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ENVIRONMENTAL QUALITY

ARBOVIRUSES AND HUMAN HEALTH IN CANADA

Ву

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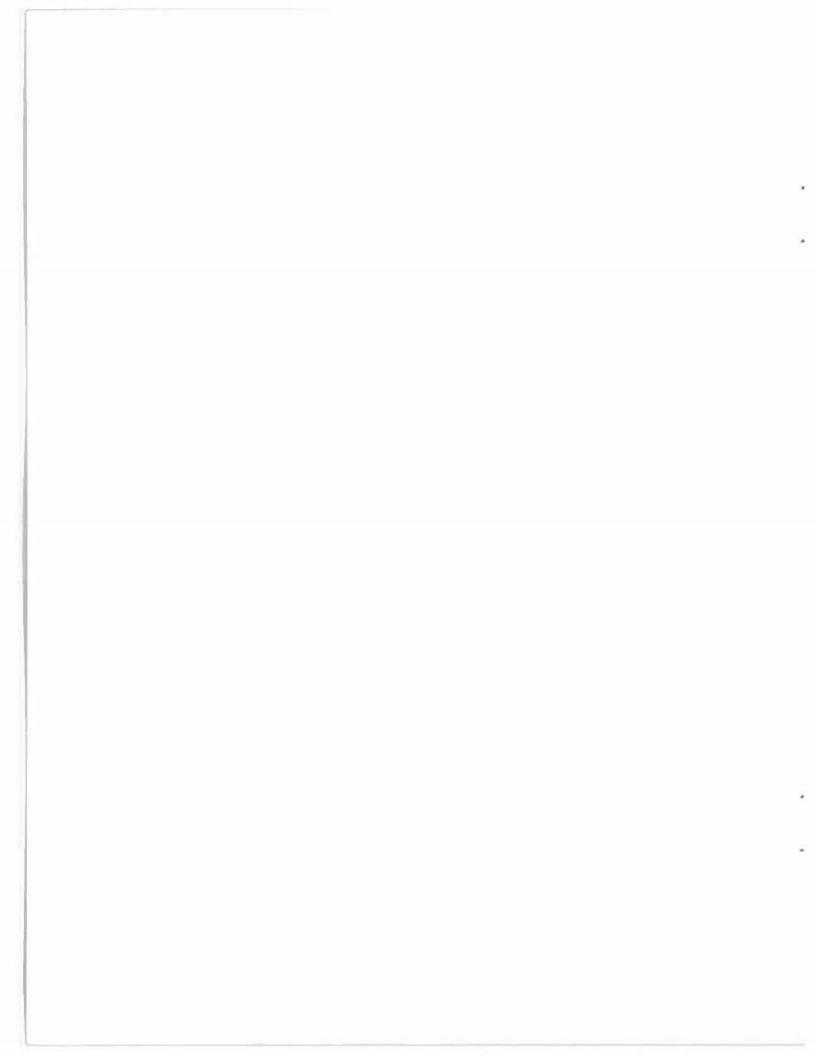
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FOREWORD

This monograph was prepared by Dr. D.M. McLean, Faculty of Medicine, Division of Medical Microbiology, University of British Columbia, Vancouver, B.C., at the request of the Biological Subcommittee of the National Research Council's Associate Committee on Scientific Criteria for Environmental Quality. The manuscript was critically reviewed and approved for publication by the Subcommittees appropriate to the subject and the Associate Committee.



ARBOVIRUSES AND HUMAN HEALTH IN CANADA

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INTRODUCTION

Arthropod-borne viruses (arboviruses) constitute a recurrent hazard to human well-being throughout Canada during the warmer months of each year, at times of high prevalence of two categories of blood-sucking arthropods - culicine and aedine mosquitoes and ixodid ticks. Despite the low incidence of clinically manifest illnesses, principally encephalitis, which are due to arbovirus infections, the severity of symptoms attracts widespread public attention. Control of the spread of arbovirus infections has been promoted by knowledge of dose/effect relationships between arboviruses and (i) their arthropod vectors on the one hand; and (ii) human and other vertebrate reservoirs on the other hand. Data applicable to the Canadian environment relating to the spread of arboviruses by mosquitoes and ticks (paragraph 4.1.3 of ACSCEQ Report No. 1) have been compiled in the present report.

The human illness which results from arbovirus infections contracted within Canada usually involves the central nervous system. Within North America, the four arboviruses associated regularly with encephalitis or aseptic meningitis are EASTERN EQUINE ENCEPHALOMYELITIS (EEE), WESTERN EQUINE ENCEPHALOMYELITIS (WEE), ST. LOUIS ENCEPHALITIS (SLE) and CALIFORNIA ENCEPHALITIS (CE) (McGowan et al. 1973). Occasional cases of encephalitis have developed following infection with POWASSAN (POW) and VENEZUELAN EQUINE ENCEPHALOMYELITIS (VEE) viruses. One North American arbovirus, COLORADO TICK FEVER (CTF) virus, usually evokes severe fever without symptoms referable to the central nervous system. Finally, no illness has been attributed to infections in Canada with arboviruses of the BUNYAMWERA group, or to TURLOCK and FLANDERS-HART PARK viruses. Within adjacent tropical areas of the Caribbean

and Central and South America and the Pacific Islands and Southeast Asia, however, additional arboviruses pathogenic for man are endemic. These include: dengue, which may induce high fever accompanied by maculopapular rash plus severe aches within the back and limbs; yellow fever, which may induce jaundice plus 'black vomit' through induction of mid-zonal necrosis of the liver; and several other agents which may induce undifferentiated fevers.

"Arboviruses are viruses which are maintained in nature principally, or to an important extent, through biological transmission between susceptible vertebrate hosts by hematophagous arthropods; they multiply and produce viremia in the vertebrates, multiply in the tissues of arthropods, and are passed on to new vertebrates by the bites of arthropods after a period of extrinsic incubation" (WHO 1967). Currently, 359 arbovirus prototype strains have been catalogued (Berge 1975). All share certain biological properties such as: ability to replicate in brains of suckling mice with production of fatal encephalitis; loss of infectivity following treatment with sodium deoxycholate or diethyl ether, which demonstrates the presence of an outer coat on the virus particle; and the presence of ribonucleic acid as the only form of nucleic acid (McLean 1968). Diameters of these enveloped virus particles range from 20 to 100 nanometres (nm). Although most serotypes exhibit cubic symmetry (TOGAVIRUSES), with a particle size of 20 to 50 nm, a substantial number exhibit helical symmetry (BUNYAVIRUSES) with a particle size of 80 to 100 nm, whilst others show a bullet-shaped internal structure (RHABDOVIRUSES) in electron micrographs (Pereira and Andrewes 1972). On the basis of hemagglutination inhibition and complement fixation tests, the arboviruses are subdivided into approximately 40 serological groups (Berge 1975). Within each serological group, individual arbovirus serotypes are identified by mouse neutralization, plague reduction neutralization, or immunodiffusion tests.

