

# OPERATION OF THE GOULDEN LARGE-SAMPLE EXTRACTOR (GLSE) (LASALLE SCIENTIFIC INC., VERSION GLSE-95)

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#### MANAGEMENT PERSPECTIVE

The Goulden Large-Sample Extractor (GLSE) was developed within the Analytical Chemistry Research Project (ACRP) of the Research and Applications Branch (RAB). It provides a means by which, through preconcentration by continuous-flow solvent extraction, detection limits for a comprehensive class of organic contaminants ("Priority Substances") could be lowered to the point that meaningful statements could be made about their distributions in freshwater systems. The technology was originally developed for use by the Water Quality Branch/Ontario Region in the Niagara River and Great Lakes Toxics Management Programs (NRTMP, GLTMP).

The GLSE technique has since received considerable interest on the part of other groups responsible for water quality monitoring (eg. the US EPA and the US Geological Survey) who share similar objectives in the ultratrace monitoring of organic contaminants in freshwater.

Because of early publications and wide-spread dissemination of information on the technique, it could not be patented. In order to assist interested groups in incorporation of the technology in their monitoring programs, the authors have set up an informal relationship to manufacture and commercialize the most "popular" GLSE prototype.

This report describes the operation of this GLSE prototype (GLSE-95) and is intended as a operating manual for LaSalle Scientific Inc. to provide along with the equipment.

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#### WAIVER

Her Majesty, as represented by the Minister of the Environment, and Environment Canada, the National Water Research Institute, the Research and Applications Branch, the Analytical Chemistry Research Project and the authors make no guarantees as to the performance of the GLSE-95 manufactured by LaSalle Scientific Inc. (Guelph, Ontario) in any application.

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#### DISCLAIMER

Her Majesty, as represented by the Minister of the Environment, and Environment Canada, the National Water Research Institute, the Research and Applications Branch, the Analytical Chemistry Research Project and the authors assume no responsibility for damages or consequences of any kind resulting from the use of the GLSE-95 manufactured by LaSalle Scientific Inc. (Guelph, Ontario) in any application including, but not limited to, the proper disposal of extraction effluent.

The use of manufacturer's equipment or trade names does not constitute endorsement or certification by Her Majesty, as represented by the Minister of the Environment or by Environment Canada, the National Water Research Institute, the Research and Applications Branch, the Analytical Chemistry Research Project or by the authors for use in any application.

#### ABSTRACT

This report describes operation of the LaSalle Scientific Inc. version of the Goulden Large-Sample Extractor GLSE-95 prototype and the minimal set-up work involved.

This material serves only as an introductory manual for getting started in the application of GLSE technology to particular water matrices. References to more comprehensive descriptions of the technique are provided.

#### INTRODUCTION

The Goulden Large-Sample Extractor (GLSE) was designed to provide a convenient means by which a certain class of organic contaminants in large samples (50-200 L) of water could be conveniently concentrated to measurable levels for environmental analytical purposes (1-11). When incorporated in an appropriate ultratrace analytical method, the technique can provide data at the parts-per-trillion (ppt) and parts-per-quadrillion (ppq) levels necessary for sensitive monitoring of contaminant distributions and trends in environmental freshwater systems.

The GLSE is a continuous-flow solvent extraction apparatus designed on the mixer/settler principle and optimized to perform an efficient single-stage extraction of contaminants having low solubility (hydrophobic) in water. Many of these contaminants are included in "Priority Substances" lists of Canadian and U.S. governments (US EPA, CEPA, MISA). With this apparatus, continuous compensation is made for solvent loss due to its solubility in water, for continuous pre-heating of the sample to 20°C and for the continuous introduction of surrogate standards for the purpose of evaluating extraction efficiency.

The GLSE-95 apparatus is manufactured and marketed by LaSalle Scientific Inc. as a package for those users who prefer not to assemble the apparatus "from scratch". The apparatus is modelled after the GLSE-95 prototype originally described by Goulden and Anthony (1). An intentional effort has been made to use components, aside from the custom-made glassware, which are easily sourced, rather than to develop and market the apparatus as a user-non-serviceable "black box".

