On the human health side, physiologically-based extraction procedures (PBET; e.g., Ruby et al. 1996) have been developed and refined (Ruby et al. 1999, Nico et al., 2006) over the past 15 years that help to quantify bioaccessibility (i.e., the degree to which a substance in soil/sediment is released into solution and available for absorption) of specific COCs (certain metals only). While similar efforts have been attempted to support ERA, particularly in characterizing bioaccessibility of mining-related contamination, standard procedures and protocols have yet to be developed. Given the large uncertainties associated with this issue, incorporation of realistic bioaccessibility estimates should be conducted where defensible. That said, in the absence of specific quantitative information regarding site-specific bioaccessibility, risk assessors should assume that 100% of the COC is available (see Section 3.3.3 of the main guidance document – EC 2012).

2.2. Receptor-Specific Tables

Tables of receptor-specific characteristics for each of the 27 species identified (**Tables 1 and 2**) are provided in **Appendix A.** The tables include each of the specific characteristics, a default value and a range of values where appropriate, and a written commentary with references. In some cases, particularly for reptiles and amphibians, information was unavailable or scarce for some of the characteristics; this was identified in the tables. References for each receptor are provided after each receptor-specific table, it is recommended that consultants review the literature to ensure project relevance.

- Estimates in the receptor-specific tables for body weight and ingestion rates are presented as follows:
 - O An average body weight is provided as a default; ranges for males and females are provided as appropriate/available. Body weights are presented in grams (g) or kilograms (kg) as appropriate depending on the weight of the animal. Average body weights were rounded to the first decimal place; or in the case of heavier wildlife (i.e., bear, moose and deer), to the nearest kilogram.

- o Food ingestion rates are provided in kg food (either dry weight or wet weight based on the source) per kg body weight per day. No attempt was made to standardize these rates to either dry or wet weight; conversions should be made on a site specific basis based on the moisture content of the diet of the ROC (see Section 2.1.6). Food ingestion rates were rounded to two decimal places.
- Water ingestion rates are provided in liters (L) per kg body weight per day and were rounded to two decimal places.
- The default foraging range provided for each ROC is a conservative estimate (smallest area) based on an up-to-date literature review. Ranges are provided where site-specific considerations are important (e.g., if habitat quality is low, range size may be larger). Foraging range is presented in square kilometers (km) /meters (m) as well as hectares, depending on the receptor. For some receptors which forage along the water, linear foraging range was also provided (in km or m). Foraging ranges were rounded to the first or second decimal place, as appropriate depending on the range size and reporting unit (or to the nearest whole number where ranges were large).
 - o For migratory birds which do not winter in Canada (spotted sandpiper and barn swallow), the breeding season range size was provided.
 - o For migratory birds which may both breed and winter in Canada (mallard, lesser scaup, common merganser, bald eagle, great blue heron, common loon, peregrine falcon and American robin), the breeding season range size was selected as the default (smallest range size).
 - o For resident birds (red-tailed hawk, ruffed grouse and spruce grouse), the foraging range is inclusive of the breeding and non-breeding seasons.
- For the purposes of risk assessment, feeding guilds assigned to ROCs in the receptor-specific tables are based on the majority of the diet. For example, although the muskrat diet is 20% carnivorous, the muskrat was classified as a herbivore due to an 80% herbivorous diet. Depending upon the potential for contamination of different food items and the nature of assumptions in the ERA, the practitioner may choose to focus on the majority dietary component.

3. REFERENCES

- Allard, P., A. Fairbrother, B. Hope, R.N. Hull, M.S. Johnson, L. Kapustka, G. Mann, B. McDonald, B.E. Sample. 2010. Recommendations for the development and application of wildlife toxicity reference values. Integrated Environmental Assessment and Management. 6:28-37.
- Environment Canada (EC). 2012. FCSAP Ecological Risk Assessment Guidance. Report prepared for Environment Canada, 31 March 2012.
- Beyer, W.N., E.E. Connor and S. Gerould. 1994. Estimates of soil ingestion by wildlife. The *Journal of Wildlife Management* 58(2):375-382.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- CCME. 1996. A Framework for Ecological Risk Assessment: General Guidance. Canadian Council of Ministers of the Environment. Winnipeg.
- CCME. 1997. A Framework for Ecological Risk Assessment: Technical Appendices. Canadian Council of Ministers of the Environment. Winnipeg.
- CEAEQ (Centre D'expertise En Analyse Environnementale Du Québec). 2006a. Paramètres d'exposition chez les mammifères Ministère du Développement durable, de l'Environnement et des Parcs du Québec.
- CEAEQ (Centre D'expertise En Analyse Environnementale Du Québec). 2006b. Paramètres d'exposition chez les oiseaux. Ministère du Développement durable, de l'Environnement et des Parcs du Québec.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128.
- Nico, P.S., M.V. Ruby, Y.W. Lowney and S.E Holm. 2006. Chemical speciation and bioaccessibility of arsenic and chromium in chromated copper arsenate-treated wood and soils. *Environmental Science and Technology*, 40(1): 402-408.
- Office of Solid Waste and Emergency Response (OSWER). 2005. Guidance for Developing Ecological Soil Screening Levels, Attachment 4-3 OSWER Directive 9285.7-55. U.S. Environmental Protection Agency, Office of Solid Waste and Emergency Response, Washington, DC
- Ruby, M.V., A.Davis, R. Schoof, S. Eberle and C.M. Sellstone.1996. Estimation of lead and arsenic bioavailability using a physiologically based extraction test. *Environmental Science and Technology*, 30(2): 422-430.
- Ruby, M.V., R. Schoof, W. Brattin, M. Goldade, G. Post, M. Harnois, D.E. Mosby, S.W. Casteel, W. Berti, M. Carpenter, D. Edwards, D. Cragin and W. Chappell. 1999. Advances in evaluating the oral bioavailability of inorganics in soil for use in human health risk assessment. Environmental Science and Technology, 33(21): 3697-3705.
- Sample, B.E., D.M. Opresko, and G.W Suter II. 1996. Toxicological Benchmarks for Wildlife: 1996 Revision. Oak Ridge National Laboratory, Oak Ridge.

- Science Advisory Board (SAB) for Contaminated Sites in British Columbia. 2008. Guidance for Detailed Ecological Risk Assessments (DERA) in British Columbia. British Columbia, Canada.
- Sparling, D.W., G. Linder, C.A. Bishop, S.K. Krest. 2010. Ecotoxicology of Amphibians and Reptiles, Second Edition. ISBN 978-1-4200-6416-2. Society of Environmental Toxicology and Chemistry (SETAC).
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

APPENDIX A

RECEPTOR-SPECIFIC TABLES

Muskrat (Ondatra zibethicus)	A-1
Moose (Alces alces)	A-4
Northern River Otter (Lontra canadensis)	A-7
American Mink (<i>Mustela vision</i>)	A-10
Meadow Vole (Microtus pennsylvanicus)	A-13
White-tailed Deer (Odocoileus virginianus)	A-15
Snowshoe Hare (Lepus americanus)	A-18
Common (Masked) Shrew (Sorex cinereus)	A-21
Ermine or Short-tailed Weasel (Mustela erminea)	A-23
Red Fox (Vulpes vulpes)	A-25
Black Bear (Ursus americanus)	A-27
Deer Mouse (Peromyscus maniculatus)	A-30
Mallard (Anas platyrhynchos)	A-32
Spotted Sandpiper (Actitus macularia)	A-35
Lesser Scaup (Aythya affinis)	A-38
Common Merganser (Mergus merganser)	A-41
Bald Eagle (Haliaeetus leucocephalus)	A-44
Great Blue Heron (Ardea herodias)	A-47
Common Loon (Gavia immer)	A-50
Ruffed Grouse (Bonasa umbellus)	A-53
Spruce Grouse (Dendragapus canadensis)	A-56
Barn Swallow (Hirundo rustica)	A-59
Red-tailed Hawk (Buteo jamaicensis)	A-61
Peregrine Falcon (Falco peregrinus)	A-64
American Robin (Turdus migratorius)	A-68
Wood Frog (Rana sylvatica)	A-71
Common Gartersnake (Thamnophis sirtalis)	A-73

Muskrat (*Ondatra zibethicus*)

Characteristic	Determination	Notes (With References)
General Habitat Type	Wetland	(Nagorsen 2005; Banfield 1974)
Specific Habitat Type		nd that is deep enough not to freeze solid in winter and bulrushes and/or sedges (Nagorsen 2005).
Foraging Range Size (All Seasons)	Area: 300 m ² (302 - 7900 m ²) Linear: 250 m (250 - 400 m)	Area: Range is variable depending on food availability (Banfield 1974; Proulx and Gilbert 1983; Caley 1987). <u>Linear</u> : (Brooks 1985)
Body Weight	$\underline{\text{Avg}}$: 1.0 kg	(Banfield 1974; Nagorsen 2005)
Foraging Guild	Herbivore	(Banfield 1974; Eder and Pattie 2001; Nagorsen 2005)
Foraging Behavior	Grazing; rarely hunting	(Nagorsen 2005; Banfield 1974)
Diet Proportions		
Aquatic Vegetation	80%	Stems, leaves, shoots, roots and tubers of aquatic
Aquatic Invertebrates	15%	plants such as cattails, bulrushes, pondweeds, water
Other (Fish, Small Mammals/Birds, Salamanders, Earthworms)	5%	lilies and horsetails are the most common food species for muskrats in North America (Errington 1941; Takos 1947; Lacki et al. 1990). In some areas where cattail is plentiful, it may represent up to 80% of muskrat diet (Proulx and Gilbert 1983). In some habitat types, muskrat will eat animal matter
		including turtles, mussels, clams, crayfish, snails, fish and small birds (Convey et al. 1989; Neves and Odom 1989; Eder and Pattie 2001; Nagorsen 2005).
Food Ingestion Rate	0.07 kg dry food/kg wet BW/day	(Campbell and MacArthur 1996). 1213
Water Ingestion Rate	0. 10 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all mammals (L/day; (0.099*BW ^{0.90}) divided by BW. The average BW provided in this table was used for these calculations.

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 $^{^{12}}$ This food ingestion rate is based on captured muskrats (both males and females, all age classes) fed a natural diet with body weights ranging from 0.8 to 1 kg.

 $^{^{13}}$ The species-specific food ingestion rates for the muskrat (0.34 and 0.26 g/g day) provided by the US EPA (1993) is based on captive animals fed corn and greens, with no mention of age or gender.

Characteristic	Determination	Notes (With References)
Incidental Soil or		
Sediment Ingestion	None identified	Refer to Section 2.1.8.
Rate		

Muskrat References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Brooks, R.P. 1985. Microenvironments and activity patterns of burrow-dwelling muskrats (Ondatra zibethicus) in rivers. *Acta Zoologica Fennica* 173: 47-49.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Caley, M.J. 1987. Dispersal and inbreeding avoidance in muskrats. *Animal Behaviour* 35: 1225-1233.
- Campbell, K.L. and R.A. MacArthur. 1996. Seasonal Changes in Gut Mass, Forage Digestibility, and Nutrient Selection of Wild Muskrats (*Ondatra zibethicus*). *Physiological Zoology* 69 (5):1215-1231.
- Convey, L.E., J.M. Hanson and W.C. MacKay. 1989. Size-selective predation on unionid clams by muskrats. *The Journal of Wildlife Management* 53(3): 654-657.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Errington, P.L. 1941. Versatility in feeding and population maintenance of the muskrat. *The Journal of Wildlife Management* 5(1): 68-69.
- Lacki, M.J., W.T. Penston, K.B. Adams, F.D. Vogt and J.C. Houppert. 1990. Summer foraging patterns and diet selection of muskrats inhabiting a fen wetland. *Canadian Journal of Zoology* 68(6): 1163-1167.
- Nagorsen, D.W. 2005. Rodents & Lagomorphs of British Columbia. Royal BC Museum, Victoria, Canada.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128.
- Neves, R.J. and M.C. Odom. 1989. Muskrat predation on endangered freshwater mussels in Virginia. *The Journal of Wildlife Management* 53(4): 934-941.
- Proulx, G. and F.F. Gilbert. 1983. The ecology of the muskrat, *Ondatra zibethicus*, at Luther Marsh, Ontario. Canadian Field-Naturalist 97(4):377-390.
- Takos, M.J. 1947. A semi-quantitative study of muskrat food habits. *The Journal of Wildlife Management* 11(4): 331-339.

USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

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Moose (Alces alces)

Characteristic	Determination	Notes (With References)
General Habitat Type	Forest, Wetland	(Banfield 1974)
		Boreal, northern and subalpine forests. Common in
Chasifia Habitat Tyma	Disturbed and	recently disturbed habitats where there is a mix of
Specific Habitat Type	open forests	young and old forest stands as well as diverse
		browse species (Shackleton 1999).
		Habitat sizes for the moose vary considerably with
Foraging Range Size	Area: 4.6 km²	geographic location and method of calculation
(All Seasons)	$(4.6 - 262 \text{ km}^2)$	(Doerr 1983; Cederlund and Okarma 1988; Leptich
(All Seasons)	(4.0 - 202 Kill)	and Gilbert 1989; Cederlund and Sand 1994;
		Stenhouse et al. 1994; Demarchi 2003).
	<u>Avg:</u> 400 kg	
Body Weight	Male: 453 kg	(Banfield 1974)
	Female: 350 kg	
Foraging Guild	Herbivore	(Banfield 1974; Eder and Pattie 2001)
Foraging Behavior	Browsers	(Banfield 1974; Eder and Pattie 2001)
Diet Proportions		
Shrubs/Trees	80%	Moose are generalist herbivores that feed on
Aquatic Plants	20%	herbaceous plants (including aquatic), leaves and
		new growth of shrubs and trees in summer and twigs
		of woody vegetation during winter (Banfield 1974).
		Up to 80% of the moose diet is woody matter
		consisting of shrubs, twigs and branches (Eder and
		Pattie 2001). The remaining 20% consists of aquatic
		vegetation that is eaten because of its high sodium
		content (Fraser et al. 1980). In winter, the most
		commonly consumed plant species is willow, but
		twigs of trembling aspen, saskatoon, birch and red-
		osier dogwood are also eaten in great quantities
		(Weixelman et al. 1998; Shackleton 1999). Conifers
		such as spruce and pine will not sustain moose,
		although some types of fir and yew are eaten readily
		(Cushwa and Coady 1976; Allen et al. 1987). In
		summer, moose are attracted to weedy lakes,
		marshes and sluggish streams where they feed on
		aquatic vegetation such as horsetail, bur-reed and
		pondweed (Nietfeld et al. 1985; MacCracken and

Characteristic	Determination	Notes (With References)
Food Ingestion Rate	0.02kg dry food/kg wet BW/day	(Renecker and Hudson 1985) ¹⁴
Water Ingestion Rate	0.05 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all mammals (L/day; (0.099*BW ^{0.90}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	<2% of dry food ingestion rate	(Beyer et al.1994)

Moose References

- Allen, A.W., P.A. Jordan and J.W. Terrell. 1987. Habitat Suitability Index Models: Moose, Lake Superior Region. Biological Report 82 (10.155). Washington, DC: U.S. Department of the Interior, Fish and Wildlife Service. 47 pp.
- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Beyer, W.N., E.E. Connor and S. Gerould. 1994. Estimates of soil ingestion by wildlife. *The Journal of Wildlife Management* 58(2):375-382.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Cederlund, G.N. and H. Okarma. 1988. Home range and habitat use of adult female moose. The *Journal of Wildlife Management* 52(2): 336-343.
- Cederlund, G.N. and H. Sand. 1994. Home-range size in relation to age and sex in moose. *Journal of Mammalogy* 75(4): 1005-1012.
- Cushwa, C.T. and J. Coady. 1976. Food habits of moose (*Alces alces*) in Alaska: A preliminary study using rumen contents analysis. *Canadian Field-Naturalist* 90:11-16.
- Demarchi, M.W. 2003. Migratory patterns and home range size of moose in the Central Nass Valley, British Columbia. Northwestern Naturalist 84:135-141.

 14 Based on a dry matter intake (for two free-ranging female moose) of 83 g/kg BW $^{0.75}$ (from a range of 38 to 129 g/kg BW $^{0.75}$) and an average body weight of 320 kg.