PREVALENCE OF ARBOVIRUSES IN CANADA

Prevalence of one or more serotypes of arbovirus has been established in Canada within all six Provinces and two Territories west of the St. Lawrence River (Fig. 1), by the isolation of virus from wild-caught unengorged mosquitoes or ticks on the one hand, or from blood or tissues of naturally infected vertebrates on the other hand, or from both arthropods and vertebrates (Table 1). To date, arbovirus foci have not been detected in the four Atlantic Provinces, but limited serological surveys suggest their presence. Among these arboviruses, only two (POW and WEE) have induced clinical illness in human residents of Canada, and subclinical infections have been caused by two additional serotypes (CE and CTF).

HUMAN INFECTIONS

WESTERN EQUINE ENCEPHALOMYELITIS (WEE) virus has caused both widespread outbreaks and sporadic cases of acute encephalitis amongst human residents of the Prairie Provinces during many summers since 1941. In Manitoba, between 1941 and 1949, the case fatality rate ranged from 0.7% to 41%, with 15% of 509 cases reported in 1941 terminating fatally (Adamson et al. 1950). In Saskatchewan during 1965, among 490 patients hospitalized with acute encephalitis, WEE infection was confirmed in 72 of whom 8 (11%) died (Rozdilsky $et\ \alpha l$. 1968). In 1971, for the first time, serologically confirmed infections with WEE virus were recorded in three residents of the Thompson-Okanagan region of British Columbia (Kettyls et αl . 1972). Subclinical infections, as determined by antibody production in selected population groups without overt encephalitis, involved 0.3% of British Columbia residents (ibid) and 9% of Alberta inhabitants (Iversen et al. 1971). During various epidemics, the subclinical infection rate in Manitoba ranged from 3 to 19% (Bowman 1947).

CALIFORNIA ENCEPHALITIS (CE) virus, SNOWSHOE HARE (SSH) subtype, has induced subclinical infections in 2.5% of British Columbia residents (Kettyls $et\ al.$ 1972) and in 30% of members of a farming community in northern Alberta (Iversen $et\ al.$ 1971), but no cases of

Fig. - | Arbovirus prevalence in Canadian vegetation zones.

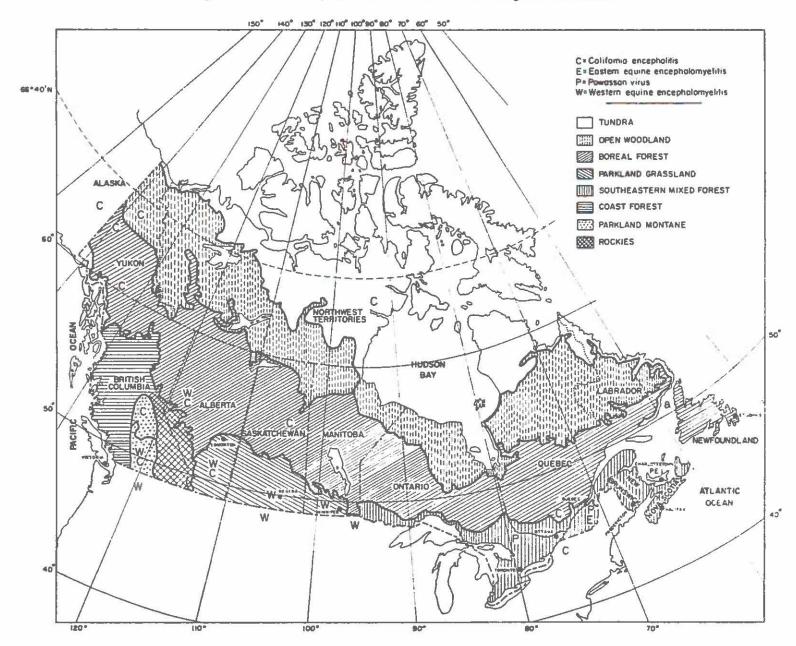


Table 1. Prevalence rates of arbovirus infections in Canadian provinces.

	Arbovirus serotype	Virus isolation rates			Antibody incidence		
Province or Territory		Vertebrate	Mosquito	Tick	Human	Vertebrate	
British Columbia	California encephalitis 1969-1970		Aedes canadensis 1:1300 (a) [†]		subclinical 48:1936, 2.5% (c)	Lepus americanus 20:31, 64.5% (b)	
						Marmota flaviven- tis 15:252, 6.0% (b)	
						Citellus columbi- anus 1:146, 0.7% (b)	
	1973		Aedes fitchii 1:8 (d)				
	Western equine encephalomyelitis	equine 2:60 (c)			clinical 3 subclinical 6:1936, 0.6% (c)		
	Colorado tick fever 1965-1966			Dermacentor andersoni 8:670 (e)	subclinical 2:1936, 0.1% (c)		
Yukon Territory	California encephalitis 1971-1974		Aedes canadensis 1:496 (f,g)			Lepus americanus 430:1076, 40% (f)	
			Aedes cinereus 1:1179 (f)			Citellus undulatus 266:3610, 7% (f)	
			Aedes communis 1:1765 (f)				
			Culiseta inor- nata 1:1778 (f)				

Table 1. (Cont'd)

Province or Territory Alberta		Vir	us isolation rates	Antibody incidence		
	Arbovirus serotype	Vertebrate	Mosquito	Tick	Human	Vertebrate
	California encephalitis 1964-1968	Lepus americanus 1 (j)	Aedes communis 1:2000 (h)		subclinical 51:160, 32% (c)	Lepus americanus 111:216, 52% (i)
		sentinel rabbits 3:36 (j)	Aedes stimulans 1:3032 (h)			sentinel rabbit 3:14, 21% (i)
	Bunyamwera group 1965		Mixed species 1:3500 approx. (k)			
	Silverwater 1965	Lepus ameri- canus 2 (j)		Haemaphysalis leporis-palus- tris 15 (j)		Lepus americanus 0-68% (j)
	Turlock 1965		Culiseta inor- nata 1:3500 approx. (k)			
	Western equine* encephalomyelitis 1965		Culex tarsalis 1:3500 approx. (k)		subclinical 20:180, 11% (i)	Lepus americanus 44:232, 19% (i)
Saskatchewan	California encephalitis 1972		Aedes cataphylla 1:313 (1)			
			Aedes excrucians 1:201 (1)			
			Aedes fitchii 1:3072 (1)			
			Aedes punctor 1:238 (1)			