The principal components of the apparatus and their orientation are shown in Figs. 1-4. The parts list is given in Appendix A.

An extensive list of documented applications of the GLSE-95 prototype is provided (Appendix B). The user is advised to consult this information before using the apparatus in a particular application. For applications outside those documented, the user is obliged to perform the necessary research and validation work before any data generated is used.

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#### PRE-INSTALLATION

1.

Pre-installation requirements for the GLSE are minimal. The following provisions should be made :

Due to the hazardous nature of the solvent used (1,2-dichloromethane (DCM), alias "methylene chloride"), adequate ventilation is important. A fume-hood or fume-cabinet would be the ideal location but any well-ventilated laboratory benchspace is satisfactory, particularly if located near a fume-hood or fume cabinet.

Personnel using or working in the vicinity of the extractor should be familiarized with the hazardous properties of the solvent thru the appropriate MSDS.

Government (municipal, provincial, federal) regulations or guidelines applying to hazardous waste discharge should be consulted to determine how extraction effluent will be disposed of (the solubility of the extractant (DCM) in water is 1.6%w/w, 1.3%v/v).

Access (2-3 ft.) to a standard 11: VAC/15 A utility outlet.

Access (2-3 ft.) to a sink, benchtop or floor drain.

3.

2.

The extractor cabinet, accessories and workspace will occupy about 5-6 ft. of bench length.

SET-UP

Bold terms indicate terms defined in Figures.

The GLSE-95 is supplied partially assembled in a carrying-case and only minimal assembly is required for those components which might cause damage to the glassware or become dislodged during shipping.

Before proceeding with set-up, ensure that the components are free of any debris from packing material and that the surfaces of glass "pick-up" and delivery tubes are rinsed free (eg. methanol, dichloromethane (DCM)) of any glue residue from tape used to hold the pieces during shipment.

- using a screw-driver, loosen the mounting straps holding the separator trap and re-orient the separator trap so that the extractor drain is located in the separator trap inlet as shown in Fig.5. Re-tighten the mounting straps and connect the separator trap outlet to a drain using Tygon tubing. Ensure that this drain tube is "free-flowing" and not constricted or of such length so as to develop "plugs" of water in the drain tubing. This may cause overflowing of the separator trap.

- if the sample pump outlet line has not been connected to the heater tube install it on the heater tube inlet at this time. Loosen the heater tube mounting straps using a screw-driver. Expand the end of the Teflon-lined polyethylene (PE) tubing slightly, (heating the tubing end in boiling water will soften it temporarily), using a twisting motion, with the tool provided or a 10 mL transfer pipette. Friction-fit the tube on the inlet to a sufficient length (3/8 to 1/2 inches) and seal the fitting with a hose clamp (provided) (Fig.6).

- if the sample pump output line was connected on shipment, loosen the mounting straps holding the accessory tube (heater tube) using a screw-driver.

- install the Teflon "drip-tip" on the tube's outlet and re-orient the tube so that the outlet is centered in the extractor inlet as shown in (Fig.6). Re-tighten the mounting straps.

- fit the two Teflon spacers on the silica immersion heater, so that when installed in the heater tube, one is located near the bottom (~2" from bottom) and one near the top of the heater (Fig.6). Clamp the heater in place, with the "micro-clamp" fitted over the plastic end-cap of the heater, to the smaller mounting post.

- if not already fitted, press a stainless steel insert (Fig.6) (provided) into the other end of the Teflon-lined PE tubing after it has been cut (scalpel, or X-acto knife) to a length sufficient to reach the sample pump without excessive stress.

- using a long "needle-nose" tweezers, lower the Teflon scrubber column base plate into the column form and rest it on the restricted end of the form (Fig 7). Install the solvent "make-up" delivery tube in the column form so that it rests close to the base plate and tape the other end of the delivery tub to the outer surface of the "scrubber column" (Fig.7). (If the Teflon solvent "make-up" line has not already been threaded through the delivery tube, do this before installing the delivery tube; the end of the Teflon tubing should extend  $\sim 1/4$ " beyond the end of the delivery tube) (Fig.8).