- Doerr, J.G. 1983. Home range size, movements and habitat use in two moose, *Alces alces*, populations in southeastern Alaska. *Canadian Field-Naturalist* 97(1): 79-88.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Fraser, D., D. Arther, J.K. Morton and B.K. Thompson. 1980. Aquatic feeding by moose *Alces alces* in a Canadian lake. *Ecography* 3(3): 218-223.
- Leptich, D.J. and J.R. Gilbert. 1989. Summer home range and habitat use by moose in northern Maine. *The Journal of Wildlife Management* 53(4): 880-885.
- MacCracken, J.G. and L.A Viereck. 1990. Browse regrowth and use by moose after fire in interior Alaska. *Northwest Science* 64(1): 11-18.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128.
- Nietfeld, M., J. Wilk, K. Woolnough and B Hoskin. 1985. Wildlife Habitat Requirement Summaries for Selected Wildlife Species in Alberta. Wildlife Resource Inventory Unit, Alberta Energy and Natural Resources. ENR Technical Report Number T/73.
- Renecker, L.A. and R.J. Hudson. 1985. Estimation of Dry Matter Intake of Free-Ranging Moose. The Journal of Wildlife Management 49(3): 785-792
- Shackleton, D. 1999. Hoofed Mammals of British Columbia. Volume 3, The Mammals of British Columbia. Royal BC Museum Handbook. UBC Press, Vancouver, Canada.
- Stenhouse, G.B., P.B. Latour, L. Kutny, N. MacLean and G. Glover. 1995. Productivity, survival, and movements of female moose in a low-density population Northwest Territories, *Canada. Arctic* 48(1):57-62.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Weixelman, D.A., R.T. Bowyer and V. Van Ballenberghe. 1998. Diet selection by Alaskan moose during winter: Effects of fire and forest succession. *Alces* 34(1): 213-238.

Northern River Otter (Lontra canadensis)

Characteristic	Determination	Notes (With References)
General Habitat Type	Shoreline,	(Eder and Pattie 2001; Banfield 1974)
General Habitat Type	Waterways	(Eder and Fattle 2001, Banneld 1974)
Specific Habitat Type	In and along the wooded shores of lakes and coastlines (Eder and Pattie	
Specific Habitat Type	2001; Banfield 19	74).
		Generally females have a smaller linear foraging
	Linear: 3.5 km	range, compared to the larger linear foraging range
Foraging Range Size	(3.5 - 30 km)	of males. Research suggests polluted areas cause
(All Seasons)	Area: 9 km ²	foraging range size to increase (Toweill and Tabor
	$(9 - 231 \text{ km}^2)$	1982; Baker 1983; Shirley et al. 1988; Reid 1994;
		Bowyer et al. 1995; Hatler et al. 2008).
	<u>Avg:</u> 7.5 kg	
Body Weight	Male: 7.7 kg	(Lariviere and Walton 1998)
	Female: 7.3 kg	
Foraging Guild	Carnivore	(Eder and Pattie 2001; Banfield 1974)
Foraging Behavior	Hunting	(Eder and Pattie 2001; Banfield 1974)
Diet Proportions		
Fish	80%	River otters are opportunistic foragers, feeding on
Aquatic Invertebrates	15%	aquatic animals, particularly fish, frogs, crayfish,
Small Mammals/Birds	5%	turtles, insects, and sometimes birds and small
		mammals (Knudson and Hale 1968; Toweill and
		Tabor 1982; Hatler et al. 2008). In particular, otters
		target slow-moving, mid-sized fish species (Guertin
		et al. 2010). In the Great Lakes Region, diet studies
		determined that fish occurred in 80-100% of
		stomachs analyzed and that crayfish, an important
		secondary food group, were present in 30-60% of
	<u>, </u>	stomachs analyzed (Knudson and Hale 1968).
	0.03 kg dry	15
Food Ingestion Rate	food/kg	(Davis et al. 1992) ¹⁵ .
	BW/day	

¹⁵Ingestion rate is based on male river otters (taken from an otter farm). Otters were fed a formulated feed consisting of fish, poultry, liver, eggs and fortified mink cereal.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.08 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all mammals (L/day; (0.099*BW ^{0.90}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Northern River Otter References

- Baker, R.H. 1983. Michigan mammals. Michigan State University Press. 642 pp.
- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Bowyer, R.T., J.W. Testa and J.B. Faro. 1995. Habitat selection and home ranges of river otters in a marine environment: Effects of the Exxon Valdez oil spill. *Journal of Mammalogy* 76(1): 1-11.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Davis, H.G., R.J. Aulerich, S.J. Bursian, J.G. Sikarskie and J.N. Stuht. 1992. Feed consumption and food transit time in northern river otters (*Lutra canadensis*). *Journal of Zoo and Wildlife Medicine* 23(2): 241-244.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Guertin, D.A., A.S. Harestad and J.E. Elliott. 2010. Summer feeding habits of river otters inhabiting a contaminated coastal marine environment. *Northwest Science* 84(1): 1-8.
- Hatler, D.F., D.W. Nagorsen and A.M. Beal. 2008. Carnivores of British Columbia. Volume 5: The Mammals of British Columbia. Royal BC Museum Handbook, Victoria, Canada.
- Knudson, G.J. and J.B. Hale. 1968. Food habits of otters in the Great Lakes region. *The Journal of Wildlife Management* 32(1): 89-93.
- Larivière, S. and L.R. Walton. 1998. Lontra canadensis. Mammalian Species 587: 1-8.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128.
- Reid, D.G., T.E. Code, A.C.H. Reid and S.M. Herrero. 1994. Spacing, movements, and habitat selection of the river otter in boreal Alberta. *Canadian Journal of Zoology* 72: 1314-1324.
- Shirley, M.G., R.G. Linscombe, N.W. Kinler, R.M. Knaus and V.L. Wright. 1988. Population estimates of river otters in a Louisiana coastal marshland. *The Journal of Wildlife Management* 52:512-515.

- Toweill, D. E. and J. E. Tabor. 1982. River Otter (Lutra canadensis). Pages 688-703 in Chapman, J.A. and G. A. Feldhamer (eds). Wild mammals of North America: biology, management, and economics. Johns Hopkins Univ. Press, Baltimore.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

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American Mink (*Mustela vison*)

Characteristic	Determination	Notes (With References)	
General Habitat Type	Shoreline, Waterways	(Eder and Pattie 2001; Banfield 1974)	
Specific Habitat Type	Stream banks, lakeshores, forest edges, large swamps and tidal flats (Banfield 1974). Coastline and wet zones in brush lands, mountains and grasslands (Eder and Pattie 2001).		
Foraging Range Size (All Seasons)	Linear: 0.4 km (0.4 - 11 km) Area: 0.06 km² (0.06 - 16.3 km ²)	Smallest foraging range size determined from adult females and largest range size determined from adult males (Ritcey and Edwards 1956; Eagle and Whitman 1987; Niemimaa 1995; Stevens et al. 1997; Hatler et al. 2008).	
Body Weight	Avg: 820 g Male: 570 g Female: 1060 g	(McCabe 1949)	
Foraging Guild	Carnivore	(Eder and Pattie 2001; Banfield 1974)	
Foraging Behavior	Hunting	(Eder and Pattie 2001; Banfield 1974)	
Diet Proportions			
Fish	30%	The American mink diet consists of invertebrates,	
Crustaceans	25%	amphibians, fish, waterfowl and their eggs, mice,	
Small Mammals/Birds	25%	voles, rabbits, snakes and aquatic invertebrates	
Amphibians	10%	(Chapman and Feldhammer 1982; Maser 1998; Eder	
Insects	10%	and Pattie 2001; Hatler et al. 2008). On Vancouver	
Insects	1070	Island, mink forage primarily in the intertidal zone where crustaceans and fish are preferred, and mammals and insects are supplemental (Hatler 1976). Scat analysis results show seasonal diet differences with increased presence of crabs from April to July (Hatler 1976). The incidence of crabs, fish, and birds and other foods found in fecal analysis were respectively 85-90%, 60-70% and less then 20% (Hatler 1976). A recent study by Kiseleva (2009) in Russia investigated stomach contents of 345 American mink from brook, small river and large river habitats. On average, diet composition consisted of 11 to 24% small mammals, 2 to 30% fish, 19 to 26% insects, 2 to 11% amphibians and	

Characteristic	Determination	Notes (With References)
		reptiles, and minor amounts of bird and mollusk
		remains. Results from Racey and Euler (1993) were
		also used to determine diet proportions.
Food Ingestion Rate	0.14 kg wet	This rate is based on the average of two ingestion
	food/kg wet	rates (0.12 and 0.16 g/g/ day) for farm raised adults,
	BW/day	both sexes as reported in the US EPA (1993) ¹⁶ .
Water Ingestion Rate	0.03 L/kg wet BW/day	This rate is based on an adult farm raised female (in US EPA 1993) in g/g day (assumes water density of 1 g/ml).
Incidental Soil or		
Sediment Ingestion	None identified	Refer to Section 2.1.8.
Rate		

American Mink References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Bleavins, M.R. and R.J. Aulerich. 1981. Feed consumption and food passage time in mink (*Mustela vison*) and European ferrets (*Mustela putorius furo*). *Laboratory Animal Science* 31: 268-269.
- Chapman, J.A. and G.A. Feldhammer. 1982. Wild Mammals of North America: Biology, Management, and Economics. John Hopkins University Press, Baltimore and London. 1147 pp\
- Eagle, T.C. and J.S. Withman. 1987. "Mink" in Wild Furbearer Management and Conservation in North America. M. Novak, J.A. Baker, M.E. Obbard and B. Malloch (eds.). University of Pittsburgh Press, Pittsburgh.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Hatler, D. 1976. The Coastal Mink. Master of Science Thesis, University of British Columbia. Vancouver, Canada.
- Hatler, D.F., D.W. Nagorsen and A.M. Beal. 2008. Carnivores of British Columbia. Volume 5: The Mammals of British Columbia. Royal BC Museum Handbook, Victoria, Canada.
- Kiseleva, N.V. 2009. The peculiarities of feeding of the American mink (*Nerovision vision*) in the Southern Urals. *Biology Bulletin* 36(4): 403-406.
- Maser, C. 1998. Mammals of the Pacific Northwest: From the Coast to the High Cascades. Oregon State University Press, Corvallis, Oregon

¹⁶ The US EPA values used are based on Bleavins and Aulerich (1981).

- McCabe, R. 1949. Notes on live-trapping mink. Journal of Mammalogy 30(4): 416-423.
- Niemimaa, J. 1995. Activity patterns and home ranges of the American mink *Mustela vison* in the Finnish outer archipelago. *Annales Zoologici Fennici* 32: 117-121.
- Racey, G.D. and D.L. Euler. 1983. Changes in mink habitat and food selection as influenced by cottage development in central Ontario. *Journal of Applied Ecology* 20(2): 387-402.
- Ritcey, R.W. and R.Y. Edwards. 1956. Live trapping mink in British Columbia. *Journal of Mammalogy* 37(1): 114-116.
- Stevens, R.T., T.L. Ashwood and J.M. Sleeman. 1997. Fall early winter home ranges, movements, and den use of male mink, *Mustela vison* in eastern Tennessee. *Canadian Field-Naturalist* 111(2): 312-314.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Meadow Vole (Microtus pennsylvanicus)

Characteristic	Determination	Notes (With References)
General Habitat Type	Grassy and open forest habitat	(Banfield 1974; Eder and Pattie 2001)
Specific Habitat Type	Grasslands, pastures, marshy areas, open woodlands, taiga and mountain meadow (Eder and Pattie 2001). Often associated with wet areas such as lakes, streams, dykes, sloughs and ditches (Nagorsen 2005).	
Foraging Range Size (All Seasons)	<u>Area</u> : 69 m² (69 – 3,480 m ²)	Females generally have a smaller foraging range than males and range is variable based on season, population density and research method (Madison 1980; Sullivan and Hogue 1987; Pugh and Ostfeld 1998).
Body Weight	<u>Avg</u> : 34.9 g	(Nagorsen 2005)
Foraging Guild	Herbivore	(Nagorsen 2005)
Foraging Behavior	Grazing	(Nagorsen 2005)
Diet Proportions		
Berries/Seeds	60%	Primary food sources are the stems, leaves and seed
Grasses	30%	heads of grasses and sedges, bark and some insects
Mushrooms	10%	(Thompson 1965; Maser and Strom 1970; Lindroth
		and Batzli 1984). For a winter food supply, meadow voles make caches of leaves, roots, rhizomes, bulbs, bark and corms in runways under the snow (van Zyll de Jong 1983). The inner bark of trees, shrubs and conifer seedlings are also eaten (Bucyanayandi et al. 1990; Nagorsen 2005).
Food Ingestion Rate	0.33 kg wet food/kg wet BW/day	This rate is based on an average of two values reported in US EPA (1993; 0.325 g/g day), with no information on age and gender of the animals. ¹⁷ .
Water Ingestion Rate	0.21 L/kg wet BW/day	This rate is based on a 0.21 g/g day water ingestion rate for adult meadow voles (both sexes) and assumes a water density of 1g/ml (US EPA 1993)
Incidental Soil or Sediment Ingestion Rate	2.4 % of dry food ingestion rate	Based on US EPA 1993 ¹⁸

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¹⁷ Ingestion rate is based on Ogdev et al. 1950.

¹⁸ Species-specific information in US EPA (1993) on soil or sediment ingestion rates are taken from Beyer et al. (1994) which was in press at the time of EPA publication.

Meadow Vole References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Beyer, W.N., E.E. Connor and S. Gerould. 1994. Estimates of soil ingestion by wildlife. *The Journal of Wildlife Management* 58(2):375-382.
- Bucyanayandi, J.-D., J.-M. Bergeron and H. Menard. 1990. Preference of meadow voles (*Microtus pennsylvanicus*) for conifer seedlings: Chemical components and nutritional quality of bark of damaged and undamaged trees. *Journal of Chemical Ecology* 16(8): 2569-2579.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Lindroth, R.L. and G.O. Batzli. 1984. Food habits of the meadow vole (*Microtus pennsylvanicus*) in bluegrass and prairie habitats. *Journal of Mammalogy* 65(4): 600-606.
- Madison, D.M. 1980. Space use and social structure in meadow voles, *Microtus pennsylvanicus*. *Behavioural Ecology and Sociobiology* 7(1): 65-71.
- Maser, C. and R.M. Strom. 1970. A key to the Microtinae of the Pacific Northwest. O.S.U. Book Stores, Inc., Corvallis, OR.
- Nagorsen, D.W. 2005. Rodents & Lagomorphs of British Columbia. Volume 4, The Mammals of British Columbia. Royal BC Museum Handbook, Royal BC Museum, Victoria, Canada.
- Ogdev, S.I. 1950. Mammals of the U.S.S.R. and adjacent countries. Translated from Russian by: Israel Program for Scientific Translations (1964), Jerusalem; 626 pp.
- Pugh, S.R. and R.S. Ostfeld. 1998. Effects of prior population density on use of space by meadow voles. *Microtus pennsylvanicus*. *Journal of Mammalogy*. 79: 551-557.
- Sullivan, T.P. and E.J. Hooge. 1987. Influence of orchard floor management on vole and pocket gopher populations and damage in apple orchards. *Journal of the American Society for Horticultural Science* 112: 972-977.
- Thompson, D.Q. 1965. Food preferences of the meadow vole (*Microtus pennsylvanicus*) in relation to habitat affinities. *American Midland Naturalist* 74(1): 76-86.
- van Zyll de Jong, C.G. 1983. Handbook of Canadian Mammals: 1 Marsupials and Insectivores. National Museum of Natural Sciences, National Museum of Canada.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

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White-tailed Deer (Odocoileus virginianus)

Characteristic	Determination	Notes (With References)	
General Habitat Type	Forest	(Eder and Pattie 2001)	
Specific Hebitet Type	Woodlands, meadows, valleys, stream courses and rolling country (Eder		
Specific Habitat Type	and Pattie 2001).		
Foraging Range Size (Minimum is for Winter Season)	<u>Area</u> : 30 ha (30 - 2435 ha)	Adult females have smaller foraging ranges relative to adult males, and average foraging range in winter is generally significantly smaller than in summer (Tierson et al. 1985; Lesage et al. 2000). Average home (foraging) ranges have varied from 129 ha (winter) to 2435 ha (summer) in Quebec (Lesage et al. 2000), 135 ha (winter) and 225 ha (summer) in the Adirondacks, New York (Tierson et al. 1985), 370 ha (winter) to 277 ha (summer) in New Brunswick (Drolet 1976), 178 ha (winter) in Wisconsin (Larson et al. 1978), and 30 ha (winter) to 75 ha (summer) in Minnesota (Mooty et al. 1987).	
Body Weight	Avg: 75 kg Male: 91 kg Female: 60 kg	(Banfield 1977)	
Foraging Guild	Herbivore	(Banfield 1974)	
Foraging Behavior	Browsers	(Banfield 1974)	
Diet Proportions			
Trees (Buds/Twigs)	35%	The diet of the white-tailed deer is seasonally	
Herbaceous Plants	20%	variable. In the winter, buds and twigs of shrubs are	
Leaves	15%	important, in the fall, fruit and mushrooms make up	
Fruits	10%	a significant part of the diet, while in the summer,	
Mushrooms	10%	grasses and herbaceous plants are preferred	
Grasses	9%	(Banfield 1974; Skinner and Telfer 1974; Hesselton	
Lichens	1%	and Hesselton 1982; Crawford 1982). In eastern	
		Canada, white cedar is a major component of the diet, with red maple, red-osier dogwood, mountainash, sumac, trembling aspen, ground hemlock, balsam fir and basswood trees also preferred (Banfield 1974). Plants eaten include asters, goldenrod, hawkweed, mayflower, ferns, leaves, water plants and mushrooms (Banfield 1974; Skinner and Telfer 1974).	