Table 1. (Cont'd)

	Arbovirus serotype Western equine* encephalomyelitis 1963-1965	٧	irus isolation rates		Antibody incidence		
Province or Territory		Vertebrate	Mosquito	Tick	Human	Vertebrate	
		birds 12:480 (n)	Aedes campestris 1:988 (m)		clinical 3 1963 (only) (n)		
		horses 47:279 (n)	Aedes dorsalis 1:766 (m)				
		(1963 only)	Aedes flavescens 1:3852 (m)				
			Aedes spencerii 1:2088 (m)				
			Aedes vexans 1:1477 (m)				
			Culex tarsalis 1:180 (m)				
			Culiseta inor- nata 1:2810 (m)				
	Cache Valley 1971		Culiseta inor- nata 1:4490 (o)				
	Flanders-Hart Park 1967		Culex tarsalis (p Aedes flavescens) (p)			
	St. Louis encephalitis 1971		Culex tarsalis 1:1004 (o)				

Table 1. (Cont'd)

	Arbovirus serotype	Vi	rus isolation rate	Antibody incidence		
Province or Territory		Vertebrate	Mosquito	Tick	Human	Vertebrate
Manitoba	Western equine* encephalomyelitis 1941-1947	human fatality 8-41% (q)	Culex tarsalis		subclinical 3-19% (q)	
Ontario	California encepha- litis 1963, 1965	sentinel rab- bits 5:9 (r)				sentinel rabbits 20:29, 69% (r)
						Lepus americanus 9:107, 8% (s)
	Powassan* 1958	human 1 (t)			subclinical 6:180, 3% (u)	
	1962			Ixodes marxi 1:14 pools (w)		
	1964-1966	Marmota monax 1:497 (v)		Ixodes cookei 1:15 pools (v)		Marmota monax 437:993, 44% (t)
						Tamiasciurus hudso- nius 20:109, 18% (v
	Silverwater 1960			Haemaphysalis leporis-palus- tris 1:24 (x)		Lepus americanus 19:211, 9% (x)
	1962			Haemaphysalis leporis-palus- tris 1:21 (w)		
	1963			Haemaphysalis leporis-palus- tris 1:57 pools (v)		Lepus americanus 21:107, 20% (s)

Table 1. Cont'd)

		V	irus isolation	rates		Antibo	dy incidence	
Province or Territory	Arbovirus serotype	Vertebrate	Mosquito	Tick		ın	Vertebrate	
)uebec	Eastern equine encephalomyeliti 1972	equine 5 (z) s					equine 5:29, 17% (y)	
	Powassan 1972				clir 1 (2	iical :)		
Northwest Territories	California encephalitis 1973		Aedes hexod 4:4757 (aa)				Lepus americanus 8:29, 27% (bb)	
These viruse	s have been isolate	d from brains of fata	l human cases	in the state	d Province.			
Legend to re	ferences.							
(a) McLean et	al. 1970 (h) Iversen et al. 196	9	(o) Burto	n et al. 1973	(v)	McLean et al. 1967	
(b) McLean et	al. 1971 (i) Iversen et al. 197	I	(p) Hall	et al. 1969	(w)	McLean et al. 1963	
(c) Kettyls et	al. 1972 (j) Yuill et al. 1969		(q) Adams	on et al. 1950	(x)	McLean et al. 1961	
(d) McLean et	al. 1974 (k) Hall <i>et al</i> . 1968b		(r) McKie	1 et al. 1966	(y)	Bellavance et al. 197	
e) Hall et al	. 1968b (1) Iversen et al. 197	3	(s) McLea	n et al. 1964	(z)	Rossier et al. 1974	
(f) McLean et	al. 1975a (m) McLintock et al. 1	970	(t) McLea	n and Donohue 195	59 (aa)	Wagner et al. 1975	
(g) McLean et	al. 1972 (n) Burton et al. 1966		(u) McLea	n et al. 1960	(bb)	Gaunt et al. 1974	

encephalitis due to CE virus have been reported to date in Canada.

Clinical cases of encephalitis due to POWASSAN (POW) virus occurred in a child resident of Ontario in 1958, who subsequently died (McLean and Donohue 1959), and in a Quebec pediatric patient during 1972 who made a slow recovery (Rossier et αl . 1974). In Ontario, 3% of human residents developed POW antibody in the absence of encephalitis (McLean et αl . 1960).

COLORADO TICK FEVER (CTF) virus has shown a subclinical infection rate of 0.1% in British Columbia residents (Kettyls $et\ \alpha l$. 1972), but no virologically-confirmed clinical cases of fever due to this agent have yet been documented in Canada.

ST. LOUIS ENCEPHALITIS (SLE) virus has not yet been associated with clinical or subclinical infection in residents of Canada, a despite its continuing prevalence as a cause of encephalitis in human residents of southeastern, midwestern and western USA and its recent isolation from mosquitoes in Saskatchewan (Burton et al. 1973). Similarly, EASTERN EQUINE ENCEPHALOMYELITIS (EEE) virus has not yet been associated with human illness in Canada.

NON-HUMAN VERTEBRATES

Infections of non-human vertebrates with WEE virus occurred in British Columbia during 1971 when brains of two of 60 horses which were kept in the Thompson-Okanagan region (51°N), yielded WEE virus (Kettyls et al. 1972). In Alberta, 44 of 232 snowshoe hares collected at 54°N during 1965 had antibody (Iversen et al. 1971). In Saskatchewan during the summer of 1963, 47 of 279 horses with encephalitis died, and WEE virus was isolated from blood or tissues of 12 of 480 nestling wild birds of 20 species, principally those which frequent sloughs (Burton et al. 1966).

In the eastern townships of Quebec, during the summer of 1972, serological evidence of EEE virus infection was detected in five horses which developed encephalitis among 29 tested, and the brain of one fatal case yielded EEE virus (Bellavance $et\ al.\ 1973$).

 $^{^{\}alpha}$ See Addendum on page 30.