- hold the Teflon restrainer in place so that the perforations are centered over the inlet of the extractor drain, and pour the Teflon "Raschig Rings" packing into the scrubber column form until the level is  $\sim 1$ " above the restrainer. The column packing will hold the restrainer in place (Fig.7).

The scrubber column functions adequately with the packing loosely-packed. Do not, at any time, exert downward pressure on the column as this will drive the base plate through the restriction allowing packing to fall into the extractor. If replacing or trying to orient the solvent "make-up" delivery tube, use a "stirring" motion with little downward pressure.

- if not already installed, thread the intake portion of the Teflon solvent delivery line through the "solvent make-up" pick-up tube so that it protrudes  $\sim 1/4$ " from the end of the tube (Fig.8).

- similarly, install the surrogate standards intake line in its pick-up tube (Fig.9) and the delivery line in its inlet on the heater tube (Fig.6).

- install the thermometer provided in the thermometer port of the heater tube (Fig.6).

- fix the **impeller** in the stirrer so that the impeller will be roughly in the recommended position when the stirrer is mounted (Fig.10). Fit the impeller blades through the stirrer port of the extractor and lower the stirrer until the impeller is located, vertically, approximately at the position indicated in Fig.10. The impeller may have to be tilted to insert it through the stirrer port.

Viewing from above, adjust the stirrer on its mounting rod so that the impeller is axially centered at its vertical position in the **mixing chamber**.

- if not already fitted, press a stainless steel insert into one end of the Teflon-lined PE sample input line and make input and output connections to the sample pump as indicated in Fig.11 using the appropriate size wrenches (5/8" and 11/16"). Make certain to brace movement of the "T" fittings with the wrench while tightening the nuts of the sample lines so that the position of the "T" fittings is not altered (the pulse suppressors must be near-vertical for proper operation). Insert a glass sampling wand into the intake end of the sample input line using a twisting motion.

- the pumps should have been set at the estimated settings for a 50L sample processed at 500 mL/min. If not, adjust them according to the settings in **Table I.** Some "fine adjustment" of these settings may be necessary during the first few extractions (see "RUN-IN").

When not in use, the extractor should be filled with high purity water and the outlets protected from dust with aluminum foil pieces.

#### "RUN-IN"

Before proceeding with the extraction of "real" samples, the extractor should be cleaned by extraction of several samples of high purity water (known to be free of the contaminants being determined, considering the sample size). This will serve as well to give the operator some practice in operation of the extractor and to make fine adjustments in pumping rates for the sample size and sample input rate desired. The example given refers to the extraction of a 50 L sample at a sample input rate of 500 mL/min. Most early validation work was done using similar conditions. The apparatus, however, is designed with the capability to operate up to 1000 mL/min and the user is encouraged to experiment to find the appropriate conditions for the particular application.

- adjust the sample pump input rate to 500 mL/min by calibration using a 1 L graduated cylinder and stopwatch.

- add 300 mL of the extractant, dichloromethane (DCM), to the extractor through the scrubber port and when the DCM level has settled (through a dry column, almost all of this DCM will pass into the extractor), mark its position on the outside of the mixing chamber using a waterproof ink marker.

- begin extraction (see **OPERATION**) of a representative sample size (50 L) of high purity water.

- during the first 30 min of operation, check the solvent "make-up" and surrogate standards delivery lines (eg. insert an air bubble) to ensure that they are delivering liquid.

- after 30 min, stop the extraction and note the level of DCM. If below the mark, add DCM until the level is at the mark, then increase the solvent "make-up" pumping rate. If above the mark, drain DCM until the level is at the mark, then decrease the solvent "make-up" pumping rate.

When adding DCM to the system through the "scrubber" port while it is full of water or while the system is operating, pour the solvent slowly to prevent it from flowing or being carried over into the separator trap.

continue the extraction.