Characteristic	Determination	Notes (With References)
	0.03 kg dry	This is based on the allometric equation (Nagy 1987 in US EPA 1993) for total dry food intake for
Food Ingestion Rate	food/kg wet BW/day	herbivorous mammals (g/day;(0.577*BW ^{0.727}) divided by the BW. The average BW provided in this table was used for these calculations.
Water Ingestion Rate	0.06 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all mammals (L/day; (0.099*BW ^{0.90}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	<2.0% of dry food ingestion rate	Beyer et al. 1994

White-tailed Deer References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Banfield, A.W.F. 1977. Les mammifères du Canada. Deuxième édition. Les Presses de l'Université Laval, Québec, 406 p. Cited in Centre D'expertise En Analyse Environnementale Du Québec. 2006. Paramètres d'exposition chez les mammifères Cerf de Virginie. Fiche descriptive. Ministère du Développement durable, de l'Environnement et des Parcs du Québec, 27 p.
- Beyer, W.N., E.E. Connor and S. Gerould. 1994. Estimates of soil ingestion by wildlife. *The Journal of Wildlife Management* 58(2):375-382.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Crawford, H.S. 1982. Seasonal food selection and digestibility by tame white-tailed deer in central Maine. *The Journal of Wildlife Management* 46: 974-982.
- Drolet, C.A. 1976. Distribution and movements of White-tailed Deer in southern New Brunswick in relation to environmental factors. Canadian Field-Naturalist 90: 123-136
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Hesselton, W.T. and R.M. Hesselton. 1982. White-tailed Deer. *Odocoileus virginianus*. Cited in Wild mammals of North America: Biology, management, and economics. J.A. Chapman and G.A. Feldhamer (eds), The Johns Hopkins University Press, Baltimore, p. 878-901.
- Larson, T.J., O.J. Rongstad and F.W. Terbilcox. 1978. Movement and habitat use of White-tailed Deer in southcentral Wisconsin. Journal of Wildlife Management 42(1): 113-117.

- Lesage, L., M. Crête, J. Huot, A. Dumont and J.-P. Ouellet. 2000. Seasonal home range size and philopatry in two northern white-tailed deer populations. *Canadian Journal of Zoology* 78(11):1930-1940. Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128.
- Mooty, J.J., P.D. Karns and T.K. Fuller. 1987. Habitat use and seasonal range size of White-tailed Deer in northcentral Minnesota. Journal of Wildlife Management 51(3): 644-648.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128
- Skinner, W.R. and E.S. Telfer. 1974. Spring, summer, and fall foods of deer in New Brunswick. *The Journal of Wildlife Management* 38: 210-214.
- Tierson, W.C., G.F. Mattfeld, R.W. Sage Jr. and D.F. Behrend. 1985. Seasonal movements and home ranges of White-tailed Deer in the Adirondacks. Journal of Wildlife Management 49(3): 760-769.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Snowshoe Hare (Lepus americanus)

Characteristic	Determination	Notes (With References)
General Habitat Type	Forest	(Eder and Pattie 2001)
Specific Habitat Type	Habitat types of particularly high suitability include conifer-dominated habitats, deciduous riparian forests, birch-willow scrub, subalpine parkland and shrub fens/carrs (Dolbeer and Clark 1975; McTaggart-Cowan and Guiguet 1978; Buehler and Keith 1982; Stevens and Lofts 1988).	
Foraging Range Size	<u>Area</u> : 1.6 ha	Foraging range size varies with season and
(All Seasons)	(1.6 -10.2 ha)	population cycle (Nagorsen 2005; O'Farrell 1965).
Body Weight	Avg: 1.3 kg (0. 9 – 1.9 kg)	(Nagorsen 2005)
Foraging Guild	Herbivore	(Nagorsen 2005; Banfield 1974)
Foraging Behavior	Grazing	(Nagorsen 2005)
Diet Proportions		
Shrubs	60%	Summer diet consists of grasses, sedges and forbs,
Grasses	30%	while winter diet consists of stems and branches of
Berries	10%	woody plants, especially the more digestible
		terminal branches (Wolff 1978; Nagorsen 2005). Favorite winter foods in western Canada and Alaska include spruce needles, and the bark and twigs of trembling aspen, paper birch, willow, alder, spruce, blueberries and soopalallie (or buffaloberry) (Shepherdia canadensis) (Banfield 1974; Wolff 1978; Smith et al. 1988). In spring, blueberries, lowbush cranberry (Vaccinium vitis-idaea), fireweed (Epilobium angustifolium) and horsetail made up 47% of hare diet in Alaska (Wolff 1978). Summer diet in Alaska was primarily the leaves of birch, willow, rose and other deciduous shrubs (Wolff 1978). In Ontario, pine were most heavily browsed by snowshoe hare, but many other species were also eaten including aspen, alder, hazelnut, elderberry, willow and saskatoon (de Vos 1964). Snowshoe hare occasionally exhibits cannibalistic tendencies in winter (Banfield 1974).

Characteristic	Determination	Notes (With References)
Food Ingestion Rate	0.06 kg dry food/kg wet BW/day	(Walski and Mautz 1977) ¹⁹
Water Ingestion Rate	0.10 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all mammals (L/day; (0.099*BW ^{0.90}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	6.3% of dry dietary ingestion rate	The jackrabbit has been estimated to have 6.3 % soil in diet (dry weight) (US EPA 1993).

Snowshoe Hare References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Buehler, D.A. and L.B. Keith. 1982. Snowshoe hare distribution and habitat use in Wisconsin. *Canadian Field-Naturalist* 96(1):19-29.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- de Vos, A. 1964. Food utilization of snowshoe hares on Manitoulin Island, Ontario. *Journal of Forestry* 62(4): 238-244.
- Dolbeer, R.A. and W.R. Clark. 1975. Population ecology of snowshoe hares in the central Rocky Mountains. *The Journal of Wildlife Management* 39(3): 535-549.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada
- McTaggart-Cowan, I. and C.J. Guiguet. 1978. The Mammals of British Columbia. Handbook No. 11, B.C. Provincial Museum, Victoria, Canada.
- Nagorsen, D.W. 2005. Rodents & Lagomorphs of British Columbia. Volume 4, The Mammals of British Columbia. Royal BC Museum Handbook, Royal BC Museum, Victoria, Canada.
- O'Farrell, T.P., 1965. Home range ecology of snowshoe hares in interior Alaska. *Journal of Mammalogy*. 46:406-418.

¹⁹ Ingestion rate is based on adults, both sexes with a mixed diet of browse and commercial rabbit pellets.

- Smith, J.N.M., C.J. Krebs, A.R.E. Sinclair and R. Boonstra. 1988. Population biology of snowshoe hares. II. Interactions with winter food plants. *Journal of Animal Ecology* 57(1): 269-286.
- Stevens, V. and S. Lofts. 1988. Wildlife Habitat Handbook for the Southern Interior Ecoprovince. Wildlife Habitat Research WHR-28, Wildlife Report No. R-15. Ministry of Environment (Wildlife Branch) and Ministry of Forests (Research Branch), Victoria, B.C.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Walski, T.W. and W.W. Mautz. 1977. Nutritional Evaluation of Three Winter Browse Species of Snowshoe Hares. *The Journal of Wildlife Management* (41):144-147
- Wolff, J.O. 1978. Food habits of snowshoe hares in interior Alaska. *The Journal of Wildlife Management* 42(1): 148-153.

Common (Masked) Shrew (Sorex cinereus)

Characteristic	Determination	Notes (With References)
General Habitat Type	Damp Forest	(Nagorsen 1996; Eder and Pattie 2001)
		oniferous or deciduous moist, cool forests as well as
Specific Habitat Type	_	and brushy riparian areas (Nagorsen 1996; Eder and
	Pattie 2001) with a	abundant rocks, stumps, and ferns (Merritt 1987).
Foraging Range Size	Area: 0.6 ha	Average foraging range size (Nagorsen 1996)
(All Seasons)		The tage to taging tange one (tage to the tage)
Body Weight	<u>Avg</u> : 4.1 g (2.5g - 8.0g)	(Nagorsen 1996)
Foraging Guild	Insectivore	(Nagorsen 1996; Eder and Pattie 2001)
Foraging Behavior	Hunting	(Nagorsen 1996)
Diet Proportions		
Insects and Larvae	65%	During the summer, the common shrew feeds
Slugs, Snails and	30%	primarily on insects including larvae of moths and
Earthworms	30%	beetles, caterpillars, centipedes, millipedes, spiders,
Other (Amphibians,		slugs, snails, earthworms, and plant material
Small Mammals,	5%	(Bellocq et al. 1994; McCay and Storm 1997; Lee
Vegetation)		2001). Up to 50% of the diet can consist of ants and
,		insect larvae (Lee 2001). Common shrews will also
		consume mice and amphibians, and occasionally
		carrion (Nagorsen 1996; Eder and Pattie 2001). In
		winter, the common shrew's diet is mostly insect
		eggs and pupae (Merritt 1987). Diet proportions
		were also determined using Bellocq et al. (1992) and Hamilton (1930).
		This is based on the allometric equation (Nagy 1987
	0.34 kg dry	in US EPA 1993) for total dry food intake for
Food Ingestion Rate	food/kg wet	rodents (g/day;(0.621*BW ^{0.564}) divided by the BW.
1 ood ingestion itute	BW/day	The average BW provided in this table was used for
	2 Wally	these calculations.
		This is based on the allometric equation (Calder and
		Braun 1983 in US EPA 1993) for total water intake
Water Ingestion Rate	0.17 L/kg wet	for all mammals (L/day; (0.099*BW ^{0.90}) divided by
8	BW/day	BW. The average BW provided in this table was
		used for these calculations.
Incidental Soil or	None identified	Refer to Section 2.1.8.
Sediment Ingestion		
Rate		

Common (Masked) Shrew References

- Bellocq, M.I., J.F. Bendell and B.L. Cadogan. 1992. Effects of the insecticide *Bacillus* thuringiensis on *Sorex cinereus* (Masked Shrew) populations, diet and prey selection in a jack pine plantation in northern Ontario. *Canadian Journal of Zoology* 70(3): 505-510.
- Bellocq, M.I., J.F. Bendell and D.G.L. Innes. 1994. Diet of *Sorex cinereus*, the Masked Shrew, in relation to the abundance of Lepidopteran larvae in northern Ontario. *American Midland Naturalist* 132(1): 68-73.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Hamilton, W.J., Jr. 1930. The food of the Soricidae. Journal of Mammalogy 11(1): 26-39.
- Lee, W. 2001. Sorex cinereus. Animal Diversity Web. Available at http://animaldiversity.ummz.umich.edu/site/. Accessed January 2011
- McCay, T.S. and G.L. Storm. 1997. Masked Shrew (*Sorex cinereus*) abundance, diet and prey selection in an irrigated forest. *American Midland Naturalist* 138(2): 268-275.
- Merritt, J.F. 1987. Guide to the Mammals of Pennsylvania. University of Pittsburgh Press. 408 pp.
- Nagorsen, D.W. 1996. Opossums, Shrews and Moles of British Columbia. Volume 2: The Mammals of British Columbia. Royal British Columbia Museum Handbook. UBC Press,
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Ermine or Short-tailed Weasel (Mustela erminea)

Characteristic	Determination	Notes (With References)
General Habitat Type	Forest	(Eder and Pattie 2001)
Specific Habitat Type	Diverse habitat types used. Coniferous or mixed forests and streamside woodlands. Alpine tundra, rock slides and talus slopes during the summer (Eder and Pattie 2001).	
Foraging Range Size (All Seasons)	<u>Area</u> : 1.0 ha (1.0 - 87.4 ha)	On average, females have smaller foraging range sizes then males. Habitat size is seasonally variable and range estimates vary between studies (Simms 1979; Robitaille and Raymond 1995; Hatler et al. 2008).
Body Weight	Avg :89.0 g Male: 118.3 g Female: 59.5g	(Raymond and Bergeron 1986)
Foraging Guild	Carnivore	(Eder and Pattie 2001)
Foraging Behavior	Hunting	(Eder and Pattie 2001)
Diet Proportions		
Small Mammals	50%	Almost entirely carnivorous, the short-tailed weasel
Birds	25%	eats most animals it can catch. Diet includes mice,
Lagomorphs (Rabbits)	20%	voles, shrews, chipmunks, pocket gophers, pikas,
Other (Amphibians, Insects, Bird Eggs)	5%	rabbits, bird eggs and nestlings, amphibians and insects (Northcott 1971; Erlinge 1981; Eder and
		Pattie 2001; Edwards and Forbes 2003). Analysis of short-tailed weasel droppings in New Zealand determined that rats and mice made of 77% of the volume of feces, while lagomorphs (rabbits, hares and pikas) contributed 12%, and birds 3% (Murphy and Bradfield 1992). After a decline in rat populations, birds became the largest contributor to the diet making up 42% of feces volume (Murphy and Bradfield 1992).
Food Ingestion Rate	0.11 kg dry food/kg wet BW/day	This is based on the allometric equation (Nagy 1987 in US EPA 1993) for total dry food intake for all mammals (g/day;(0.235*BW ^{0.822}) divided by BW. The average BW provided in this table was used for these calculations.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.13 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all mammals (L/day; (0.099*BW ^{0.90}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Ermine or Short-tailed Weasel References

- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Edwards, M.A. and G.J. Forbes. 2003. Food habits of ermine, *Mustela erminea*, in a forested landscape. *Canadian Field-Naturalist* 117(2): 245-248.
- Erlinge, S. 1981. Food preference, optimal diet and reproductive output in stoats *Mustela erminea* in Sweden. *Oikos* 36(3): 303-315.
- Hatler, D.F., D.W. Nagorsen and A.M. Beal. 2008. Carnivores of British Columbia. Volume 5: The Mammals of British Columbia. Royal BC Museum Handbook, Victoria, Canada.
- Murphy, E. and P. Bradfield. 1992. Change in diet of stoats following poisoning of rats in a New Zealand forest. *New Zealand Journal of Ecology* 16: 137-140.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada
- Northcott, T.H. 1971. Winter predation of *Mustela erminea* in northern Canada. *Arctic* 24(2): 141-143.
- Raymond, M. and J.M. Bergeron. 1986. Sexual dimorphism and morphometric variations in the ermine, *Mustela erminea cicognanii*. *Canadian Journal of Zoology* 64: 1966-1972.
- Robitaille, J.-F. and M. Raymond. 1995. Spacing patterns of ermine, *Mustela erminea L.*, in a Quebec agrosystem. *Canadian Journal of Zoology*. 73: 1827-1834
- Simms, D.A. 1979. Studies of an ermine population in southern Ontario. *Canadian Journal of Zoology* 57(4): 824-832.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Red Fox (Vulpes vulpes)

Characteristic	Determination	Notes (With References)
General Habitat Type	Open Country	(Eder and Pattie 2001)
	Agricultural areas	, lakeshores, river valleys, natural forest clearings and
Specific Habitat Type	alpine and arctic tundra (Eder and Pattie 2001). Typically avoids dense	
	forest (Banfield 19	974).
		Variable depending on geographic location and
Foraging Range Size	<u>Area</u> : 2.8 km ²	season (Lemieux 1989). Widely different range sizes
(All Seasons)	$\frac{\text{Arca.}}{(2.8 - 34.2 \text{ km}^2)}$	between reports. Juveniles have smaller rangers then
(All Seasons)	(2.6 - 34.2 Kili)	adults, and adults with young have reduced range
		sizes (Jones and Theberge 1982; Banfield 1974).
	<u>Avg</u> : 3.8 kg	(Voigt 1987; Banfield 1977)
Body Weight	Male: 4.1 kg	
	Female: 3.4 kg	
Foraging Guild	Omnivore	(Banfield 1974; Eder and Pattie 2001)
Foraging Behavior	Hunting	(Eder and Pattie 2001)
Diet Proportions		
Small Mammals	40%	The red fox is omnivorous and primarily consumes
Invertebrates	25%	small rodents, rabbits and birds during the winter,
Birds	20%	and fruits, berries, bird eggs, small mammals and
Fruits and Plant	15%	nesting birds in the summer (Jones and Theberge
Material	1370	1982; Sargeant et al. 1984; Eder and Pattie 2001;
		Banfield 1974). Diet studies comparing summer and
		winter feeding habits found a 10% increase (by
		volume) in the presence of fruits in the feces of red
		foxes during the summer season (Fortin 1995). Diet
		proportions were also determined in consultation
	T	with Banfield (1974).
	0.09 kg wet	This rate is based on an average of three ingestion
Food Ingestion Rate	food/kg wet	rates for adults in captivity as reported in US EPA
	BW/day	$(1993)^{20}$.
	0.09 L/kg wet	This is based on the allometric equation (Calder and
Water Ingestion Rate	BW/day	Braun 1983 in US EPA 1993) for total water intake
	Diriday	for all mammals (L/day; (0.099*BW ^{0.90}) divided by
		BW. The average BW provided in this table was
		used for these calculations.