CALIFORNIA ENCEPHALITIS virus infections have been demonstrated serologically in 20 of 31 snowshoe hares and 15 of 252 marmots (Marmota flaviventris) collected in British Columbia during 1970 near Penticton (49秒°N, 120°W) (McLean et al. 1971). In 1971, between Williams Lake (52°N) and Prince George (54°N), 39 of 78 snowshoe hares (Lepus americanus) and 1 of 35 Columbian ground squirrels (Citellus columbianus) showed CE neutralizing antibodies (McLean et al. 1972). In the Yukon Territory, between latitudes 61 and 66°N, 430 of 1076 L. americanus and 266 of 3610 Arctic ground squirrels (Citellus undulatus) showed CE neutralizing antibodies between the summers of 1971 and 1974 (McLean et αl . 1975a). Near Rochester, Alberta (54°N, 113°W), between 1964 and 1968, CE virus (snowshoe hare subtype) was isolated from the blood of one L. americanus plus 3 of 63 sentinel rabbits. CE antibodies were found in 111 of 216 wild-caught L. americanus and antibody conversions in 3 of 14 sentinel domestic rabbits (Yuill et al. 1969). Near Ottawa, Ontario (45°N, 76°W), during 1963 and 1965, CE virus was isolated from the blood of 5 of 9 sentinel rabbits, and another 20 of 29 sentinel rabbits showed CE antibody conversions (McKiel et al. 1966). In 1963, near North Bay, Ontario, (46°N, 79°W), sera from 9 of 107 L. americanus showed CE antibody (McLean et αl . 1964).

POWASSAN virus was isolated from blood of 2 of 993 groundhogs (Marmota monax) collected near North Bay, Ontario, between 1964 and 1966, and POW neutralizing antibody was detected in 437 of 993 M. monax plus 20 of 109 red squirrels (Tamiasciurus hudsonicus) at the same location (McLean et al. 1967).

SILVERWATER (SIL) virus, which has not yet been incriminated as a human pathogen, was isolated from the blood of two L. americanus collected near Rochester, Alberta during 1965, and 0 to 68% L. americanus collected during each summer between 1964 and 1968 had antibody (Yuill et al. 1969). Near North Bay, Ontario, in 1963, sera from 21 of 107 L. americanus showed SIL complement fixing antibody (McLean et al. 1964), whilst on Manitoulin Island (46°N, 83°W), 19 of 211 L. americanus sera contained SIL antibody during the summers of 1960 and 1961 (McLean et al. 1961).

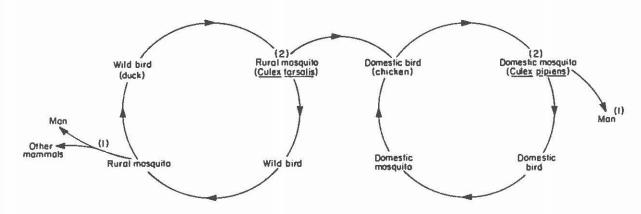
ARTHROPODS

(i) MOSQUITO VECTORS

The principal mosquito vector of WEE virus in the Prairie Provinces during successive summers has been Culex tarsalis (Fig. 2). In Saskatchewan, during 1963-1965, the minimum field infection rate (MFIR = number of pools of wild-caught mosquitoes which yielded virus. divided by the total number of wild-caught mosquitoes processed in pools for virus isolation) for Culex tarsalis was 1:180 (McLintock et αl . 1970). This was substantially lower than the MFIR for five Aedes species which ranged from 1:766 for A. dorsalis to 1:3852 for A. flavescens and 1:2810 for Culiseta inornata (ibid.) (Fig. 3). The high MFIR for Culex tarsalis, the repeated virus isolations during successive summers, the readiness with which it feeds on both wild and domestic birds despite ingestion of earlier blood meals, its readiness to bite man, and the isolation of virus from wild-caught mosquitoes as early as 22 June until 15 August, both before and during the occurrence of human and horse cases of encephalitis, provide adequate criteria in support of its role as the principal natural vector species. Culex tarsalis was found as far north as 53°N (Burton and McLintock 1970), which was the northern limit of prevalence of cases of encephalitis. Although virus was isolated from Culiseta inornata less frequently, this species yielded virus between 20 and 24 September, which was the latest time of year at which virus-infected mosquitoes were collected. Both Culex tarsalis and Culiseta inornata overwinter as adults, thus providing a means by which mosquitoes infected during late summer may maintain virus until extensive activity of vertebrates and arthropods commences the following spring. There is no field or laboratory evidence of transovarial transfer of WEE to date. Low MFIR in Aedes mosquitoes, together with the relative infrequency with which they imbibe more than one vertebrate blood meal, render the role of these five species as natural vectors unlikely.

In Alberta and Manitoba, *Culex tarsalis* has been incriminated repeatedly as the principal vector of WEE virus. No details are available regarding WEE vectors in British Columbia.

Figure 2

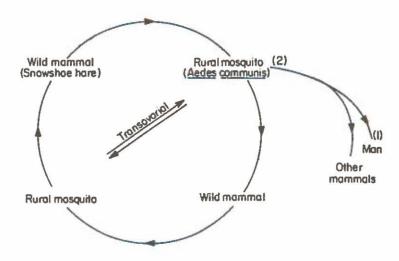


Bird-mosquito cycle (e.g. western equine encephalitis)

No evidence of transovarial transfer.

Transmission of infection to man may be prevented by:
(1) repellants and protective clothing
(2) mosquito abatement (larval and adult)

Figure 3



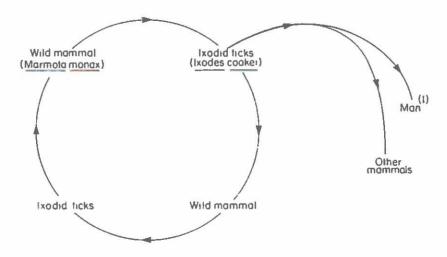
Mammal-mosquito cycle (e.g. California encephalitis)

Transovarial transfer is indicated. Birds are insusceptible hosts
Transmission of infection toman may be prevented by:
(1) repellants and protective clothing
(2) mosquito abatement (larval and adult)