- at 60 min, stop the extraction again and check the DCM level. Repeat the above adjustments if necessary.

- at 90 min, stop the extraction and repeat the above adjustment process if necessary. At this point, 10 min are left in the extraction of 50 L if the sample pump is correctly set. Additionally, the 100 mL of surrogate standard solution should have been delivered by this time (last 10% of the extraction time), allowing time for rinsing the standards container, pump-head and transfer line. If these conditions are not met, adjust the pumps accordingly and finish the extraction.

- practice the extract collection process (see **OPERATION**) then drain all components of the extractor (extractor body, separator trap and heater tube).

- repeat this "run-in" procedure with at least two more 50 L samples of high purity water and until all pump settings are seen to be correct for this sample size and sample input rate, (ie. the DCM level in the extractor should be maintained near the "300 mL" mark through the extraction, the 100 mL surrogate standards solution should be delivered with 10 min allowed to rinse the delivery system during the last 10% of the extraction, and the sample input rate should be well calibrated ((N>/=5) at 500 mL/min).

#### **OPERATION**

Given here is an operating procedure for processing a 50 litre sample of relatively "clean" aqueous sample (ie. low concentrations of dissolved organic matter) which has been clarified of particulate matter. Operation in this application and with this sample size has been thoroughly validated (4,5,6,9).

For operation with more complex matrices, different suites of contaminants, sample sizes and processing rates, the user is referred to more comprehensive applications literature (10,11) and to the list of documentation provided in this manual (Appendix B). If this applications material is insufficient for the intended application, the user, necessarily, must conduct the required research, development and validation.

Extractions are generally conducted at 20°C as most physical constants (eg. octanol-water partition coefficient, Kow) describing solvent extraction processes are derived from data at or near this temperature. Use of the heater tube accessory, then, depends on sample temperature and must be adjusted to bring the sample temperature to 20°C. If the sample temperature is above 20°C, the accessory is not used and the ambient sample temperature is reported.

Control of dispersion is perhaps the most critical aspect of extraction with the GLSE. The advantage with GLSE prototypes is that dispersion is, controllable. The solvent/water dispersion must be restricted primarily to the mixing chamber: this is accomplished by proper adjustment of the stirring speed at a particular sample input rate. A significant flow of solvent should be allowed to recirculate to the mixing chamber through the first solvent recirculation arm, with a minimal amount reaching the second recirculation area, the settling chamber. The "instinct" to maximize dispersion (by stirring at an excessive rate) in order to increase mass transfer of analyte from sample to solvent must be avoided as this serves only to increase analyte loss by the escape of fine droplets of solvent (containing analyte) through the "scrubber" column and separator trap. With some matrices, high in content of dissolved organics or gases, this loss may be so significant as to exceed the solvent "make-up" provision and, if attention is not paid to the extraction, an excessive amount of extract may be lost, making a re-sample necessary. Using a sink tap, provide sufficient dilution water (5-10 L/min) in the drain for dilution of the extraction effluent (see **PRE-INSTALLATION** for comments on disposal of extraction effluent).

Ensure that all drain lines actually feed to the drain and that they are unobstructed (ie. no "kinks", not compressed by other equipment).

Ensure that all stopcocks on the GLSE are closed.

Ensure that stirrer, heater (if to be used) and pump power switches are in the "OFF" position, and that power is switched "ON" at the power bar main switch (Fig.1).

Ensure that there is sufficient DCM in the solvent "makeup" container for the extraction and that the "pick-up" tube is in place.

If on-line surrogate standard addition is being used, ensure that the surrogate standard solution is at room temperature, then fill the standard addition container (100 mL graduated cylinder) with solution and insert the "pick-up tube in the solution.

Sample volume, which must be measured and reported to the analytical laboratory for calculation of concentrations, may be estimated by reliance on sample container calibration. Alternatively, the sample volume may be estimated by timing the extraction period (from the moment sample enters the extractor to the moment water stops entering the extractor) and calculation, using this measurement and the calibrated sample input rate. If the calibrated container method is used, a correction must be made for the volume of the heater tube and sample input line (~300-400 mL).