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 $^{^{20}}$ The rates reported in US EPA (1993) are based on Sargeant (1978)

Characteristic	Determination	Notes (With References)
Incidental Soil or	2 90/ of dry food	
Sediment Ingestion	2.8% of dry food ingestion rate	US EPA (1993)
Rate		

Red Fox References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Fortin, C. 1995. Écologie comparée du coyote, du lynx du Canada et du renard roux au Parc national Forillon. Mémoire de maîtrise. Département de biologie, Université Laval, Québec, 199 p. Cited in Centre D'expertise En Analyse Environnementale Du Québec. 2006. Paramètres d'exposition chez les mammifères Renard roux. Fiche descriptive. Ministère du Développement durable, de l'Environnement et des Parcs du Québec, 19 p.
- Jones, D.M. and J.B. Theberge. 1982. Summer home range and habitat utilisation of the red fox (*Vulpes vulpes*) in a tundra habitat, northwest British Columbia. Canadian Journal of Zoology 60(5): 807-812.
- Lemieux, R. 1989. Superficie des domaines vitaux et utilisation de l'habitat des renards roux en milieu agro-forestier dans le sud-est du Québec. Mémoire de maîtrise. Département de biologie, Université Laval, Québec, 56 p. Cited in Centre D'expertise En Analyse Environnementale Du Québec. 2006. Paramètres d'exposition chez les mammifères Renard roux. Fiche descriptive. Ministère du Développement durable, de l'Environnement et des Parcs du Québec, 19 p.
- Sargeant, A.B., S.H. Allen and R.T. Eberhardt. 1984. Red fox predation on breeding ducks in midcontinent North America. *Wildlife Monographs* No. 89: 3-41.
- Sargeant, A.B. 1978. Red fox prey demands and implications to prairie duck production. *Journal of Wildlife Management* 42(3): 520-527.
- Voigt, D.R. 1987. "Red Fox." In Wild Furbearer Management and Conservation in North America. Novak, M., J.A. Baker, M.E. Obbard and B. Malloch (eds), The Ontario Trappers Association, Toronto, pp. 378-392
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Black Bear (*Ursus americanus*)

Characteristic	Determination	Notes (With References)
General Habitat Type	Forest	(Eder and Pattie 2001)
Specific Habitat Type	Deciduous or coni 1974).	ferous forests, swamps, and berry patches (Banfield
Foraging Range Size (All Seasons)	Area: 3 km ² (3 – 1147 km ²⁾	Range is extremely variable and dependent on season, gender and habitat. Females typically have smaller ranges then males. The smallest range estimate was determined from adult males between the end of July and end of October. The largest range was determined from adult males between the end of April and beginning of July (Samson and Huot 1994; Boileau at al.1994; Nilsen et al. 1995; Leblanc and Huot 2000).
Body Weight	Avg: 68 kg Male (June- July): 90 kg Female (June- July): 46 kg	Seasonally variable. During hibernation, black bears lose 20-40% of body weight (Eder and Pattie 2001).
Foraging Guild	Omnivore	(Eder and Pattie, 2001; Banfield, 1974)
Foraging Behavior	Hunting/Grazing	
Diet Proportions		
Berries/Seeds	50%	Black bears are omnivorous and utilize seasonally
Grasses and Leaves	30%	and locally abundant food. The bulk of their diet is
Carrion	10%	vegetation and includes leaves, flowering parts,
Fish	5%	roots, bulbs, berries, nuts, fruits of grasses, forbs,
Other (Insects, Small Mammals)	5%	shrubs and trees (Fish and Wildlife Branch 1980). Black bears will also kill small mammals, fish and young ungulates in addition to eating carrion and insects (Banfield 1974; Fish and Wildlife Branch
		1980; Graber and White 1983; Bull et al. 2001). An analysis of 621 black bear scats in northeastern Oregon found the mean estimated volume of food items to be 35% grasses, 24% insects, 16% fruit, 11% soil and wood, 10% animal remains and 4% leaves and stems (Bull et al. 2001). Of scats collected in July, 98% contained insects including

Characteristic	Determination	Notes (With References)
		primarily ants and some yellowjackets (Vespula
		sp.)(Bull et al. 2001). In spring, black bears forage
		for succulent vegetation in wet meadows, riparian
		inclusions, skunk cabbage swamps, avalanche
		chutes, and burns (McDonald and Fuller 2005;
		Stevens and Lofts 1988). During this period, they
		feed mainly on poplar catkins, spruce needles, newly
		emerging grasses and sedges, insects, ants, tree buds
		and carrion resulting from winter losses (Banfield
		1974; Kolenosky and Strathearn 1987; Raine and
		Kansas 1990).
		This is based on the allometric equation (Nagy 1987
	0.03 kg dry	in US EPA 1993) for total dry food intake for all
Food Ingestion Rate	food/kg wet	mammals (g/day;(0.235*BW ^{0.822}) divided by BW.
	BW/day	The average BW provided in this table was used for
		these calculations.
		This is based on the allometric equation (Calder and
	0.06 L/kg wet	Braun 1983 in US EPA 1993) for total water intake
Water Ingestion Rate		for all mammals (L/day; (0.099*BW ^{0.90}) divided by
	BW/day	BW. The average BW provided in this table was
		used for these calculations.
Incidental Soil or		
Sediment Ingestion	None identified	Refer to Section 2.1.8.
Rate		

Black Bear References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Boileau, F., M Crete and J. Huot. 1994. Food habits of the black bear, *Ursus americanus*, and habitat use in Gaspesie Park, eastern Quebec. *Canadian Field-Naturalist* 108:163-169.
- Bull, E.L., T.R. Torgersen and T.L. Wertz. 2001. The importance of vegetation, insects, and neonate ungulates in black bear diet in northeastern Oregon. *Northwest Science* 75(3): 244-253.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada

- Fish and Wildlife Branch. 1980. Preliminary Black Bear Management Plan for British Columbia. Fish and Wildlife Branch, Ministry of Environment. 16 p.
- Graber, D.M. and M. White. 1983. Black bear food habits in Yosemite National Park. International Conference on Bear Research and Management 5:1-10.
- Kolenosky, G.B. and S.M. Strathearn. 1987. Black Bear. Cited in M. Novak, J.A. Baker, M.E. Obbard and B. Malloch (eds.). Wild Furbearer Management and Conservation in North America. Ontario Ministry of Natural Resources, Queen's Printer for Ontario.
- Leblanc, N. and J. Huot. 2000. Ecologie de l'ours noir (Ursus americanus) au Parc national Forillon. Service de la conservation des ecostystemes, Parcs Canada, Quebec, 115p. Cited in: Centre D'expertise En Analyse Environnementale Du Québec. 2006. Paramètres d'exposition chez les mammifères Ours noir. Fiche descriptive. Ministère du Développement durable, de l'Environnement et des Parcs du Québec, 17 p.
- McDonald, J.E. and T.K. Fuller. 2005. Effects of spring acorn availability on black bear diet, milk composition and cub survival. *Journal of Mammalogy* 86(5): 1022-1028.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada
- Nilsen, E.B., I. Herfindal and J.D.C. Linnell. 2005. Can intra-specific variation in carnivore home-range size be explained using remote-sensing estimates of environmental productivity? *Ecoscience* 12(1): 68-75.
- Raine, R.M. and J.L. Kansas. 1990. Black bear seasonal food habitats and distribution by elevation in Banff National Park, Alberta. International Conference on Bear Research and Management 8:297-304.
- Samson, C and J. Huot. 1994. Ecologie et dynamique de la population d;ours noirs (Ursus americanus) du Parc national de la Mauricie. Rapport final remis a Parcs Canada. Department de biologie, University Laval, Sainte-Foy.214 pp. Cited in: Centre D'expertise En Analyse Environnementale Du Québec. 2006. Paramètres d'exposition chez les mammifères Ours noir. Fiche descriptive. Ministère du Développement durable, de l'Environnement et des Parcs du Québec, 17 p.
- Stevens, V. and S. Lofts. 1988. Wildlife Habitat Handbook for the Southern Interior Ecoprovince. Wildlife Habitat Research WHR-28, Wildlife Report No. R-15. Ministry of Environment (Wildlife Branch) and Ministry of Forests (Research Branch), Victoria, B.C
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Deer Mouse (Peromyscus maniculatus)

Characteristic	Determination	Notes (With References)
General Habitat Type	Variable habitats	(Banfield 1974)
		f diverse habitats. Forested, shrub-steppe, arid
Specific Habitat Type	grassland, rocky alpine habitats, grassy areas, human dwellings and	
	caves (Banfield 19	974; Sullivan et al. 2000).
		Foraging range is variable based on food availability
	_	(Teferi and Millar 1993; Bowman et al. 1999).
Foraging Range Size	<u>Area</u> : 120 m ²	Increased food abundance will decrease range
(All Seasons)	$(120-4000 \text{ m}^2)$	(Nagorsen 2005). In British Columbia, females have
		a smaller range limited to ~2500 m ² , while males
		range can extend to ~4000 m ² (Taitt 1981).
Body Weight	<u>Avg</u> : 21.7 g (15.0 – 41.8g)	(Nagorsen 2005)
Foraging Guild	Omnivore	(Eder and Pattie 2001)
Foraging Behavior	Grazer/hunter	(Eder and Pattie 2001)
Diet Proportions		
Ground Insects	45%	The deer mouse is omnivorous, feeding on seeds of
Berries/seeds	30%	trees, shrubs and grasses, a wide range of
Grasses	15%	invertebrates, and occasionally fungi (Nagorsen
Mushroom	5%	2005). A study by Wolff et al. (1985) presented the
Earthworms	5%	results of dissected stomach content from seventy (70) deer mice captured from the Mountain Lake
		Biological Station in southwestern Virginia.
		Although diet varied seasonally, the overall averages
		for deer mouse stomach contents were as follows:
		47.1% arthropods, 2.4% lepidopteran (moth and
		butterfly) larvae, 10.2% adult lepidopterans, 15.1%
		fruit, 8.6% green vegetation, 4.3% fungi, 10.2% nuts
	0.27 kg wet	and seeds, and 2.0% unknown. This rate is based on an average of 6 food ingestion
Food Ingestion Rate	food/kg wet	rates for adults, both sexes in US EPA 1993 in g/g
Food ingestion ivate	BW/day	day (wet weight).
	Dirauy	This rate is based on a 0.19 g/g day water ingestion
Water Ingestion Rate	0.19 L/kg wet	rate for adult deer mice (both sexes) and assumes a
	BW/day	water density of 1g/ml)
Incidental Soil or	<2.0 % of dry	Based on white-footed mouse (US EPA 1993; Beyer
Sediment Ingestion	food ingestion	et al., 1994)
		Ct al., 1777)
Rate	rate	

Deer Mouse References

- Banfield, A.W.F. 1974. The Mammals of Canada. University of Toronto Press, Toronto, Canada.
- Bowman, J.C., M. Edwards, L.S. Sheppard and G.J. Forbes. 1999. Record distance for non-homing movement by a deer mouse, *Peromyscus maniculatus*. *Canadian Field-Naturalist* 113:292-293.
- Beyer, W.N., E.E. Connor and S. Gerould. 1994. Estimates of soil ingestion by wildlife. *The Journal of Wildlife Management* 58(2):375-382.
- Eder, T. and D. Pattie. 2001. Mammals of British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Nagorsen, D.W. 2005. Rodents & Lagomorphs of British Columbia. Volume 4, The Mammals of British Columbia. Royal BC Museum Handbook, Royal BC Museum, Victoria, Canada.
- Sullivan, T.P., D.S. Sullivan and P.M.F. Lindgren. 2000. Small mammals and stand structure in young pine, seed-tree, and old-growth forest, southwest Canada. *Ecological Applications* 10(5): 1367-1383.
- Taitt, M.J. 1981. The effect of extra food on small rodent populations: I. Deermice (*Peromyscus maniculatus*). *Journal of Animal Ecology* 50(1): 111-124.
- Teferi, T. and J.S. Millar. 1993. Long distance homing by the deer mouse, *Peromyscus maniculatus*. *Canadian Field–Naturalist* 107(1): 109-111.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Wolff, J.O., R.D. Dueser and K.S. Berry. 1985. Food habits of sympatric *Peromyscus leucopus* and *Peromyscus maniculatus*. *Journal of Mammalogy* 66: 795-798.

Mallard (Anas platyrhynchos)

Characteristic	Determination	Notes (With References)
General Habitat Type	Wetlands	(Godfrey 1986)
Specific Habitat Type	Very adaptable. Preferred habitats include ponds, lakes, marshes, river bends, bays ditches and city ponds. Prefers freshwater but frequents saltwater in the winter (Godfrey 1986).	
Foraging Range Size (Minimum is for Breeding Season)	<u>Area</u> : 9.2 ha (9.2 – 240 ha)	The mallard is migratory but may breed and overwinter in Canada. Habitat size is highly dependent upon location. Smaller range sizes are found in prairie pothole habitats (~9 ha) and larger range sizes are found in northern Minnesota forests (~240 ha) (Gilmer at al. 1975; Titman 1983; Dwyer at al. 1979).
Body Weight	Avg: 1.2 kg Male: 1.1 kg (0.5 -1.7 kgg) Female: 1.2 kg (0.7 -1.7 kg)	(Bellrose 1976)
Foraging Guild	Omnivore	(Drillling et al. 2002)
Foraging Behavior	Dabbling	(Ehrlich et al. 1988)
Diet Proportions		
Aquatic Plants	50%	Mallards are dabbling ducks; therefore, they do not
Aquatic Invertebrates	40%	dive underwater for food, but feed at or just below
Berries/Seeds	5%	the surface. Most of the mallard diet is made up of
Other (Ground Insects, Flying Insects, Fish)	5%	plant material, such as wetland plants, seeds and grains (Palmer 1976; Hughes and Young 1982; Delnicki and Reinecke 1986; Gruenhagen and Fredrickson 1990). In southwestern British Columbia, diet analysis, by percent dry weight of food in the esophagus, determined that individuals
		ate 30.5% leaves and roots, 13% potatoes, and the remainder seeds (Lovvorn and Baldwin 1996). In winter, green vegetation such as weeds, grass and sedge seeds increased to 52% and potatoes remained constant at 13% (Hirst and Easthope 1981). Laying females were found to eat more animal food (72%) when compared to males (38%) and non-laying

Characteristic	Determination	Notes (With References)
		females (37%) (Swanson et al. 1985). Mallards in
		other areas have also been observed eating insects
		and other invertebrates (Ehrlich et al. 1988;
		Gruenhagen and Fredrickson 1990; Drilling et al.
		2002).
		This is based on the allometric equation (Nagy 1987
	0.05 kg dry	in US EPA 1993) for total dry food intake for all
Food Ingestion Rate	food/kg wet	birds (g/day;(0.648*BW ^{0.651}) divided by BW. The
	BW/day	average BW provided in this table was used for
		these calculations.
	0.06 L/kg wet	This is based on the allometric equation (Calder and
	BW/day	Braun 1983 in US EPA 1993) for total water intake
Water Ingestion Rate		for all birds (L/day; (0.059*BW ^{0.67}) divided by BW.
		The average BW provided in this table was used for
		these calculations.
Incidental Soil or	2.0 -3.3% of dry	Beyer et al. 1994, US EPA 1993
Sediment Ingestion	food ingestion	
Rate	rate	

Mallard References

- Bellrose, F.C. 1976. Ducks, Geese and Swans of North America. 2nd ed. Stackpole Books, Harrisburg, Pennsylvania, 534 p.
- Beyer, W.N., E.E. Connor and S. Gerould. 1994. Estimates of soil ingestion by wildlife. *The Journal of Wildlife Management* 58(2):375-382.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Delnicki, D. and K.J. Reinecke. 1986. Mid-winter food use and body weights of mallards and wood ducks in Mississippi. *The Journal of Wildlife Management* 50(1):43-51.
- Drilling, N., R. Titman and F. Mckinney. 2002. Mallard (Anas platyrhynchos), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/658.
- Dwyer, T.J., G.L. Krapu and D.M. Janke. 1979. Use of prairie pothole habitat by breeding mallards. *The Journal of Wildlife Management* 43:526-531.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.