The principal mosquito vectors of CE virus in the boreal forest regions of British Columbia (McLean et al. 1974), Alberta (Iversen et al. 1969), Saskatchewan (Iversen et αl . 1973), the Yukon (McLean et αl . 1975a) and the Northwest Territories (Wagner et al. 1975), comprise several species of Aedes and Culiseta. These mosquitoes also serve as vectors in irrigated farmlands of southcentral British Columbia (McLean et αl . 1970) and southeastern Alberta (Morgante and Semanchuk 1967). Aedes canadensis and A. communis have shown the highest MFIR, and repeated isolations during successive summers point towards these species as principal vectors, although seven other Aedes species caught in nature have yielded virus (Rozdilsky et al. 1968) (Fig. 3). Virus replication has been demonstrated in salivary glands of A. canadensis, cinereus, communis and hexodontus following intrathoracic injection and incubation at 21° and 13°C (McLean et al. 1975a); A. cinereus transmitted virus by biting suckling mice 15 days after intrathoracic injection following incubation at 13°C (McLean et al. 1974). The isolation of CE virus from Aedes larvae collected at Kusawa Lake (61°N, 136°W), Yukon Territory, on 16 May 1974 before the springtime emergence of adult mosquitoes at that location (McLean et al. 1975a), strongly suggests that this agent has overwintered by transovarial transmission. Culiseta inornata showed MFIR comparable to those for Aedes mosquitoes in the Yukon. This species has transmitted CE virus 15 days after intrathoracic injection following incubation at 28° C (McLean et αl . 1974); infectivity has been demonstrated in salivary glands of mosquitoes held continuously for 138 days at 0°C and from day 77 to day 194 at -1°C, following intrathoracic injection (McLean et al. 1975b). In the Yukon, Culiseta inornata emerges during early May, along with some Aedes species, and has been collected until mid-June, thus permitting ample opportunity for one or more blood meals from vertebrates. Daytime temperatures of 10 to 15°C would be sufficient to permit prompt viral replication in mosquito salivary glands, following which the virus could overwinter within the adult mosquitoes.

BUNYAMWERA GROUP viruses (CACHE VALLEY) have been isolated from Aedes mosquitoes of mixed species collected in central Alberta near Coronation (52°N, 111°W) during August 1965 (Hall et αI . 1968a), when the MFIR was

Figure 4



Mammal-tick cycle (e.g. Powassan virus)

No evidence of transovarial transfer Transmission of infection to man may be prevented by (1) repellants and protective clothing

approximately 1:3500, and from Culiseta inornata collected near Weyburn, Saskatchewan (50°N, 105°W) during July 1971 (Burton et al. 1973). Both of these BUNYAMWERA GROUP isolates in Canada were achieved from mosquitoes at prairie locations where horses have contracted encephalitis due to WEE virus repeatedly during summer, but to date there is no evidence that encephalitis due to these viruses has affected human residents and horses. To date, these are the only two isolations of BUNYAMWERA GROUP viruses from mosquitoes in Canada but another BUNYAMWERA GROUP agent, NORTHWAY (NOR) virus, was isolated in June 1970 from A. hexodontus mosquitoes near Northway, Alaska (63°N, 142°W) (Calisher et al. 1974), where CE virus was also isolated from mosquitoes (Ritter and Feltz 1974). This is 150 miles southwest of Dawson City (64°N, 139°W), Y.T., where CE virus has been isolated from mosquitoes (McLean et al. 1975a).

TURLOCK virus was recovered from Culiseta inornata mosquitoes collected near Coronation, Alberta
(52°N, 111°W) during August 1965 (Hall et al. 1968a).
Both TURLOCK and mosquito-borne arboviruses of the
FLANDERS-HART PARK group have also been recovered on
several occasions from mosquitoes collected in southeastern Saskatchewan (J. McLintock, personal communication).

(ii) TICK VECTORS

COLORADO TICK FEVER virus has been recovered from 8 of 22 tick pools comprising 670 Dermacentor andersoni ticks which were collected in forested mountain terrain in southeastern British Columbia mainly near Salmo (49°N, 118°W) during the spring of 1965 and 1966 (Hall et al. 1968b). These comprise the northernmost isolations of CTF virus from D. andersoni in locations adjacent to their principal areas of prevalence in the States of Washington, Idaho and Montana.

Tick vectors of POWASSAN virus have been identified only in Ontario near North Bay $(46\,^\circ\text{N})$ where 18 of 273 pools of $Ixodes\ cookei$ collected between 1964 and 1966 (McLean et al. 1964) and 1 of 14 pools of $I.\ marxi$ collected in 1962 (McLean and Larke 1963) (containing 1 to 9 ticks per pool) have yielded virus. These ticks feed

principally on marmots and squirrels respectively, but both species will bite man. Although *D. andersoni* ticks collected in southcentral British Columbia have not yielded POW virus, laboratory-infected larval and nymphal ticks have transferred virus transstadially, and hamsters and mice have been infected by bites of infected ticks (Chernesky and McLean 1969).

SILVERWATER virus was first recovered from 2 of 49 pools of Haemaphysalis leporis-palustris (HLP) ticks collected on Manitoulin Island, Ontario (46°N, 83°W) during the summer of 1960 (McLean et al. 1961). Near North Bay, Ontario (46°N, 79°W), this agent was recovered from a pool of HLP ticks collected during July 1962 among 24 pools tested between July 1959 and October 1962 (McLean and Larke 1963). During May 1963, one HLP tick pool collected in the same region yielded SIL virus, among 57 tick pools tested during the summer of 1963 (McLean et al. 1964). Subsequently, SIL virus was isolated from 15 pools of HLP ticks collected from snowshoe hares near Rochester, Alberta (54°N, 113°W) during the spring and summer of 1962 to 1965 (Yuill et al. 1969).

(iii) VEGETATION AND TERRAIN

Mosquito-borne arboviruses are distributed widely throughout the grassland and boreal forest regions west of the Great Lakes. In grassland regions of the Prairie Provinces, Culex tarsalis is an abundant summertime mosquito species. The increasing use of irrigation in agriculture promotes the buildup of mosquito populations, especially Culex tarsalis. This facilitates the natural transmission of several arboviruses, but favors particularly the prevalence of WEE virus (McLintock $et\ \alpha l.$ 1970). The northern limit of distribution of C. tarsalis is 53°N in Saskatchewan (Burton and McLintock 1970), which corresponds both with the northerly extent of documented WEE infection in man and mosquitoes, and the northernmost fringe of prairie grassland. In the boreal forest, however, both near St. Walburg and in Prince Albert National Park (53°N), Aedes mosquitoes and Culiseta inornata predominate. These species are associated particularly with the transmission of CE virus, and during the summer of 1972, eight CE isolates were achieved from four Aedes species (Iversen et al.