### Procedure

- 1. Dry the floor of the extractor case before conducting an extraction. This makes visual determination of leaks in connections, tubing, glassware and at stopcocks much easier.
- 2. Drain the extractor components of storage water using the Tygon tubing drain lines.
- 3. Close all stopcocks.
- 4. Add ~300 mL of DCM to the extractor through the "scrubber" port directly from the solvent container or from a cleaned container reserved for this purpose only.
- 5. Fit a cleaned sampling "wand" in the end of the sample input line, then insert the "wand" in the sample.
- 6. Turn on the sample pump.

If estimating sample volume by timing the extraction period at a calibrated sample input rate, start timing as soon as sample begins flowing into the mixing chamber from the heater tube. At the same moment, turn on the solvent "make-up" pump, the surrogate standards pump and the heater (if to be

used) (see General, above).

7.

8.

- After a few hundred mL of sample has entered the extractor (the extractor will be  $\sim 1/3$  to 1/2 full), start the stirrer.
- 9. After 5-10 min of operation, make any adjustments to stirring speed necessary to confine the majority of solvent/water dispersion to the **mixing chamber** with some solvent return in the **first solvent recirculation arm** and minimal dispersion to the **second recirculation chamber**.
- 10. After 20 min of operation, stop the extraction (pause the timing device) and check the DCM level. Make adjustments as necessary (see "RUN-IN").

With some matrices (eg, high dissolved organics, dissolved gases) solvent consumption may be increased above that due to its solubility in water, making an increase in solvent "make-up" necessary.

13

- Re-start the extraction (continue the timing device). At this point, assuming that an effective "run-in" has been done, there should be no need to attend the extraction until 90 min, at which point the surrogate standards container should be rinsed with ~5 mL of methanol to "flush" the standards distribution system.
- 12. As the sample is completed, raise the sampling wand above the level of the extractor inlet and as air begins to enter the heater tube, stop the timing and record the extraction time (min & s, convert to min).
- 13. Turn off all components except the stirrer. Continue stirring for at least 2 min to extract the last portion of sample entering the extractor.
- 14. Turn off the stirrer and allow the phases to separate and clear.
- 15. Before collecting extract, give the stirrer a few momentary "bursts", or stir slowly for several minutes, if necessary, to assist in clearing the DCM phase.

## Extract collection

11.

16. When phases have cleared sufficiently, collect, using the extract drain, the bulk of the extract directly into the extract container, including any "scum"

which has formed at the DCM/water interface. Collect any DCM caught in the sample drain (Fig.4) or which has been carried over into the separator trap (separator trap drain, Fig.5) directly into the extract container.

Ideally, with proper control of the degree of phase mixing, no DCM should be carried over into the separator trap. With some matrices (high concentrations of organics, dissolved gases, dissolved inorganic, or high loadings of fine silt, colloidal material, etc.), this may be unavoidable. Other techniques of dealing with poor separation and clearing of phases are dealt with in (10).

- 17. Drain the separator trap using a Tygon tubing drain line connected to the separator trap drain.
- 18. Drain ~1/2 of the water remaining in the extractor to the sink drain using the sample drain and Tygon tubing.

As water drains from the "scrubber column", DCM which has collected in the column will fall out until the water level is below the column. In draining the extractor, be careful not to drain so fast that this DCM is carried out through the drain line.

- 19. Using the 500 mL separatory funnel supplied with the GLSE, collect DCM which has fallen out of the column and a sufficient amount of extracted water to fill the funnel.
- 20. Add this DCM to the extract container and pour ~1/2 the water in the separatory funnel back into the extractor through the "scrubber" column to flush out remaining DCM.
- 21. Continue this draining and column flushing process using the separatory funnel. With each collection, add any DCM collected to the extract container and use a portion of the water to flush the "scrubber column". The column should have been flushed 3-5 times during this process.
- 22. Before the water level in the mixing chamber has fallen below the impeller, momentarily activate the stirrer to dislodge any DCM which has collected on the upper surface of the impeller.
- 23. Ensure that the DCM extract in its container is layered with  $\sim 1/4$ " of extracted water. This serves to minimize evaporation of DCM during storage.