- Gilmer, D.S., I.J. Ball, L.M. Cowardin, J.H. Riechmann and J.R. Tester. 1975. Habitat use and home range of mallards breeding in Minnesota. *The Journal of Wildlife Management* 39: 781-789.
- Godfrey, W.E. 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa.
- Gruenhagen, N.M. and L.H. Fredrickson. 1990. Food use by migratory female mallards in northwest Missouri. *The Journal of Wildlife Management* 54(4): 622-626.
- Hirst, S.M. and C.A. Easthope. 1981. Use of agricultural lands by waterfowl in southwestern British Columbia. *The Journal of Wildlife Management* 45:454-462.
- Hughes, J.H. and E.L. Young, Jr. 1982. Autumn foods of dabbling ducks in southeast Alaska. The *Journal of Wildlife Management* 46:259-263.
- Lovvorn, J.R. and J.R. Baldwin. 1996. Intertidal and farmland habitats of ducks in the Puget Sound region: A landscape perspective. *Biological Conservation* 77(1): 97-114.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada
- Palmer, R.S. 1976. Handbook of North American birds. Volume 2. Waterfowl (first part). Whistling Ducks, Swans, Geese, Shelducks, Dabbling ducks. Yale University Press, New Haven. 521 pp.
- Swanson, G.A., M.I. Meyer and V.A. Adomaitis. 1985. Foods consumed by breeding mallards on wetlands of south-central North Dakota. *The Journal of Wildlife Management* 49:197-20.
- Titman, R.D. 1983. Spacing and three-bird flights of mallards breeding in pothole habitat. *Canadian Journal of Zoology* 61:839-84
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Spotted Sandpiper (*Actitis macularia*)

Characteristic	Determination	Notes (With References)
General Habitat Type	Shoreline	(Godfrey 1986)
Specific Habitat Type	_	or muddy shores of interior lakes, ponds and streams salt water. Prefers relatively open areas (Godfrey
Foraging Range Size (Minimum is for Breeding Season)	<u>Area</u> : 0.08 ha (0.08 - 1.2 ha)	The spotted sandpiper is migratory and does not overwinter in Canada. Breeding range for spotted sandpiper is generally quite small (Miller and Miller 1948; Hays 1972; Oring and Knudson 1972; Oring et al. 1991; Oring et al. 1997).
Body Weight	Avg: 37.5 g Males: 36.9 g (30.0 - 46.0 g) Females: 38.0 g (no range given)	Season and location dependent (Irving 1960).
Foraging Guild	Insectivore	(Oring et al. 1997)
Foraging Behavior	Ground Cleans	(Ehrlich et al. 1988)
Diet Proportions		
Ground Insects	50 %	The spotted sandpiper forages principally for
Aquatic Invertebrates	30%	terrestrial and aquatic invertebrates but their diet can
Flying Insects	10%	also include fish (Oring et al. 1997). Primary prey
Aquatic Plants	5%	items include flies, grasshoppers, crickets, beetles,
Other (Amphibians, Fish)	5%	caterpillars, worms, mollusks and crustaceans, and spiders (Nelson 1939; Cramp and Simmons 1983; Ehrlich et al. 1988). Several studies have determined that midges and mayflies are a major food source for
	0.18 kg dry	spotted sandpipers (Maxson and Oring 1980; Lank et al. 1985). Ehrlich et al. (1988) noted that spotted sandpipers can deftly catch flying insects and that invertebrates are also picked off the water's surface. This is based on the allometric equation (Nagy 1987)
Food Ingestion Rate	food/kg wet BW/day	in US EPA 1993) for total dry food intake for all birds (g/day;(0.648*BW ^{0.651}) divided by BW. The average BW provided in this table was used for these calculations. ²¹

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²¹ The US EPA (1993) species specific value for spotted sandiper is based on juveniles

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.17 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Spotted Sandpiper References

- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Cramp, S. and K.E.L. Simmons. 1983. Handbook of the Birds of Europe, the Middle East and North Africa: The Birds of the Western Palearctic. Volume 3. Oxford University Press, Oxford, U.K.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Godfrey, W.E. 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa.
- Hays, H. 1972. Polyandry in the spotted sandpiper. Living Bird 11: 43-57.
- Irving, L. 1960. Birds of Anaktuvuk Pass Kobuk and Old Crow: A Study in Arctic Adaptation. Bulletin U.S. Natural Museum. 217: 1-409
- Lank, D.B., L.W. Oring and S.J. Maxson. 1985. Mate and nutrient limitation of egg-laying in a polyandrous shorebird. *Ecology* 66: 1513-1524
- Maxson, S.J. and L.W. Oring. 1980. Breeding season time and energy budgets of the polyandrous Spotted Sandpiper. *Behaviour* 74: 200-263.
- Miller, J.R. and J.T. Miller. 1948. Nesting of the spotted sandpiper at Detroit, Michigan. *Auk* 65:558-567.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128.Vancouver, Canada

- Nelson, T. 1939. The biology of the spotted sandpiper (Actitis macularia, Linn.). Ph.D. Thesis, University of Michigan, Ann Arbor.
- Oring, L.W. and M.L. Knudson. 1972. Monogamy and polyandry in the spotted sandpiper. *Living Bird* 11: 59-73.
- Oring, Lewis W., Elizabeth M. Gray and J. Michael Reed. 1997. Spotted Sandpiper (Actitis macularius), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/289
- Oring, L.W., J.M. Reed, M.A. Colwell, D.B. Lank and S.J. Maxson. 1991. Factors regulating annual mating success and reproductive success in spotted sandpipers (*Actitis macularia*). *Behavioral Ecology and Sociobiology* 28:433-44
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Lesser Scaup (Aythya affinis)

Characteristic	Determination	Notes (With References)
General Habitat Type	Marine and Freshwater	(Godfrey 1986; Campbell et al. 1990)
Specific Habitat Type	Interior lakes and ponds, low islands and moist sedge meadows. During migration found in coastal bays, estuaries, rivers and large lakes (Godfrey 1986).	
Foraging Range Size (Minimum is for Breeding Season)	<u>Area</u> : 0.1 km² (0.1 -17.1 km ²)	The lesser scaup is migratory but may breed and overwinter in Canada. Foraging range in winter is quite large (Herring and Collazo 2005). References for summer range were not found, but range is likely much smaller than in winter. A range of approximately 10 ha, similar to mallard, was considered to be reasonable.
Body Weight	Avg: 707 g Male: 745 g Female: 669 g	Mass variable based on season (Moore 1991).
Foraging Guild	Insectivore	(Austin et al. 1998)
Foraging Behavior	Surface Dives	(Ehrlich et al. 1988)
Diet Proportions		
Insects	45%	Diet is variable based on season location and
Leeches	25%	reproductive stage (Afton et al. 1991). The primary
Amphipods	20%	foods consumed are insects, crustaceans
Aquatic plants and seeds	10%	(amphipods) and mollusks, while seeds and portions of aquatic plants are a significant component in
	ı	some areas (Dirschl 1969; Austin et al. 1998; Lindeman and Clark 1999; Strand 2005). In Manitoba, diet studies determined that before periods of rapid follicular growth the percent occurrence of insects, amphipods, leeches and seeds were respectively (50.2%), (28.9%), (17.4%) and (3.1%) (Afton et al.1991).
Food Ingestion Rate	0.07 kg dry food/kg wet BW/day	This is based on the allometric equation (Nagy 1987 in US EPA 1993) for total dry food intake for all birds (g/day;(0.648*BW ^{0.651}) divided by BW. The average BW provided in this table was used for these calculations.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.07 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Lesser Scaup References

- Afton, A.D., R.H. Hier and S.L. Paulus. 1991. Lesser scaup diets during migration and winter in the Mississippi Flyway. *Canadian Journal of Zoology* 69:328-333.
- Austin, J.E., C.M. Custer and A.D. Afton. 1998. Lesser Scaup (*Aythya affinis*). The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/338.
- Campbell, R.W., N.K. Dawe, I. McTaggart-Cowan, J.M. Cooper, G.W. Kaiser and M.C.E. McNall. 1990. The Birds of British Columbia, Volume 1: Introduction and loons through waterfowl. Royal British Columbia Museum, Victoria.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Dirschl, H.J. 1969. Foods of lesser scaup and blue-winged teal in the Saskatchewan River Delta. *The Journal of Wildlife Management* 33(1): 77-87.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Godfrey, W.E. 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa.
- Herring, G. and J.A. Collazo. 2005. Habitat use, movements and home range of wintering lesser scaup in Florida. *Waterbirds* 28 (1): 71-78.
- Lindeman, D.H. and R.G. Clark. 1999. Amphipods, land-use impacts, and lesser scaup (*Aythya affinis*) distribution in Saskatchewan wetlands. *Wetlands* 19(3): 627-638.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada
- Moore, J. L. 1991. Habitat-related Activities and Body Mass of Wintering Redhead Ducks on Coastal Ponds in South Texas. Masters Thesis, Texas A&M University. College Station.
- Strand, K.A. 2005. Diet and Body Composition of Migrating Lesser Scaup (Aythya affinis) in Eastern South Dakota. Masters of Science Thesis, South Dakota State University.

USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Common Merganser (Mergus merganser)

Characteristic	Determination	Notes (With References)
General Habitat Type	Freshwater	(Godfrey 1986)
Specific Habitat Type	Lakes and rivers in or near woodlands during summer. In winter, they are found in rivers and open water lakes. Less often observed in salt water (Godfrey 1986).	
Foraging Range Size (Minimum is for Breeding Season)	<u>Area</u> : 0.04 km ² (0.04 - 13.9 km ²) <u>Linear</u> : 0.7 km (0.7 - 2.5 km)	The common merganser is migratory but may breed and overwinter in Canada. Habitat size is variable based on season, location and habitat suitability. Area Estimates - (Ross 1987; Cadman et al. 1987; Erskine 1987). Linear estimates - (Wood 1986).
Body Weight	Avg: 1.5 kg Male:1.7 kg (1.3-2.2 kg) Female: 1.3 kg (0.9-1.8 kg)	Variable by gender and season (Erskine 1972; Cramp and Simmons 1977).
Foraging Guild	Piscivore	(Ehrlich et al. 1988)
Foraging Behavior	Surface Dives	(Ehrlich et al. 1988)
Diet Proportions		
Fish	90%	Common mergansers primarily consume fish;
Aquatic Invertebrates	8%	however, they also occasionally eat aquatic
Aquatic Plants	2%	invertebrates (e.g., mollusks, crustaceans and
		worms), frogs, small mammals, birds and plants (Palmer 1976; Ehrlich et al. 1988; Del Hoyo et al. 1992; Mallory and Metz 1999). Diet studies from stomach analyses of 48 common mergansers in interior British Columbia during the summer found that the following fish species were consumed (% in diet): sculpin and chub (each 23.6%), shiner (13.7%), sucker (9.8%) and crayfish (5.9%) (Munro and Clemens 1932). Downy young eat mostly aquatic invertebrates but switch to fish when they are about 12 days old (White 1957).
Food Ingestion Rate	0.05 kg dry food/kg wet BW/day	This is based on the allometric equation (Nagy 1987 in US EPA 1993) for total dry food intake for all birds (g/day;(0.648*BW ^{0.651}) divided by BW. The average BW provided in this table was used for these calculations.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.05 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Common Merganser References

- Cadman, M.P., P.F.J. Eagles and F.M. Helleinen. 1987. Atlas of Breeding Birds of Ontario. University of Waterloo, Waterloo.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Cramp, S. and K.E.L. Simmons. 1977. The Birds of the Western Palearctic, Vol. 1. Ostrich to Ducks. Oxford University Press, Oxford.
- Del Hoyo, J., A. Elliot and J. Sargatal. 1992. Handbook of the Birds of the World, Volume 1: Ostrich to Ducks. Lynx Edicions, Barcelona.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Erskine, A.J. 1972. Buffleheads. Monograph Series No. 4, Canadian Wildlife Service.
- Erskine, A.J. 1987. Waterfowl breeding population surveys, Atlantic provinces. Canadian Wildlife Service Occasional Paper Series No. 60.
- Godfrey, W.E. 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa.
- Mallory, M. and K. Metz. 1999. Common Merganser (Mergus merganser), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/442.
- Munro, J. A. and W. A. Clemens. 1932. Food of the American merganser (*Mergus merganser americanus*) in British Columbia: A preliminary paper. *Canadian Field-Naturalist* 46:166-168.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128.Vancouver, Canada

- Palmer, R.S. 1976. Handbook of North American Birds. Volume 2. Waterfowl (first part). Whistling Ducks, Swans, Geese, Shelducks, Dabbling ducks. Yale University Press, New Haven. 521 pp.
- Ross, R.K. 1987. Interim report on waterfowl breeding pair surveys in northern Ontario, 1980-1983. Canadian Wildlife Service Progress Report No. 168.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- White, H.C. 1957. Food and Natural History of Mergansers on Salmon Waters in the Maritime Provinces of Canada. Fish Resource Board of Canada Bulletin No. 116.
- Wood, C.C. 1986. Dispersion of common merganser (*Mergus merganser*) breeding pairs in relation to the availability of juvenile Pacific salmon in Vancouver Island streams. *Canadian Journal of Zoology* 64:756-765.

Bald Eagle (Haliaeetus leucocephalus)

Characteristic	Determination	Notes (With References)
General Habitat Type	Near Water	(Godfrey 1986)
Specific Habitat Type	In the areas adjacent to large rivers, lakes and coastline (Godfrey 1986).	
Foraging Range Size (Minimum is for Breeding Season)	<u>Area</u> : 2.1 km² (2.1 – 21.6 km ²)	The bald eagle is migratory but may breed and overwinter in Canada. Variable based on habitat, food availability and season (Frenzel 1984; Gerrard et al. 1992; Garrett et al. 1993; Watson 2002).
Body Weight	Avg: 4.7 kg Male: 4.1 kg (3.7 – 4.9 kg) Females: 5.4 kg (4.6 – 6.4 kg)	(Imler and Kalmbach 1955)
Foraging Guild	Piscivore	(Ehrlich et al. 1988)
Foraging Behavior	Hunting (High patrol and swoops)	(Ehrlich et al. 1988)
Diet Proportions		
Fish	65%	The bald eagle is an opportunistic forager and
Small Mammals	20%	scavenger that preferentially eats fish but also
Birds	15%	consumes a variety of mammalian, avian, and reptilian prey (Todd et al. 1982; Ehrlich et al. 1988; Buehler 2000; Watson 2002). Stalmaster (1987) also concluded that fish are preferred throughout range but consumption is variable based on location and seasonal availability. After a review of 20 different diet composition studies, Stalmaster (1987) determined the average bald eagle diet to be composed of 56% fish, 28% birds, 14% mammals and 2% other. In prey remains collected from eagle nests in western Washington, Knight et al. (1990) found 49 bird species, eight mammal species and 14 invertebrate species with 55% of remains being birds; however, based on direct observation, 92% of delivered prey items were fish. Watson (2002) studied bald eagles in four different aquatic habitats and determined that those living on freshwater lakes consumed the highest amount of fish (84%) and had the smallest home range size (210 ha) compared those in marine, bay or river habitats.

Characteristic	Determination	Notes (With References)
	0.12 kg wet	This rate is based on 0.12 g/g day ingestion rate for
Food Ingestion Rate	food/kg wet BW/day	adults, both sexes (US EPA 1993)
Water Ingestion Rate	0.04 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Bald Eagle References

- Buehler, D.A. 2000. Bald Eagle (Haliaeetus leucocephalus), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/506.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: A Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Frenzel, R.W. 1984. Environmental contaminants and ecology of bald eagles in southcentral Oregon. Phd Thesis. Oregon State University, Corvallis.
- Garrett, M.G., J.W. Watson and R.G. Anthony. 1993. Bald eagle home range and habitat use in the Columbia River estuary. *The Journal of Wildlife Management* 57:19-27.
- Gerrard, J.M., P.N. Gerrard, G.R. Bortolotti and E.H. Dzus. 1992. A 24-year study of bald eagles on Besnard Lake, Saskatchewan. *Journal of Raptor Research* 26:159-16.
- Godfrey, W.E., 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa.
- Imler, R. H. and E. R. Kalmbach. 1955. The Bald Eagle and its Economic Status. U.S. Fish and Wildlife Service. Circular 30, Washington, D.C.
- Knight, R.L., P.J. Randolph, G.T. Allen, L.S. Young and R.T. Wigen. 1990. Diets of nesting bald eagles, Haliaeetus leucocephalus, in western Washington. *Canadian Field-Naturalist* 104:545-551.