1973). In Alberta, $Culex\ tarsalis$ which was collected at the northern limit of grassland near Stettler (52°N) has yielded WEE virus (Hall $et\ al.$ 1968a). In the boreal forest near Rochester (54°N), however, the mosquito population was composed predominantly of $Culiseta\ inornata$ and several Aedes species, which have repeatedly yielded arboviruses of the CE group (Iversen $et\ al.$ 1969).

In mountain forest regions of British Columbia. especially in the Okanagan and Shushwap valleys where fruit is grown extensively, large populations of Aedes mosquitoes have built up during early summer of each year. Extensive mosquito abatement procedures which relied upon DDT and organophosphates, until banned during the summer of 1971, reduced mosquito populations to negligible numbers by mid-July, thus eliminating opportunities for natural transmission of arboviruses. Prevalence of CE virus was first established positively in the south Okanagan region during June 1969 by the isolation of CE virus from wildcaught A. canadensis (McLean et al. 1970). During the summer of 1971, recovery of WEE virus from the brain of a horse which developed encephalitis near Salmon Arm, plus diagnostically significant WEE antibody increments in three human residents of, or visitors to, the Shushwap and Okanagan valleys who developed encephalitis (Kettyls et al. 1972), confirmed the long-suspected prevalence of this arbovirus in southeastern British Columbia.

In montane and subalpine forest regions of central British Columbia, the endemic prevalence of CE virus was firmly established in July 1973 by the isolation of CE virus from A. fitchii mosquitoes collected near Williams Lake (52°N) (McLean $et\ al$. 1974). Serological evidence of infection of small rodents has revealed infection with this agent from this point northward into boreal forest regions of northeastern British Columbia and the Yukon Territory (McLean $et\ al$. 1971, 1975a).

The boreal forest covers virtually all of the Yukon Territory south of the Arctic Circle. Repeated isolations of CE virus from Aedes mosquitoes and Culiseta inornata in or adjacent to forest habitats between 60 and 66°N during the summers of 1971 through 1974 (McLean et al. 1975a) suggest its endemic prevalence throughout the entire boreal forest region of the Yukon.

In the tundra region of the Northwest Territories near Rankin Inlet (63°N, 92°W), the prevalence of CE virus was clearly demonstrated in July 1973 by its isolation from A. hexodontus mosquitoes (Wagner et αl . 1975), a species typical of boreal forest and tundra regions.

Although the boreal forest extends eastward to the province of Newfoundland and Labrador, insufficient studies have been undertaken east of Saskatchewan to determine whether or not arboviruses are prevalent in this vegetation zone.

In southeastern mixed forest regions of Ontario, endemic foci of mosquito-borne CE virus activity have been identified at three locations at latitudes 45 to 46°N: Monitoulin Island (McLean et al. 1961), North Bay (McLean et al. 1964) and near Ottawa (McKiel et al. 1966). In each location, Aedes comprises the dominant mosquito genus within the mosquito population. Although to date no isolations of CE virus have been achieved in this vegetation zone east of Ottawa, another mosquito-borne agent, EEE virus, was identified for the first time in September 1972 as the etiological agent in an epizootic of encephalitis which affected horses in the eastern townships of Quebec (Bellavance et al. 1973).

In the southeastern mixed forest zone, foci of activity of two tick-borne arboviruses have also been identified. POWASSAN virus has been isolated from a human patient, pools both of Ixodes cookei and Ixodes marxiticks and the blood of groundhogs (Marmota monax), near North Bay (McLean and Donohue 1959; McLean et al. 1967; McLean and Larke 1963). Serological evidence of infection of a human patient was detected southeast of Montreal (Rossier et al. 1974). SILVERWATER virus foci were identified by virus isolation from Haemaphysalis leporis-palustris ticks collected on Manitoulin Island (McLean et al. 1961) and near North Bay (McLean et al. 1964).

GAPS IN KNOWLEDGE

1. Although the prevalence of mosquito-borne CE virus has been establish clearly in the boreal forest zone of the Yukon Territory, British Columbia, Alberta, and Saskatchewan, an urgent need is to determine and map its prevalence throughout the open woodland vegetation zone along the Mackenzie Valley, immediately east of the border between the Yukon and the Northwest Territories. This need has become imminent through the immediate prospect of location of several thousand workers in primitive rural campsites during construction of the proposed Mackenzie Valley pipeline, highway, and other services. Under these circumstances, a substantial human population will become exposed simultaneously to mosquito bites throughout May, June, and July of each year. Whilst the possibility of development of encephalitis in man following bites by infective mosquitoes is estimated at 1:200 to 1:500, two to five cases could be expected among every 1000 persons located in an endemic focus.

An equally urgent requirement is to define the prevalence of CE virus in open woodland and tundra regions near Inuvik and other coastal settlements on the Arctic Ocean on account of: (i) extensive oil drilling operations in Arctic Ocean coastal districts, and (ii) the isolation of CE virus along the Arctic seacoast at Beaufort Lagoon, Alaska, about 200 miles west of Inuvik.

- 2. Although CE virus was recovered from mosquitoes collected on the tundra near Rankin Inlet, N.W.T., there is a complete absence of information regarding possible arbovirus prevalence throughout tundra and transitional zones of the eastern Arctic.
- 3. Arbovirus prevalence has not yet been established in boreal forest and southeastern mixed forest zones of Quebec, Newfoundland and Labrador, Nova Scotia, New Brunswick, and Prince Edward Island. Preliminary serological evidence has suggested the prevalence of CE virus in Nova Scotia.

- 4. Intensive field investigations are required in the eastern townships of Quebec to determine quantitative host-vector relationships in foci of activity of mosquito-borne EEE and tick-borne POW viruses.
- Quantitative relationships must be determined between the incidence of infection in mosquito larvae. adult mosquitoes and animal reservoirs for CE virus in the Yukon Territory. With the decline of incidence of usual wild vertebrate hosts during 1975, it is essential to investigate whether transovarial transfer of this virus will alone ensure a sufficiently high proportion of infected mosquitoes in successive summers to maintain foci of natural infection without fresh introduction of virus into the mosquito populations by imbibing viremic blood of infected vertebrates before ovulation each summer. In Wisconsin, the serologically related CE group agent, LA CROSSE virus, has infected about 5% of emergent A. triseriatus adults derived from parent mosquitoes which imbibed infective blood 9 months earlier (Watts et al. 1974).
- 6. Despite some 30 years of intensive investigation of the prevalence of WEE virus in mosquitoes, vertebrate reservoirs and human inhabitants of prairie portions of Saskatchewan, including the provision of good laboratory diagnostic facilities for clinical infections in man, new epidemiological investigations are required whenever new tracts of prairie are converted from dry-farming to irrigation.
- 7. The influence of temperatures approaching 0°C on the replication of CE, NOR, WEE and EEE virus in natural vector species such as wild-caught Aedes spp. and Culiseta spp. from various vegetation zones in Canada has only commenced to be investigated for CE and NOR viruses, and no data exist for WEE and EEE viruses. Such knowledge may improve our ability to predict outbreaks of mosquito-borne diseases and to prevent them.