- A useful measure at this point is to mark the phase meniscus (water/DCM, water/"scum"/DCM) on the outer surface of the extract container with a "permanent" ink marker (cover mark with transparent tape) or with similar marking on a strip of "lab tape" placed vertically on the container. This marking may later be useful in assessing extract loss during transport and storage and in noting the "clearing" of "scum".
- 25. Cap and label the extract container and store refrigerated. Re-tighten the capa few hours later as it may loosen on cooling.
- 26. Close all stopcocks on the GLSE apparatus and fill the apparatus with "high purity" water.

## Intersample Cleaning and Short-Term Storage

24.

If the extract is collected as described in this section, analyte loss due to failure to collect all extract in the GLSE should be minimized to 0-5 mL of extract (ie.< 2% of the total extract collected). It is impractical to attempt to collect this remaining few % of "lost" analyte; the effect will be accounted for in the surrogate standards correction. The intersample cleaning procedure should minimize any significant effect on the subsequent sample. For relatively "pristine" matrices, intersample cleaning of the extractor is limited to filling it with "high purity" water, stirring for 10 min, then draining the water. Alternatively, the extractor body may be rinsed by stirring an "extractor-full" of high purity water with 300 mL of DCM or the entire apparatus may be rinsed by processing 10L or more of high purity water as though it were a sample. More rigorous cleaning methods are discussed in (10).

## PRECAUTIONARY ADVICE

Although application of the GLSE with some aqueous matrices and certain classes of contaminants has been well- studied, the technique remains a research tool and successful application depends on the extent to which the user has done the necessary background investigation (ie. "homework") and is willing to perform the required development and validation work in cases where documentation does not exist.

# SAFETY/HAZARD INFORMATION

The extractant (DCM) is a hazardous substance and all personnel working in the area of the GLSE during operation must be so informed by the provision of the appropriate WHMIS and MSDS information (APPENDIX C). Government regulations (municipal, provincial, federal) must be consulted for limits applicable to the disposal of extraction effluent (see **PRE-INSTALLATION**).

The power distribution bar should be located so that it will not be wetted by activities associated with operation of the GLSE.

#### REFERENCES

- Goulden, P.D. and Anthony, D.H.J. 1985. "Design of a Large-Sample Extractor for the Determination of Organics in Water", Environment Canada, National Water Research Institute. Contribution No. 85-121. Burlington, Ontario, Canada, L7R-4A6.
- Goulden, P.D. and Anthony, D.H.J. 1986. "A Modified Large-Sample Extractor for a 24-Hour Sampling Period", (unpublished manuscript), presented at the 1986 CAPMON (Canadian Atmospheric Precipitation Monitoring Network) Workshop.
- Anthony, D.H.J. and Goulden, P.D. 1987. "Partition Coefficients of Some Niagara River Organic Contaminants", Environment Canada, National Water Research Institute. Contribution No. 86-222. Burlington, Ontario, Canada, L7R 4A6.
- Neilson, M., Stevens, R.J.J., Biberhofer, H., Goulden, P.D. and Anthony, D.H.J. 1987. "A Large-Sample Extractor for Determining Organics in the Great Lakes". Environment Canada, Water Quality Branch/Ontario Region, Inland Waters Directorate (IWD) Technical Bulletin No. 157. Canada Centre for Inland Waters, Burlington, Ontario, Canada, L7R 4A6.