- Stalmaster, M.V. 1987. The Bald Eagle. Universe Books, New York.
- Todd, C.S., L.S. Young, R.B. Owen and F.J. Gramlich. 1982. Food habits of bald eagles in Maine. *The Journal of Wildlife Management* 46:636-645.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Watson, J.W. 2002. Comparative home ranges and food habits of bald eagles nesting in four aquatic habitats in western Washington. *Northwestern Naturalist* 83(3): 101-108

Great Blue Heron (*Ardea herodias***)**

Characteristic	Determination	Notes (With References)
General Habitat Type	Marine and Freshwater	(Godfrey 1986)
Specific Habitat Type	_	water at the edges of streams, river sloughs, lakes, udflats and marshes (Godfrey 1986).
Foraging Range Size (Minimum is for Breeding Season)	<u>Area</u> : 16.6 km ² (16.6 – 2827 km ²)	The great blue heron is migratory but may breed and overwinter in Canada. Little information is available on foraging home range. References indicate that great blue herons forage between 2.3 km and 30 km linear distance away from heronries (Parris 1979; Dowd and Flake 1985; Butler 1991; Vennesland and Butler 2011)
Body Weight	Avg: 2.3 kg Male: 2.5 kg Female: 2.1 kg	(Simpson 1984)
Foraging Guild	Piscivore	(Ehrlich et al. 1988)
Foraging Behavior	Stalk and Strike	(Ehrlich et al. 1988)
Diet Proportions		
Fish	65%	The diet of the great blue heron consists principally
Small Mammals	25%	of fish; however, small mammals, amphibians,
Aquatic Invertebrates	10%	invertebrates, birds and crustaceans are also taken
		when they are available (Palmer 1962; Kushlan 1978; Peifer 1979; Verbeek and Butler 1989; Butler 1991). Small fish such as gunnels, sculpins, flounders, sticklebacks and shiner perch are the primary foods of breeding herons in British Columbia (Harfenist et al. 1995; Butler 1997). Herons also eat marine invertebrates such as mud shrimp, isopods and crabs (Verbeek and Butler 1989). Butler (1991) suggested foraging preference is based on individual ability with juveniles learning hunting skills in grasslands, adult females focusing on intertidal beaches, and males favoring riverbanks. Some research suggests that voles may be important in the diet of nestlings (Collazo 1979; Butler 1991)

Characteristic	Determination	Notes (With References)
	0.18 kg wet	This rate is based on 0.18 g/g day ingestion rate for
Food Ingestion Rate	food/kg wet	adults, both sexes (US EPA 1993). ²²
	BW/day	
	0.04 L/kg wet BW/day	This is based on the allometric equation (Calder and
		Braun 1983 in US EPA 1993) for total water intake
Water Ingestion Rate		for all birds (L/day; (0.059*BW ^{0.67}) divided by BW.
		The average BW provided in this table was used for
		these calculations.
Incidental Soil or		
Sediment Ingestion	None identified	Refer to Section 2.1.8.
Rate		

Great Blue Heron References

- Butler, R.W. 1991. Habitat Selection and Time of Breeding in the Great Blue Heron (Ardea herodias). Ph.D Thesis, University of British Columbia, Vancouver.
- Butler, R.W. 1997. The Great Blue Heron: A Natural History and Ecology of a Seashore Sentinel. UBC Press, Vancouver.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Collazo, J.A. 1979. Breeding Biology and Food Habits of the Great Blue Heron at Heyburn State Park, Benewah County, Idaho. Master of Science Thesis, University of Idaho.
- Dowd, E. M. and L. D. Flake. 1985. Foraging habitats and movements of nesting Great Blue Herons in a prairie river ecosystem, South Dakota. J. Field Ornithol. 56:379-387.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Godfrey, W.E., 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa
- Harfenist, A., P.E. Whitehead, W.J. Cretney and J.E. Elliot. 1995. Food Chain Sources of Polychlorinated Dioxins and Furans to Great Blue Herons (*Ardea herodias*) Foraging in the Fraser River Estuary, British Columbia. Canadian Wildlife Service Technical Report No. 169. Canadian Wildlife Service, Delta, Canada.

²² Sample et al. (1996) use a food ingestion rate of 0.42 kg/day. Using the body weight from this table, this converts to 0.183 kg/kg bw/day, very similar to the US EPA food ingestion rate reported in the table above.

- Kushlan, J. A. 1978. Feeding Ecology of Wading Birds. Pages 249-298 Cited in Wading birds. (Sprunt IV, A., J.C. Ogden, and S. Winkler, Eds.) National Audubon Society Research Report No. 7, New York.
- Palmer, R.S. 1962. Handbook of North American Birds. Volume 1. Yale University Press, New Haven, CT.
- Parris, R.W. 1979. Aspects of great blue heron (*Ardea herodias*) foraging ecology in southwest Lake Erie. Master's Thesis. Ohio State University, Columbus.
- Peifer, R.W. 1979. Great blue herons foraging for small mammals. *The Wilson Bulletin* 91:63-631.
- Simpson, K. 1984. Factors Affecting Reproduction in Great Blue Herons (*Ardea herodias*). Masters of Science Thesis, University of British Columbia, Vancouver, Canada.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Vennesland, Ross G. and Robert W. Butler. 2011. Great Blue Heron (*Ardea herodias*), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/025
- Verbeek, N.A.M. and R.W. Butler. 1989. Feeding Ecology of Shoreline Birds in the Strait of Georgia. Pages 74-81. Cited in The ecology and status of marine and shoreline birds in the Strait of Georgia, British Columbia. (Vermeer, K. and R.W. Butler, Eds.) Canadian Wildlife Service Special Publication, Ottawa.

Common Loon (Gavia immer)

Characteristic	Determination	Notes (With References)
General Habitat Type	Marine and fresh water	(Godfrey 1986)
Specific Habitat Type	Lakes and rivers d winter (Godfrey 1	luring the summer; seacoasts and large lakes in the 986).
Foraging Range Size (Minimum is for Breeding Season)	<u>Area</u> : 4.4 ha (4.4 – 59.0 ha)	The common loon is migratory but may breed and overwinter in Canada. Foraging range is highly variable based on territory quality, prey and nest site availability (Zimmer 1979; Miller and Dring 1988; Evers et al. 2000; Evers 2001; Evers et al. 2010)
Body Weight	Avg: 5.3 kg Male: 6.0 kg (5.5 – 6.4 kg) Female: 4.7 kg (4.3 – 5.0 kg)	(Evers et al. 2010)
Foraging Guild	Piscivore	(Ehrlich et al. 1988)
Foraging Behavior	Surface Dives	(Ehrlich et al. 1988)
Diet Proportions		
Fish	90%	Loons are opportunistic predators that primarily
Aquatic Invertebrates	10%	consume fish (Barr 1996). Perch are known to be a
		common component of the diet and are preferentially consumed relative to salmonids perhaps because salmonids are more difficult to capture (Evers et al. 2004). Aquatic invertebrates, such as crayfish, make up a significant secondary component of the diet, especially when fish are difficult to capture due to poor water visibility (Barr 1973). In some cases, aquatic invertebrates made up over 30% percent of the common loon diet (Barr 1973).
Food Ingestion Rate	0.19 kg wet food/kg wet BW/day	(Barr 1996) ²³ .

²³ Based on a pair of hand-reared birds (male and female, with young) fed a diet of fish and an average body weight of 5 kg.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.03 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Common Loon References

- Barr, J. F. 1973. Feeding Biology of the Common Loon (*Gavia immer*) in Oligotrophic Lakes of the Precambrian Shield. Phd Thesis, University of Guelph, Ontario.
- Barr, J. F. 1996. Aspects of common loon (*Gavia immer*) feeding biology on its breeding ground. *Hydrobiologia* 321:119-144.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Evers, D. 2001. Common Loon Population Studies: Continental Mercury Patterns and Breeding Territory Philopatry. Ph.D. dissertation, University of Minnesota, St. Paul, Minnesota.
- Evers, D., J.D. Kaplan, P.S. Reaman, J.D. Paruk and P. Phifer. 2000. A Demographic Characterization of the Common Loon in the Upper Great Lakes. Pages 78-90 in Loons: Old history and new findings. Proceedings of a Symposium from the 1997 meeting. (McIntyre, J. W. and D. Evers, Eds.) American Ornithology Union, North America. Loon Fund, Holderness, NH.
- Evers, D., O. P. Lane, C. R. DeSorbo and L. Savoy. 2004. Assessing the Impacts of Methylmercury on Piscivorous Birds Using a Wildlife Criterion Value Based on the Common Loon, 1998-2001. Report BRI 2002-08 submitted to Maine Dept. Environ. Protection, BioDiversity Research Institute, Gorham, Maine.
- Evers, D.C., J.D. Paruk, J.W. Mcintyre and J.F. Barr. 2010. Common Loon (*Gavia immer*), The Birds of North America Online (A. Poole, Edits.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/313.
- Godfrey, W.E., 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa
- Miller, E. and T. Dring. 1988. Territorial defense of multiple lakes by common loons: A preliminary report. Pages 1-14 in Papers from the 1987 Conference on Common Loon

- Research and Management. (Strong, P. I. V., Ed.) North American Loon Fund, Meredith, NH.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Zimmer, G. E. 1979. The Status and Distribution of the Common Loon in Wisconsin. University of Wisconsin.

Ruffed Grouse (Bonasa umbellus)

Characteristic	Determination	Notes (With References)
General Habitat Type	Forest	(Godfrey 1986)
Specific Habitat Type	Second growth and mixed woodlands. Forest and stream edges,	
Specific Habitat Type	openings and alde	r or willow bordered ravines (Godfrey 1986).
Foraging Range Size (All Seasons)	<u>Area</u> : 1.0 ha (1.0 – 180 ha)	The ruffed grouse is a resident species. Foraging ranges are quite variable between studies (Thompson and Fritzell 1989). Seasonally variable with summer ranges smaller then winter ranges (Bump et al. 1947; Archibald 1975; Godfrey 1975; Epperson 1988; Maxson 1989; Scott et al. 1998).
Body Weight	Avg: 552 g Males: 604 g Females: 500 g	(Rusch et al. 2000)
Foraging Guild	Omnivore	(Ehrlich et al. 1988)
Foraging Behavior	Foliage Browsing	(Ehrlich et al. 1988)
Diet Proportions		
Leaves (Shrubs/Trees)	55%	Twigs and buds, particularly of trembling aspen are
Berries	30%	important food for ruffed grouse (Svoboda and
Insects and Invertebrates	15%	Gullion 1972; Jakubas and Gullion 1991). A number of other plant species as well as fruits also make up
	0.06 kg dry	part of ruffed grouse diet in some regions (Stafford and Dimmick 1979; Rusch et al. 2000). Insects and invertebrates are believed to be an important food source for chicks and are also eaten by adults (Bump et al. 1947; Ehrlich et al. 1988).
Food Ingestion Rate	food/kg wet BW/day	(Guglielmo and Karasov 1995) ²⁴

²⁴ Based on captured ruffed grouse (males and females, adults and juveniles) fed a natural diet. Grouse were fed a series of diets (Series 1 in Table 2 of the Guglielmo and Karasov (1995), those included in this study were diets where grouse did not have significant weight loss (aspen flower buds, hazel catkins and mixed wild browse). Although juveniles were included, body weight listed are similar to those presented in the module table.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.07 L/kg wet	This is based on the allometric equation (Calder and
	BW/day	Braun 1983 in US EPA 1993) for total water intake
		for all birds (L/day; (0.059*BW ^{0.67}) divided by BW.
		The average BW provided in this table was used for
		these calculations.
Incidental Soil or		
Sediment Ingestion	None identified	Refer to Section 2.1.8.
Rate		

Ruffed Grouse References

- Archibald, H.L. 1975. Temporal patterns of spring space use by ruffed grouse. *The Journal of Wildlife Management* 39:472-481.
- Bump, G., R.W. Darrow, F.C. Edminster and W.F. Crissey. 1947. The Ruffed Grouse: Life History, Propagation, Management. New York State Conservation Department. Albany.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Epperson, Jr., R.G. 1988. Population status, movements and habitat utilization of ruffed grouse on the Catoosa Wildlife Management Area, Cumberland County, Tennessee. Master's Thesis. University of Tennessee, Knoxville.
- Godfrey, G.A. 1975. Home range characteristics of ruffed grouse broods in Minnesota. *The Journal of Wildlife Management* 39:287-298.
- Godfrey, W.E., 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa
- Guglielmo, C.G. and W.H. Karasov. 1995. Nutritional Quality of Winter Browse for Ruffed Grouse. *The Journal of Wildlife Management* 59(3):427-436.
- Jakubas, W.J. and G.W. Gullion. 1991. Use of quaking aspen flower buds by ruffed grouse: Its relationship to grouse densities and bud chemical composition. *Condor* 93: 473–485.
- Maxson, S.J. 1989. Patterns of Activity and Home Range of Hens. Pp. 118–129. Cited in Ruffed Grouse (S. Atwater and J. Schnell, eds.). Stackpole Books, Harrisburg, PA.
- Rusch, D.H., S. Destefano, M.C. Reynolds and D. Lauten. 2000. Ruffed Grouse (*Bonasa umbellus*), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/515.
- Scott, J.G., M.J. Lovallo, G.L. Storm and W.M. Tzilkowski. 1998. Summer habitat use by ruffed grouse with broods in central Pennsylvania. *Journal of Field Ornithology* 69:474-485

- Stafford, S.K. and R.W. Dimmick. 1979. Autumn and winter foods of ruffed grouse in the southern Appalachians. *The Journal of Wildlife Management* 43: 121–127.
- Svoboda, F.J. and G.W. Gullion. 1972. Preferential use of aspen by ruffed grouse in northern Minnesota. *The Journal of Wildlife Management* 36: 1166–1180.
- Thompson III, F.R. and E.K. Fritzell. 1989. Habitat use, home range, and survival of territorial male ruffed grouse. *The Journal of Wildlife Management* 53:15-21.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Spruce Grouse (Dendragapus canadensis)

Characteristic	Determination	Notes (With References)
General Habitat Type	Forest	(Godfrey 1986)
Specific Habitat Type	Coniferous and m	ixed wood forests, muskeg, forest edges and openings
Specific Habitat Type	(Godfrey 1986).	
		The spruce grouse is a resident species and is
		generally sedentary with discrete territories and
Foraging Range Size	Area: 3.0 ha	overlapping but restricted ranges, generally never
(All Seasons)	(3 - 24 ha)	more than a few hectares (Ellison 1971; Herzog and
		Boag 1977; Robinson 1980; Lattner 1982; Campbell
		et al. 1990; Boag and Schroeder 1992).
		Variable depending on location and season (Boag
		and Schroeder 1992). Throughout the year, males
		generally weigh more then females with the
Body Weight	<u>Avg</u> : ~600 g	exception of the prelaying and laying stage (April
		and May) when females are heavier then males
		(Ellison and Weeden 1979; Boag and Schroeder
		1992).
Foraging Guild	Herbivore	(Ehrlich et al. 1988)
Foraging Behavior	Foliage Browse	(Ehrlich et al. 1988)
Diet Proportions		
Conifer Needles and	65%	Throughout the majority of its range, the spruce
Buds	0370	grouse eats pine needles (Pinus banksiana or Pinus
Berries	30%	contorta) as the main component of its diet
Other (Insects, Fungi)	5%	(Crichton 1963; Pendergast and Boag 1970; Naylor
		and Bendell 1989). In the absence of pine needles,
		spruce needles (<i>Picea glauca or Picea mariana</i>) are
		consumed. In periods without snow, the freshly growing tips, leaves, flowers and fruits of
		blueberries are a significant secondary component of
		the diet (Boag and Schroeder 1992). Spruce grouse
		are also known to occasionally eat insects and
		especially grasshoppers (Ehrlich et al. 1988) while
		fungi are believed to be an important component of
		the diet for chicks (DeFranceschi and Boag 1991).
	0.07 kg dry	This is based on the allometric equation (Nagy 1987
Food Ingestion Rate	food/kg wet	in US EPA 1993) for total dry food intake for all
	BW/day	birds (g/day;(0.648*BW ^{0.651}) divided by BW. The
		average BW provided in this table was used for
		these calculations.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.07 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Spruce Grouse References

- Boag, D.A. and M.A. Schroeder. 1992. Spruce Grouse (Falcipennis canadensis). The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/005.
- Campbell, R.W., N.K. Dawe, I. McTaggart-Cowan, J.M. Cooper, G.W. Kaiser and M.C.E. McNall. 1990. The Birds of British Columbia, Volume 2: Nonpasserines, Diurnal Birds of Prey through Woodpeckers. Royal British Columbia Museum and Canadian Wildlife Service
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Crichton, V. 1963. Autumn and winter foods of spruce grouse in central Ontario. *The Journal of Wildlife Management* 27(4):597.
- Defranceschi, P.F. and D.A. Boag. 1991. Summer foraging by spruce grouse: Implications for galliform food habits. *Canadian Journal of Zoology* 69: 1708-1711
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Ellison, L. N. 1971. Territoriality in Alaskan spruce grouse. Auk 88: 652-664.
- Ellison, L.N. and R.B. Weeden. 1979. Seasonal and local weights of Alaskan spruce grouse. *The Journal of Wildlife Management* 43:176-183.
- Godfrey, W.E. 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa
- Herzog, P. W. and D. A. Boag. 1977. Seasonal changes in aggressive behavior of female spruce grouse. *Canadian Journal of Zoology* 55:1734-1739.
- Lattner, M. T. 1982. Foods, Mobility, Habitat Selection and Density of Spruce Grouse. Master's Thesis. University, Toronto, Ontario.

- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada
- Naylor, B.J. and J.F. Bendell. 1989. Clutch size and egg size of spruce grouse in relation to spring diet, food supply, and endogenous reserves. *Canadian Journal of Zoology* 67: 969-980.
- Pendergast, B.A. and D.A. Boag. 1970. Seasonal changes in the diet of spruce grouse in central Alberta. *The Journal of Wildlife Management* 34(3): 605-611.
- Robinson, W. L. 1980. Fool Hen: the Spruce Grouse on the Yellow Dog Plains. University of Wisconsin Press, Madison.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Barn Swallow (Hirundo rustica)

Characteristic	Determination	Notes (With References)
General Habitat Type	Open areas	(Campbell et al. 1997)
	Urban backyards	of rural farmyards, farmlands, roadsides, weedy fields,
Specific Habitat Type	neat water such as	s lakes, marshes, streams, estuaries, sloughs and
	meadows, orchards and vineyards (Campbell et al. 1997).	
		The barn swallow is migratory and does not
Foraging Range Size	<u>Area</u> : 0.8 km²	overwinter in Canada. Foraging distance between
(Breeding Season)	$(0.8 - 4.5 \text{ km}^2)$	0.5 km and 1.2 km linear distance from nest site
		(Møller 1987; Samuel 1971).
	<u>Avg</u> : 18.7 g	
Body Weight	Males: 18.1 g	(Brown and Brown 1999)
	Females: 19.2 g	
Foraging Guild	Insectivore	(Ehrlich et al. 1988)
Foraging Behavior	Aerial Foraging	(Ehrlich et al. 1988)
Diet Proportions		
Insects	99%	Barn swallows are almost exclusively insectivorous,
Seeds/berries	1%	hawking insects (e.g., flies, moths, butterflies,
		dragonflies, and beetles) during flight over open land
		and water habitats (Brown and Brown 1999). In an
		analysis of 467 barn swallow stomachs from across
		North America, Beal (1918) revealed 99.8% animal
		matter. Many of the insects hawked aerially may
		have had an aquatic life stage. Berries and seeds
		have also been reported as being used occasionally
		(Ehrlich et al. 1988).
		This is based on the allometric equation (Nagy 1987
	0.26 kg dry	in US EPA 1993) for total dry food intake for
Food Ingestion Rate	food/kg wet	passerine birds (g/day;(0.398*BW ^{0.850}) divided by
	BW/day	BW. The average BW provided in this table was
		used for these calculations.
		This is based on the allometric equation (Calder and
Water Ingestion Rate	0.22 L/kg wet	Braun 1983 in US EPA 1993) for total water intake
	BW/day	for all birds (L/day; (0.059*BW ^{0.67}) divided by BW.
	D W/day	The average BW provided in this table was used for
		these calculations.
Incidental Soil or	None identified	Refer to Section 2.1.8.
Sediment Ingestion		
Rate		

Barn Swallow References

- Beal, F.E.L. 1918. Food Habits of the Swallows, a Family of Valuable Native Birds. U.S. Department of Agriculture Bulletin 619.
- Brown, C.R. and M. Bomberger Brown. 1999. Barn Swallow (Hirundo rustica), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/452.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Campbell, R.W., N.K. Dawe, I. McTaggart-Cowan, J.M. Cooper, G.W. Kaiser, M.C.E. McNall and G.E.J. Smith. 1997. The Birds of British Columbia. Volume 3: Passerines, Flycatchers through Vireos. Ministry of Environment, Lands and Parks and Canadian Wildlife Service. UBC Press, Vancouver.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Møller, A.P. 1987. Advantages and disadvantages of coloniality in the swallow, *Hirundo rustica*. Animal Behaviour 35:819-832
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada
- Samuel, D.E. 1971. The breeding biology of barn and cliff swallows in West Virginia. *The Wilson Bulletin* 83: 284–301.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Red-tailed Hawk (Buteo jamaicensis)

Characteristic	Determination	Notes (With References)
General Habitat Type	Forest and open habitats	(Godfrey 1986)
Specific Habitat Type	-	en country including pastures, fields and shrub- Usually near trees during nesting season (Godfrey
Foraging Range Size (Minimum is for Winter Season)	<u>Area</u> : 0.2 km² (0.2 - 50 km ²)	The red-tailed hawk is a resident species. Foraging range is variable and based on season, habitat, food availability and human disturbance. Foraging range is significantly lower in winter than in summer in most studies (Fitch et al. 1946; Orions and Kuhlman 1956; Austing 1964; Shelton 1971; Gates 1972; Misztal 1974; Lowe 1978; Peterson 1979; Bildstein 1987; Preston and Beane 2009).
Body Weight	Avg: 1.1 kg Males: 1.0 kg Females: 1.2 kg	Measured during the breading season (Palmer 1988).
Foraging Guild	Carnivore	(Ehrlich et al. 1988)
Foraging Behavior	Hunting (High Patrol; Swoops)	(Ehrlich et al. 1988)
Diet Proportions		
Rodents	85%	In Alberta, Luttich et al. (1970) found that 66% of
Birds	10%	the diet of red-tailed hawks consisted of small
Amphibians and Reptiles	5%	mammals including hares, ground-squirrels and voles. Waterfowl (i.e., approximately 18% of the
Food Ingestion Rate	0.10 kg wet food/kg wet BW/day	diet) were also taken on a regular basis (Luttich et al. 1970). Generally, more than 80% of a red-tailed hawks diet consists of rodents, with amphibians, snakes, birds and fish taken to a lesser extent (Gates 1972; Adamcik et al. 1979; Stinson 1980; Ehrlich et al. 1988; Janes 1984; Preston and Beane 2009). This rate is based on an average (0.099g/g per day) of 3 values (adult male and female in winter, adult male in summer) for adults, both sexes (US EPA 1993). ²⁵

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²⁵ Sample et al. (1996) reports a food ingestion rate of 0.109 kg wet food/day for the red-tailed hawk (derived from an updated version of Craighead and Craighead 1969; the 1956 version was used by the US EPA). Using the body weight in this table, this would convert to a rate of 0.097 kg/kg wet BW/day, which is very similar to the ingestion rate calculated using US EPA 1993 values.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.06 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

Red-tailed Hawk References

- Adamcik, R.S., A.W. Todd, and L.B. Keith. 1979. Demographic and dietary responses of redtailed hawks during a snowshoe hare fluctuation. *Canadian Field-Naturalist* 93(1):16-27
- Austing, G.R. 1964. The World of the Red-tailed Hawk. J.B. Lippincott Co. Philadelphia, PA.
- Bildstein, K.L. 1987. Behavioral ecology of red-tailed hawks (*Buteo jamaicensis*), rough-legged hawks (*Buteo lagopus*), northern harriers (*Circus cyaneus*), and American kestrels (*Falco sparverius*) in south central Ohio. Ohio Biological Survey Biology Notes 18.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American Journal of Physiology* 224:R601-R606.
- Craighead, J.J., and F.C. Craighead. 1969, Hawks, Owls, and Wildlife. Dover Publ. Co. New York. 443 pp.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: a Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Fitch, H.S., F. Swenson and D.F. Tillotson. 1946. Behavior and food habits of the red-tailed hawk. *Condor* 48:205-257.
- Gates, J.M. 1972. Red-tailed hawk populations and ecology in east central Wisconsin. *The Wilson Bulletin* 84:421-433.
- Godfrey, W.E. 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa.

- Janes, S.W. 1984. Influences of territory composition and interspecific competition on red-tailed hawk reproductive success. *Ecology* 65(3): 862-870.
- Lowe, C. 1978. Certain life history aspects of the red-tailed hawk, central Oklahoma and interior Alaska. Master's Thesis. University of Alaska, Fairbanks.
- Luttich, S., D.H. Rusch, E.C. Meslow and L.B. Keith. 1970. Ecology of red-tailed hawk predation in Alberta. *Ecology* 51(2): 190-203.
- Misztal, A. 1974. The Population and Nesting of Red-tailed Hawks and Great Horned Owls on the Wisconsin Till Plains of Western Ohio. Master's Thesis. Ohio State University, Columbus.
- Orians, G. and F. Kuhlman. 1956. Red-tailed hawk and great horned owl populations in Wisconsin. *Condor* 58:371-385.
- Palmer, R. S. 1988. Red-tailed Hawk. Pages 96-134. Cited in Handbook of North American birds, Volume 5, Part 2. Yale University Press, New Haven, Connecticut.
- Petersen, L. 1979. Ecology of Great Horned Owls and Red-tailed Hawks in Southeastern Wisconsin. Wisconsin Department of Natural Resources. Technical Bulletin No. 111.
- Preston, C.R. and R.D. Beane. 2009. Red-tailed Hawk (Buteo jamaicensis), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/052doi:10.2173/bna.52 Accessed January 2011.
- Shelton, A.D. 1971. A population study of the breeding hawks genus Buteo in Marion County, Ohio. Master's Thesis. Ohio State University, Columbus.
- Stinson, C.H. 1980. Weather-dependent foraging success and sibling aggression in red-tailed hawks in Central Washington. Condor 82(1): 76-80.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.

Peregrine Falcon (Falco peregrinus)

Characteristic	Determination	Notes (With References)
General Habitat Type	Open habitats	(Ehrlich et al. 1988; Godfrey 1986)
Specific Habitat Type	Tundra, savannah, marshes and seacoasts to high mountains and open	
	forests (Ehrlich et al. 1988; Godfrey 1986).	
Foraging Range Size (Minimum is for Breeding Season)	Area: 16.0 km ² (16 - 1,508 km ²) Linear: 1.2 km (1.2 – 80 km) coastline or river	The peregrine falcon is migratory but may breed and overwinter in Canada. Range is variable depending on prey availability (Mearns 1985). Believed to largely forage within 5 km of breeding sites or within a 78.5 km² area (Beebe 1960; Nelson 1973; Nelson 1977; Enderson and Kirven 1983; White et al. 2002). Smallest range was determined in the Canadian Arctic and Greenland (Fyfe 1969; Bromley 1988; Court et al. 1988; Folk and Moller 1988; Mattox and Seegar 1988; Swem and Ambrose 1994) while the largest was determined in Colorado (Enderson and Craig 1997). Linear range estimates were determined in consultation with Brooks 1926; White 1975; Ambrose at al. 1988; Enderson et al 1995; Mesta 1999; White et al. 2002).
Body Weight (g)	Avg: 814.5 g Male: 652 g (590 -810 g) Female: 977g (760 – 1194 g)	Considerably variable between geographic regions and throughout the year. Breeding season weights of <i>anatum</i> species given (Court et al. 1988).
Foraging Guild	Carnivore	(Ehrlich et al. 1988)
Foraging Behavior	Hunting (Aerial Pursuit)	(Ehrlich et al. 1988)
Diet Proportions		
Birds	85%	Dietary composition is greatly variable depending on
Mammals	10%	location, habitat and season, and even between
Other (Fish, Insects)	5%	different birds (White et al. 2002). A comprehensive review of diet studies determined that birds are the most common prey species (Sherrod 1978; Hunter et
		al. 1988; Paine et al. 1990; White et al. 2002). Based on frequency of occurrence, birds are consumed between 77% and 99% of the time with secondary

Characteristic	Determination	Notes (With References)
		prey sources including mammals and rarely
		amphibians, fish and insects (Sherrod 1978). In
		North America, 429 different species of birds have
		been recorded as prey, while 10 bat species and 13
		other mammals were reported (White et al. 2002). A
		study by Bradley and Oliphant (1991) found
		unusually high proportions of small mammal prey in
		the diet of Arctic falcons during years of high small
		mammal abundance.
		This is based on the allometric equation (Nagy 1987
	0.06 kg dry	in US EPA 1993) for total dry food intake for all
Food Ingestion Rate	food/kg wet	birds (g/day;(0.648*BW ^{0.651}) divided by BW. The
	BW/day	average BW provided in this table was used for
		these calculations.
		This is based on the allometric equation (Calder and
	0.06 I /lea wat	Braun 1983 in US EPA 1993) for total water intake
Water Ingestion Rate	0.06 L/kg wet	for all birds (L/day; (0.059*BW ^{0.67}) divided by BW.
	BW/day	The average BW provided in this table was used for
		these calculations.
Incidental Soil or		
Sediment Ingestion	None identified	Refer to Section 2.1.8.
Rate		

Peregrine Falcon References

- Ambrose, R.E., R.J. Ritchie, C.M. White, P.F. Schempf, T. Swem and R. Dittrick. 1988. Changes in Status of Peregrine Falcon populations in Alaska. Pages 73-82 in Peregrine Falcon Populations: Their Management and Recovery. (Cade, T.J., J.H. Enderson, C.G. Thelander and C. M. White, Eds.) The Peregrine Fund, Inc. Boise, ID.
- Beebe, F.L. 1960. The marine peregrines of the northwest Pacific coast. *Condor* (62):154-189.
- Bradley, M. and L.W. Oliphant. 1991. The diet of peregrine falcons in Rankin Inlet, Northwest Territories: An unusually high proportion of mammalian prey. Condor 93(1): 193-197.
- Bromley, R.G. 1988. Status of Peregrine Falcons in the Kitikmeot, Baffin, and Keewatin Regions, Northwest Territories, 1982-1985. Pages 51-57 in Peregrine Falcon Populations: Their Management and Recovery. (Cade, T.J., J.H. Enderson, C.G. Thelander and C.M. White, Eds.) The Peregrine Fund, Inc. Boise, ID.
- Brooks, A. 1926. Notes on the status of the Peale falcon. Condor 28:77-79.

- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Court, G.S., D.M. Bradley, C.C. Gates and D.A. Boag. 1988. The Population Biology of Peregrine Falcons in the Keewatin District of the Northwest Territories, Canada. Pages 729-739 Cited in Peregrine Falcon Populations: Their Management and Recovery (Cade, T.J., J.H. Enderson, C.G. Thelander and C.M. White, Eds.) The Peregrine Fund Inc., Boise, Idaho.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: A Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Enderson, J.H. and M.N. Kirven. 1983. Flights of nesting peregrine falcons recorded by telemetry. *Raptor Research* 17:33-37.
- Enderson, J.H. and G.R. Craig. 1991. Wide ranging nesting by peregrine falcons (*Falco peregrinus*) determined by radiotelemetry. *Journal of Raptor Research* 31: 333-338.
- Enderson, J.H., W. Heinrich, L. Kiff, and C.M. White. 1995. Population changes in North American peregrines. Transactions of the 60th North American Wildlife and Natural Resources Conference.
- Falk, K. and S. Møller. 1988. Status of the Peregrine Falcon in South Greenland: Population Density and Reproduction. Pages 37-43 in Peregrine Falcon Populations: Their Management and Recovery. (Cade, T.J., J.H. Enderson, C.G. Thelander and C.M. White, Eds.) The Peregrine Fund, Inc. Boise, ID.
- Fyfe, R. 1969. The Peregrine Falcon in Northern Canada. Pages 101-114 in Peregrine Falcon Populations: Their Biology and Decline. (Hickey, J. J., Ed.) University of Wisconsin Press, Madison.
- Godfrey, W.E., 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa
- Hunter, R.E., J.A. Crawford and R.E. Ambrose. 1988. Prey selection by peregrine falcons during the nestling stage. *The Journal of Wildlife Management* 52(4): 730-736.
- Mattox, W.G. and W.S. Seegar. 1988. The Greenland Peregrine Falcon Survey, 1972-1985, with Emphasis on Recent Population Status. Pages 27-36 in Peregrine Falcon Populations: Their Management and Recovery. (Cade, T.J., J.H. Enderson, C.G. Thelander and C.M. White, Eds.) The Peregrine Fund, Inc. Boise, ID.
- Mearns, R. 1985. The hunting ranges of two female peregrines toward the end of a breeding season. *Raptor Rescue* 19:20-26.
- Mesta, R. 1999. Endangered and threatened wildlife and plants; final rule to remove the American peregrine falcon from the federal list of endangered and threatened wildlife, and to remove the similarity of appearance provision for free-flying peregrines in the coterminous United States. Federal Registry 64(164):46542-46558.
- Nagy, K. A. 1987. Field metabolic rate and food requirement scaling in mammals and birds. *Ecological Monographs* 57:111-128. Vancouver, Canada

- Nelson, R. W. 1977. Behavioral ecology of coastal peregrines (Falco peregrinus pealei). Phd Thesis. University of Calgary, Calgary, AB.
- Nelson, R.W. and J.A. Campbell. 1973. Breeding and behavior of captive Arctic peregrines. *Hawk Chalk* (12):39-54.
- Paine, R.T., J.T. Wootton and P.D. Boersma. 1990. Direct and indirect effects of peregrine falcon predation on seabird abundance. *The Auk* 107(1): 1-9.
- Sherrod, S.K. 1978. Diets of North American Falconiformes. Raptor Rescue (12): 49-121.
- Swem, T. and R.E. Ambrose. 1994. Removal of Arctic peregrine falcon from the list of endangered and threatened wildlife. Federal Registry 59(192):50796-50805
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- White, C.M. 1975. Studies on peregrine falcons in the Aleutian Islands. *Raptor Research Report* 333-350.
- White, C.M., N.J. Clum, T.J. Cade and W.G. Hunt. 2002. Peregrine Falcon (*Falco peregrinus*), The Birds of North America Online (A. Poole, Edits.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/660.