SUMMARY

Quantitative data have been assembled on the prevalence of five mosquito-borne arboviruses in Canada. including two ALPHAVIRUSES, EASTERN (EEE) and WESTERN (WEE) EQUINE ENCEPHALOMYELITIS, one FLAVIVIRUS, ST. LOUIS ENCEPHALITIS (SLE), and two BUNYAVIRUSES, the SNOWSHOE HARE (SSH) subtype of CALIFORNIA ENCEPHALITIS (CE). and a CACHE VALLEY-like agent. All of these, with the possible exception of the CACHE VALLEY agent, have induced encephalitis in man. Data have also been assembled for two human pathogenic tick-borne arboviruses, a FLAVIVIRUS, POWASSAN (POW), and an ungrouped agent, COLORADO TICK FEVER (CTF). Minimum field infection rates of arboviruses in mosquitoes, plus virus isolation rates and/or antibody prevalence rates in humans and mammalian or avian reservoirs, have been tabulated. Correlations have been developed between foci of arbovirus activity and the several vegetation zones of Canada. Gaps in knowledge of the potential hazards of arboviruses under Canadian conditions of climate, vegetation zones, and human settlement, have been identified; in particular: (i) there are vast regions where evidence of arbovirus activity has not yet been sought; and (ii) little is known about the effects of atmospheric temperature and transovarial transfer rates on the maintenance and spread of arboviruses in regions of demonstrated arbovirus endemicity.

ADDENDUM

HUMAN INFECTIONS WITH SLE AND WEE VIRUSES DURING THE SUMMER OF 1975

During the summer of 1975, clinically-manifest human infections with SLE virus were encountered for the first time in Canada. By 6 October, 44 of 191 human residents of southwestern Ontario who developed meningoence-phalitis had serological evidence of SLE infection (Canada Diseases Weekly Report 1: 92, 1975).

In Manitoba, 4 human residents who developed encephalitis during August 1975 had laboratory evidence of WEE virus infection. Horses were also affected, sentinel chickens showed rising WEE antibody titers, and WEE virus was isolated from Culex tarsalis mosquitoes following a substantial population build-up. Following commencement of extensive mosquito abatement programs in Winnipeg and environs on 15 August, the C. tarsalis population declined, and no further human cases of WEE infection were reported in the area. In Saskatchewan, although no human cases of WEE virus infection have been reported, several horse deaths have been attributed to this virus, and WEE virus has been isolated from C. tarsalis (Canada Diseases Weekly Report 1: 89, 1975).

In the United States up to 30 September 1975, a total of 541 laboratory-confirmed cases of SLE virus infection had been reported from 19 States and the District of Columbia, with the highest incidence reported in Illinois, Indiana, Ohio and Mississippi (Morbidity and Mortality Weekly Report 24: 339, 1975). To 2 September, 82 human cases of encephalitis had been reported in Minnesota and North Dakota, and 9 had serological evidence of current WEE virus infection (*ibid*. 24: 295, 1975). Thus, throughout the Mississippi River drainage basin extending northwards into southwestern Ontario, SLE virus prevalence has been high during the summer of 1975, whilst in the Red River basin extending northwards into the prairie portion of Manitoba, WEE virus has been widely prevalent.

REFERENCES

- Adamson, J.D., Bowman, M. and MacDonell, J.A. 1950. The future of western equine encephalitis. Manitoba Med. Rev. 30: 366-368.
- Bellavance, R., Rossier, E., Lemaitre, M., Willis, N.G. and Belanger, P. 1973. Eastern equine encephalomyelitis in eastern Canada 1972. Can. J. Public Health 64: 189-190.
- Berge, T.O. (editor) 1975. International catalogue of arboviruses. 2nd edition. U.S. Department of Health, Education and Welfare, Publication No. (CDC)75-8301.
- Bowman, M. 1947. Epidemiology of poliomyelitis and encephalitis (western equine). Manitoba Med. Rev. 27: 670.
- Burton, A.N. and McLintock, J.R. 1970. Further evidence of western encephalitis infection in Saskatchewan mammals and birds and in reindeer in northern Canada. Can. Vet. J. 11: 232-235.
- Burton, A.N., McLintock, J.R. and Francy, D.B. 1973. Isolation of St. Louis encephalitis and Cache Valley viruses from Saskatchewan mosquitoes. Can. J. Public Health 64: 368-373.
- Burton, A.N., McLintock, J.R., Spalatin, J. and Rempel, J.G. 1966. Western equine encephalitis in Saskatchewan birds and mammals 1962-1963. Can. J. Microbiol. 12: 132-141.
- Calisher, C.H., Lindsey, H.S., Ritter, D.G. and Sommerman, K.M. 1974. Northway virus: a new Bunyamwera group arbovirus from Alaska. Can. J. Microbiol. 20: 219-223.
- Chernesky, M.A. and McLean, D.M. 1969. Localization of Powassan virus in *Dermacentor andersoni* ticks by immunofluorescence. Can. J. Microbiol. 15: 1399-1408.