- Neilson, M. and Stevens, R.J.J. 1988. "Evaluation of a Large-Volume Extractor for Determining Trace Organic Contaminant Levels in the Great Lakes". Water Poll. Res. J. Canada, <u>23</u>, (4), 578-588.
- Foster, G.D. and Rogerson, P.F. 1990. "Enhanced Preconcentration of Pesticides from Water Using the Goulden Large-Sample Extractor", Intern. J. Environ. Anal. Chem., <u>41</u>, (3+4), 105-117.
- Kuntz, K. 1988. "Niagara River Sampling Protocol". Environment Canada, Water Quality Branch/Ontario Region, Canada Centre for Inland Waters, Burlington, Ontario, Canada, L7R 4A6.
- 8. Afghan, B.K. 1987. "Analytical Protocol for Monitoring Ambient Water Quality at the Niagara-on-the-Lake and Fort Erie Monitoring Stations". Environment Canada, National Laboratory for Environmental Testing (NLET), Canada Centre for Inland Waters, Burlington, Ontario, Canada, L7R 4A6.
- 9. Afghan, B.K., Agemian, H. and Forbes, M.A. 1987. "Validation of the Analytical Protocol for Monitoring Ambient Water Quality at the Niagara-on-the-Lake and Fort Erie Monitoring Stations". Environment Canada, National Laboratory for Environmental Testing (NLET), Canada Centre for Inland Waters, Burlington, Ontario, Canada, L7R 4A6.

- Anthony, D.H.J. 1992. "Assembly, Set-Up and Operation of the Goulden Large-Sample Extractor in Field and Laboratory Settings". Environment Canada, National Water Research. Contribution No. 93-xx (in preparation). Burlington, Ontario, Canada, L7R 4A6.
- Anthony, D.H.J. 1992. "Incorporation of Goulden Large-Sample Extraction (GLSE) Technology in Water Quality Monitoring and Research Programs". Environment Canada, National Water Research Institute. Contribution No. 93-xx (in preparation). Burlington, Ontario, Canada, L7R 4A6.

# TABLE I : OPERATIONAL SETTINGS

Component Description Setting Comments					
Component Description S					
SAMPLE PUMP (FMI Model # QD 2CSC)	4.2	500 mL/min (nominal)			
SOLVENT "MAKE-UP" PUMP (FMI Model # RRP SY 2CSC)	1.4	6.50 mL/min (nominal)			
SURROGATE STANDARDS PU (FMI Model # RP SY 0CSC)	VMP 2.2	1.11 mL/min (nominal)			
STIRRER Caframo Model # RZR50	800-1000	Setting approximates RPM and is dependent on phase mixing characteristics (See <b>OPERATION</b> )			
HEATER CONTROLLER Cole-Parmer # (Used with Corning Model No. Silica Immersion Heater)	0-100%	Setting depends on sample temperature (eg. 40% for sample at 5°C processed at 500 mL/min to reach 20°C)			

### **APPENDIX I**

A1

# PARTS LIST FOR THE GOULDEN LARGE-SAMPLE EXTRACTOR (LASALLE SCIENTIFIC INC. PACKAGE)

	ALLE F NUMBER
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GLSE-95

# MAJOR GLASSWARE COMPONENTS

Extractor Body	1	LASALLE SCIENTIFIC INC.
Separator Trap	1	121 Malcolm Rd. Guelph, Ontario N1K 1A8
Accessory Tube	1	(519) 824-7301 FAX (519) 824-9576

# MINOR GLASSWARE COMPONENTS

Spiking Inlet (Set/2)	1
Spike Pick-up Tube (Set/4)	1
Solvent Delivery Tube (Set/4)	1
Solvent Pick-up Tube (Set/4)	1
Vent Tube (Set/4)	1

Pulse Suppressor (Set/2) Sampling Wand (Straight) (Set/6) Sampling Wand (Right-angle) (Set/6)

# EXTRACTOR CABINET

LASALLE SCIENTIFIC INC. (above)

# ELECTRICAL/MECHANICAL COMPONENTS

Power Outlet	6 outlets, idependently switched, independently fused, 115VAC/15A	1
Immersion Heater	Silica,115VAC/15A, 1000W	1
Power Controller	SCR, 115VAC/15A, 1000W	1
Stirrer	High torque, constant torque, chuck size to 10 mm.OD, suitable for continuous operation, variable speed to 2000 rpm	1
Pump (Sample)	Positive displacement, high precision delivery (±1 % or better), non-contaminating and chemically inert wetted components, suitable for continuous operation, 0-1000 mL/min.	1