American Robin (Turdus migratorius)

Characteristic	Determination	Notes (With References)
General Habitat Type	Open habitat;	(Godfrey 1986)
General Habitat Type	generalist	(Godiney 1700)
	Forest edges, woodlands, gardens, parks, country farmland, open fields,	
Specific Habitat Type		rests and riparian areas. Less common in heavily
	forested areas (Eh	rlich et al. 1988; Godfrey 1986).
		The American robin is migratory but may breed and
		overwinter in Canada. Small foraging range size
Foraging Range Size		found in deciduous forest habitats (Weatherhead and
(Minimum is for	<u>Area</u> : 0. 7 ha	McRae 1990). Knupp at al. (1977) found that
Breeding Season)	(0. 7 - 28.3 ha)	American Robins use undefended feeding grounds
		up to 300 meters away from nesting site (28.3 ha
		range).
	<u>Avg</u> : 79 g	
Body Weight	Male: 77.4 g	(Wheelright 1986)
	Female: 80.6 g	
Foraging Guild	Omnivore	(Campbell et al. 1997)
Foraging Behavior	Ground Clean	(Ehrlich et al. 1988)
Diet Proportions		
Fruit	60%	Earthworms are one of the most important food
Invertebrates	40%	sources in many areas (Campbell et al. 1997), but
		fruit, berries and invertebrates can also be important,
		particularly during the migratory and winter periods
		(Beal 1915; Wheelwright 1986; Ehrlich et al. 1988;
		Sallabanks and James 1999). During the period
		before reproduction, insects make up a significant
		component of the diet, and insects are also fed to
		young. (Martin et al. 1951; Ehrlich et al. 1988).
		This rate is based on the average of two ingestion
	1.21 kg wet	rates (0.89 and 1.52 g/g day) as reported in US EPA
Food Ingestion Rate	food/kg wet	(1993) for both genders, adults and juveniles
	BW/day	(juvenile body weights were less than 5% different
		than adults). ²⁶

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 $^{^{26}}$ The US EPA 1993 reports food ingestion rates (0.89 and 1.52 g/g day) from Skorupa and Hothem 1985 and Hazelton et al. 1984.

Characteristic	Determination	Notes (With References)
Water Ingestion Rate	0.14 L/kg wet BW/day	This is based on the allometric equation (Calder and Braun 1983 in US EPA 1993) for total water intake for all birds (L/day; (0.059*BW ^{0.67}) divided by BW. The average BW provided in this table was used for these calculations.
Incidental Soil or Sediment Ingestion Rate	4% of dry food ingestion rate	This is based on a calculation by Sample and Suter (1994) for the American robin using the following assumptions: (1) assumed that soil consumption is proportional to earthworm consumption; (2) used sediment ingestion values for the American woodcock (10.4% soil ingestion, diet is 99% earthworms); and (3) assumed the robin consumes 40% earthworms. ²⁷

American Robin References

- Beal, F.E.L. 1915. Food of the Robins and Bluebirds of the United States. U.S. Department of Agriculture Bulletin 171.
- Calder, W. A., and Braun, E. J. 1983. Scaling of osmotic regulation in mammals and birds. *American. Journal of Physiology* 224:R601-R606.
- Campbell, R.W., N.K. Dawe, I. McTaggart-Cowan, J.M. Cooper, G.W. Kaiser, M.C.E. McNall and G.E.J. Smith. 1997. The Birds of British Columbia. Volume 3: Passerines, Flycatchers through Vireos. Ministry of Environment, Lands and Parks and Canadian Wildlife Service. UBC Press, Vancouver.
- Ehrlich, P.R., D.S. Dobkin and D. Wheye. 1988. The Birder's Handbook: A Field Guide to the Natural History of North American Birds. A Fireside Book by Simon & Schuster Inc., New York.
- Godfrey, W.E., 1986. The Birds of Canada, revised edition. National Museums of Canada, Ottawa.
- Hazelton, P. K., R. J. Robel, and A. D. Dayton. 1984. Preferences and influence of paired food items on energy intake of American robins and gray catbirds. *The Journal of Wildlife Management* 48(1): 198-202.
- Knupp, D.M., R.B. Owen, Jr., and J.B. Dimond. 1977. Reproductive biology of the American robin in northern Maine. *Auk* 94:80-85.

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 $^{^{27}}$ = (40/(99/10.4))

- Martin, A.C., H.S. Zim and A.L. Nelson. 1951. American Wildlife and Plants. McGraw-Hill Book Company Inc., New York, 500 p.
- Sallabanks, R. and F.C. James. 1999. American Robin (Turdus migratorius), The Birds of North America Online (A. Poole, Ed.). Ithaca: Cornell Lab of Ornithology; Retrieved from the Birds of North America Online: http://bna.birds.cornell.edu/bna/species/462.
- Sample, B.E. and G.W. Suter. 1994. Estimating exposure of terrestrial wildlife to contaminants. DRAFT for Internal Use Only. Prepared for the Environmental Sciences Division, Oak Ridge National Laboratory, Oak Ridge, Tennessee.
- Skorupa, J. P. and R. L. Hothem. 1985. Consumption of commercially-grown grapes by American robins: A field evaluation of laboratory estimates. *Journal of Field Ornithology* 56(4): 369-378.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Weatherhead, P.J. and S.B. McRae. 1990. Brood care in American robins: Implications for mixed reproductive strategies by females. *Animal Behavior* 39: 1179-1188.
- Wheelwright, N.T. 1986. The diet of American robins: An analysis of U.S. biological survey records. *The Auk* 103: 710-725.

Wood Frog (Lithobates sylvatica)

Characteristic	Determination	Notes (With References)
General Habitat Type	Wet habitats	(Matsuda et al. 2006; Corkran and Thoms 1996)
Specific Habitat Type	Forests, fields, muskegs, marshes, wet meadows, moist woodlands and brush (Matsuda et al. 2006; Corkran and Thoms 1996).	
Foraging Range Size (All Seasons)	Area: 25 ha	(Baldwin et al. 2006)
Body Weight	<u>Avg</u> : 8 g	Encyclopedia Britannica
Foraging Guild	Carnivore	(Muths et al. 2005)
Foraging Behavior	Hunting	(Muths et al. 2005)
Diet Proportions		
Insects and Spiders	70%	Diet studies in Alberta determined that adult and
Snails and Slugs	25%	juvenile wood frogs are carnivorous with their diet
Earthworms	5%	dominated by arthropods such as insects and spiders
		(Moore and Stickland 1955). Secondary diet
		components include snails and slugs and the
		occasional earthworm (Moore and Stickland 1955).
		Tadpoles are opportunistic and omnivorous feeding
		on algae, bacteria and single cell organisms, but will
		also eat amphibian eggs and hatchlings of American
		toad (Bufo americanus), gray tree frog (Hyla
		chrysocelis), pickerel frog (Rana palustris) and
		spotted salamander (Ambystoma maculata)
		(Petranka and Thomas 1995).
Food Ingestion Rate	None identified	
		Water ingestion rates were not identified for the
		wood frog. The water balance of amphibians is
		complex, as they absorb water through their skin as
Water Ingestion Rate	None identified	well as extract water from their food (US EPA
		1993). Nearly all amphibians rely on the skin for
		rehydration and are not known to drink water
		through the mouth (Wells 2007).
Incidental Soil or		
Sediment Ingestion	None identified	Refer to Section 2.1.8.
Rate		

Wood Frog References

- Baldwin, R.F., A.J.K. Calhoun and P.G. deMaynadier. 2006. Conservation planning for amphibian species with complex habitat requirements: A case study using movements and habitat selection of the Wood Frog *Rana sylvatica*. *Journal of Herpetology* 40(4): 442-453
- Corkran, C.C. and C. Thoms. 1996. Amphibians of Oregon, Washington and British Columbia. Lone Pine Publishing, Edmonton, Canada.
- Matsuda, B.M., D.M. Green and P.T. Gregory. 2006. Amphibians and Reptiles of British Columbia. Royal BC Museum Handbook, Victoria, Canada.
- Moore, J.E. and E.H. Strickland. 1955. Further notes on the food of Alberta amphibians. *American Midland Naturalist* 54(1): 253-256.
- Muths, E., S. Rittman, J. Irwin, D. Keinath and R. Scherer. 2005. Wood Frog (*Rana sylvatica*): A Technical Conservation Assessment. USDA Forest Service, Rocky Mountain Region.
- Petranka, J.W. and D.A.G. Thomas. 1995. Explosive breeding reduces egg and tadpole cannibalism in the wood frog, Rana sylvatica. *Animal Behaviour* 50(3):731-739.
- Storey, K.B. and J.M. Storey. 1984. Biochemical adaption for freezing tolerance in the wood frog, Rana sylvatica. *Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology* 155(1): 29-36.
- USEPA (United States Environmental Protection Agency). 1993. Wildlife Exposure Factors Handbook. Vol.1. EPA/600/R-93/187a. Office of Research and Development, Washington, DC.
- Wells, K.D. 2007. The Ecology and Behaviour of Amphibians. The University of Chigaco Press.
- Wood Frog. 2011. In Encyclopdia Brittanica Retrieved From: http://www.britannica.com/EBchecked/topic/647426/wood-frog. Accessed: March 2011.

Common Gartersnake (Thamnophis sirtalis)

Characteristic	Determination	Notes (With References)
General Habitat Type	Generally near aquatic habitats	(St. John 2002; Matsuda et al. 2006)
Specific Habitat Type	Habitat ranges from forest, fields and prairies and is often found near water in moist forests and meadows, vegetated riparian zones of creeks, rivers, lakes and marshes (St. John 2002; Matsuda et al. 2006).	
Foraging Range Size (All Seasons)	Avg: 1 ha	Carpenter (1952) estimated the home range to be approximately 2 acres (~ 1 ha) in size.
Body Weight	Avg: 90 g	(Whittier and Crews 1990)
Foraging Guild	Carnivore	(Matsuda et al. 2006)
Foraging Behavior	Hunting	(Matsuda et al. 2006)
Diet Proportions		
Amphibians	60%	Prey of common gartersnake is variable and habitat
Ground Insects/ Earthworms	30%	dependent, but generally includes amphibians, earthworms, small mammals and birds, freshwater
Fish	6%	fishes and leeches (White and Kolb 1974; Kephart
Other (Birds, Small Mammals)	4%	and Arnold 1982; Gregory and Nelson 1991; Matsuda et al. 2006). Diet studies conducted in Michigan determined that common gartersnakes
		earthworms and trace amounts of mammals, fish and caterpillars (Carpenter 1952). St. John (2002) noted that adult common gartersnakes prefer fish and amphibians, while juveniles primarily ate earthworms.
Food Ingestion Rate	0.03 kg wet food/kg wet BW/day	(Bessler et al. 2010) ²⁸ .
Water Ingestion Rate	None identified	
Incidental Soil or Sediment Ingestion Rate	None identified	Refer to Section 2.1.8.

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²⁸ This ingestion rate is based on a "moderate normal feeding schedule" of 5 meals per month which ranged from 12 to 19% of the snakes body weight. For the purposes of this module, the intakes for each meal over the course of the month were summed and divided by 30 days (one month). Snakes in this study were wild caught and approximately 50 g in body weight.

Common Gartersnake References

- Bessler, S.M., M.C. Stubblefield, G.R. Ultsch, and S.M. Secor. 2010. Determinants and modeling of specific dynamic action for the Common Garter Snake (*Thamnophis sirtalis*). *Canadian Journal of Zoology* 88: 808–820.
- Carpenter, C.C. 1952. Comparative ecology of the common garter snake (*Thamnophis s. sirtalis*), the ribbon snake (*Thamnophis s. sauritus*) and Butler's garter snake (*Thamnophis butleri*) in mixed populations. *Ecological Monographs* 22:235-238.
- Gregory, P.T. and K.J. Nelson. 1991. Predation on fish and inter-site variation in diet of the garter snake, *Thamnophis sirtalis*, on Vancouver Island. *Canadian Journal of Zoology* 69: 988-994 (Erratum in *Canadian Journal of Zoology* 70: 2501).
- Kephart, D.G. and S.J. Arnold. 1982. Garter snake diets in a fluctuating environment: A seven-year study. *Ecology* 63(5): 1232-1236.
- Matsuda, B.M., D.M. Green and P.T. Gregory. 2006. Amphibians and Reptiles of British Columbia. Royal BC Museum Handbook.
- St. John, A. 2002. Reptiles of the Northwest: British Columbia to California. Lone Pine Publishing, Edmonton, Canada.
- White, M. and J.A. Kolb. 1974. A preliminary study of Thamnophis near Sagehen Creek, California. *Copeia* (1): 126-136.
- Whittier, J.M. and D. Crews. 1990. Body mass and reproduction in female red-sided garter snakes (*Thamnophis sirtalis parietalis*). *Herpetologica* 46(2): 219-226

List of Acronyms

BW	Body Weight
CCME	Canadian Council of Ministers of the Environment
CEAEQ	Centre d'expertise en analyse environnementale Quebec
COC	Contaminant of Concern
ERA	Ecological Risk Assessment
FCSAP	Federal Contaminated Sites Action Plan
PBET	Physiologically-Based Extraction Procedures
ROC	Receptor of Concern
SAB	Science Advisory Board
SARA	Species at Risk Act
TRV	Toxicity Reference Value
US EPA	US Environmental Protection Agency
VEC	Valued Ecosystem Component

Glossary

Best professional judgement – The thorough application of critical judgement in professional practice, in which an experiential, reflective, self-corrective, and purposeful thinking process is applied to consider knowledge, context, evidence, methods, conceptualizations, and criteria. BPJ is a means by which a practitioner can incorporate a diverse range of information without articulating a mechanical process for processing the information.

Contaminants of Concern (COCs) – Contaminants that have been selected for evaluation in the ERA, usually based on a completed problem formulation. The process used to select COCs is not covered in this module.

Ecological Risk Assessment (ERA) – The process by which responses of natural organisms to human-induced environmental alteration is evaluated. ERA entails the application of a formal framework, analytical process, or model to estimate the effects of human actions on a natural resource and to interpret the significance of those effects in light of the uncertainties identified in each study component.

Exposure pathways – The routes of exposure from environmental media (soil, water, air or sediment) to the receptors of concern.

Feeding guild – A group of organisms that use the same ecological resource in a similar way for feeding (e.g., insectivores, granivores, detritivores, carnivores); or, a group of species that overlap significantly in their niche requirements.

Foraging range—The geographic area typically explored by an organism when feeding.

Home range – The geographic area to which an organism normally confines its activity.

Practitioner – The primary investigator in an ecological risk assessment responsible for the design, implementation, and interpretation of results. The practitioner, who may be a consultant, interacts with the responsible party for the site (client), the regulators, and other interested parties.

Receptor of Concern (ROC) – Any non-human individual organism, species, population, community, habitat or ecosystem that is potentially exposed to contaminants of potential concern and that is considered in the ERA.

Surrogate ROC – an ROC that is representative of a receptor type (e.g., a shrew may be used as a surrogate ROC for insectivorous mammal). More than one surrogate ROC may be used to represent a particular receptor type.

Threshold – Dividing line (in units of exposure concentration or dose) between a zone of potential response and a zone of negligible response. Thresholds may be estimated using theory, data, or a combination of both. In nature, thresholds generally do not occur as precise or static entities, due to the variations among individuals and environmental factors that influence responses. Therefore, a threshold is usually expressed as a best estimate considered protective of most of the population, and often includes a margin of safety in the derivation.

Toxicity Reference Value (TRV) – An exposure concentration or dose that is not expected to cause an unacceptable level of effect in receptor(s) exposed to the contaminant of potential concern. A TRV is a specific type of *threshold*, as defined above.

Value ecosystem component (VEC) – for purposes of ERA, this term should be considered synonymous with receptor of concern (ROC). The term VEC originates in environmental assessment literature rather than early ERA literature. Either term can be used by practitioners, but ROC is used exclusively in this guidance document.

Wildlife – In the context of ERA, the term is generally applied to birds and mammals, and sometimes defined to include reptiles and amphibians. Generally it excludes fish and invertebrates.

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