- Gaunt, R.A., Stowe, T.C. and Watson, C.G. 1974. Antibody to human pathogens in the wildlife of the Yellowknife, N.W.T. area. Can. J. Public Health 65: 61 (Abstract only).
- Hall, R.R., McKiel, J.A. and Brown, J.H. 1968a. Isolation of Turlock virus and a member of the Bunyamwera group, probably Cache Valley virus, from Alberta mosquitoes. Can. J. Public Health 59: 159-160.
- Hall, R.R., McKiel, J.A. and Gregson, J.D. 1968b. Occurrence of Colorado tick fever virus in *Dermacentor* andersoni ticks in British Columbia. Can. J. Public Health 59: 273-275.
- Hall, R.R., McKiel, J.A., McLintock, J. and Burton, A.N.
 1969. Arboviruses from Saskatchewan mosquitoes
 isolation of a member of the Flanders-Hart
 Park group and of a strain as yet unidentified.
 Can. J. Public Health 60: 486-488.
- Iversen, J.O., Hanson, R.P., Papadopoulos, O., Morris, C.V. and De Foliart, G.R. 1969. Isolation of viruses of the California encephalitis virus group from boreal *Aedes* mosquitoes. Am. J. Trop. Med. Hyg. 18: 735-742.
- Iversen, J.O., Seawright, G. and Hanson, R.P. 1971. Serologic survey for arboviruses in Central Alberta. Can. J. Public Health 62: 125-132.
- Iversen, J.O., Wagner, R.J., de Jong, C. and McLintock, J.R. 1973. California encephalitis virus in Saskatchewan: isolation from boreal *Aedes* mosquitoes. Can. J. Public Health 64: 590-594.
- Kettyls, G.D., Verrall, V.M., Wilton, L.D., Clapp, J.B., Clarke, D.A. and Rublee, J.D. 1972. Arbovirus infections in man in British Columbia. Can. Med. Assoc. J. 106: 1175-1179.
- McGowan, J.K., Bryan, J.A. and Gregg, M.B. 1973. Surveillance of arboviral encephalitis in the United States 1955-1971. Am. J. Epidemiol. 97: 198-207.
- McKiel, J.A., Hall, R.R. and Newhouse, V.F. 1966. Viruses of the California encephalitis complex in indicator rabbits. Am. J. Trop. Med. Hyg. 15: 98-102.

- McLean, D.M. 1968. The arboviruses. *In* Textbook of virology. 5th edition, Section 6. *Edited by* Rhodes, A.J. and van Rooyen, C.E. Williams and Wilkins, Baltimore.
- McLean, D.M., Bergman, S.K.A., Goddard, E.J., Graham, E.A. and Purvin-Good, K.W. 1971. North-south distribution of arbovirus reservoirs in British Columbia, 1970. Can. J. Public Health 62: 120-124.
- McLean, D.M., Bergman, S.K.A., Gould, A.P., Grass, P.N., Miller, M.A. and Spratt, E.E. 1975a. California encephalitis virus prevalence throughout the Yukon Territory 1971-1974. Am. J. Trop. Med. Hyg. 24: 676-684.
- McLean, D.M., Bergman, S.K.A., Graham, E.A., Greenfield, G.P., Olsen, J.A. and Patterson, R.D. 1974. California encephalitis virus prevalence in Yukon mosquitoes during 1973. Can. J. Public Health 65: 23-28.
- McLean, D.M., Cobb, C., Gooderham, S.E., Smart, C.A., Wilson, A.G. and Wilson, W.E. 1967. Powassan virus: persistence of virus activity during 1966. Can. Med. Assoc. J. 96: 660-664.
- McLean, D.M., Crawford, M.A., Ladyman, S.R., Peers, R.R. and Purvin-Good, K.W. 1970. California encephalitis and Powassan virus activity in British Columbia, 1969. Am. J. Epidemiol. 92: 266-272.
- McLean, D.M., De Vos, A. and Quantz, E.J. 1964. Powassan virus: field investigations during the summer of 1963. Am. J. Trop. Med. Hyg. 13: 747-753.
- McLean, D.M. and Donohue, W.L. 1959. Powassan virus: isolation of virus from a fatal case of encephalitis. Can. Med. Assoc. J. 80: 708-711.
- McLean, D.M., Goddard, E.J., Graham, E.A., Hardy, G.J. and Purvin-Good, K.W. 1972. California encephalitis virus isolations from Yukon mosquitoes, 1971. Am. J. Epidemiol. 95: 347-355.

- McLean, D.M., Gubash, S.M., Grass, P.N., Miller, M.A., Petric, M. and Walters, T.E. 1975b. California encephalitis virus development in mosquitoes as revealed by transmission studies, immunoperoxidase staining, and electron microscopy. Can. J. Microbiol. 21: 453-462.
- McLean, D.M. and Larke, R.P.B. 1963. Powassan and Silverwater viruses: ecology of two Ontario arboviruses. Can. Med. Assoc. J. 88: 182-185.
- McLean, D.M., MacPherson, L.W., Walker, S.J. and Funk, G. 1960. Powassan virus: surveys of human and animal sera. Am. J. Public Health 50: 1539-1544.
- McLean, D.M., Walker, S.J., MacPherson, L.W., Scholten, T.H., Ronald, K., Wyllie, J.C. and McQueen, E.J. 1961. Powassan virus: investigations of possible natural cycles of infection. J. Infect. Dis. 109: 19-23.
- McLintock, J.R., Burton, A.N., McKiel, J.A., Hall, R.R. and Rempel, J.G. 1970. Known mosquito hosts of western encephalitis virus in Saskatchewan. J. Med. Entomol. 7: 446-454.
- Morgante, O. and Shemanchuk, J. 1967. Virus of California encephalitis complex: isolation from Culiseta inornata. Science 157: 292.
- Pereira, H.G. and Andrewes, C.H. 1972. Viruses of vertebrates. 3rd edition. Bailliere, Tindall and Cassell, London.
- Ritter, D.G. and Feltz, E.T. 1974. On the natural occurrence of California encephalitis virus and other arboviruses in Alaska. Can. J. Microbiol. 20: 1359-1366.
- Rossier, E., Harrison, R.J. and Lemieux, B. 1974. A case of Powassan virus encephalitis. Can. Med. Assoc. J. 110: 1173-1180.
- Rozdilsky, B., Robertson, H.E. and Chorney, J. 1968. Western encephalitis: report of eight fatal cases: Saskatchewan epidemic 1965. Can. Med. Assoc. J. 98: 79-86.

- Wagner, R.J., de Jong, C., Leung, M.K., McLintock, J. and Iversen, J.O. 1975. Isolations of California encephalitis virus from tundra mosquitoes. Can. J. Microbiol. 21: 574-576.
- Watts, D.M., Thompson, W.H., Yuill, T.M., De Foliart, G.R. and Hanson, R.P. 1974. Overwintering of La Crosse virus in *Aedes trisinatus*. Am. J. Med. Hyg. 23: 694-700.
- World Health Organization 1967. Arboviruses and human disease. Technical Report Series No. 369.
- Yuill, T.M., Iversen, J.O. and Hanson, R.P. 1969. Evidence for arbovirus infections in a population of snowshoe hares: a possible mortality factor. Bull. Wildl. Dis. Assoc. 5: 248-253.