## A2

2

1

1

Pump (Solvent ( 1 Compensation) 0-45 mL/min. Pump (Standard ( ) 1 Addition) 0-5mL/min. PUMP FITTINGS Male Run Tee 316 Stn.Stl., 3/8" tube OD to 1/4" 2 male pipe (NPT) thread Male Elbow 316 Stn.Stl.,1/8" tube OD to 1/4" 2 male pipe (NPT) thread Inserts 316 Stn.Stl.,3/8" tube OD to 1/4" 2 tube ID Ferrules Teflon, 3/8" tube OD, set of front 10 and back ferrules Adapter Teflon, low-flow isolation 1 adapter for Stn.Stl. pump-heads, Set/2

A3

Small-bore1/4-28 fittings kit for 1/16" ODfitting kitTeflon tubing

1 LASALLE SCIENTIFIC INC. (above)

## MATERIALS

Teflon TFE (virgin),1/32" thick, 1 : Sheet, Teflon 12" \* 12 "

6ft. Teflon TFE (virgin), Tubing, Teflon 1/8" OD \* 1/16" ID

Teflon TFE (virgin),5/16" ID, 15ft. Tubing, Teflon fractional spaghetti tubing, standard wall

Teflon TFE (virgin),3/16" ID, 1ft. Tubing, Teflon fractional spaghetti tubing, standard wall

10ft. Polyethyene, Teflon FEP lined, Tubing 3/8" OD \* 1/4" ID, Penntube Plastics type "Pureline"

4 mm. OD \* 0.8 mm. ID Tubing, Silicone (Size 13)

1/4" OD \* 1/8" ID Tubing, Silicone

9/16" ID \* 3/32" wall, Tubing, Tygon Norton Performance Plastics type R-3603

1ft.

1ft.

10ft.

### A5

# Tubing, Tygon

3/8" ID \* 3/32" wall, Norton Performance Plastics type R-3603

## MISCELLANEOUS PARTS

Clamp Micro, Polymer Tech Model No.58858 Clamp Holder Lab-Line Model No.7040 Clamp Holder Jumbo, Polymer Tech Model No. 58875 Base Foot Lab frame Rod Aluminum, hardened, corrosionresistant, 1/2" diameter, 36" length cut to 26" Stirring Paddle 3 bladed, 2" diameter, 10" length (eg. Talboy's Engineering Corp. Model No. 151) Separatory Funnel

10ft.

1

· 3

1 4

1

Teflon FEP, 500 mL, "Nalgene"

1

1

Support Ring	Extension, "Humboldt" Model No. H18440		2	•	
Thermometer	Dial type, -10 to +100°C	1.			
Hose clamps	5" OD (nom.) 3 & 1/2" OD (nom.) 2 & 1/2" OD (nom.) 1/2" OD (nom.)		3 1 3 2		

A6

# GLSE-95-PKG

Complete GLSE system, including all the above components, assembled and mounted in a transportable cabinet

A7

1

1

## 1 LASALLE SCIENTIFIC INC. (above)

# EXTRACT PROCESSING GLASSWARE

Goulden Evaporator I Distillation of dichloromethane Fig. 12 (References 8,10)

Goulden Evaporator II Distillation of hexane Fig. 13 (References 8,10) LASALLE SCIENTIFIC INC. (above)

### DOCUMENTATION OF GOULDEN LARGE-SAMPLE EXTRACTOR (GLSE) TECHNOLOGY:

**B**1

"APPENDIX B

#### DEVELOPMENT, VALIDATION AND APPLICATIONS STUDIES

From:

Anthony, D.H.J. 1993. "Incorporation of Goulden Large-Sample Extraction Technology in Water Quality Monitoring and Research Programs", National Water Research Institute, Contribution No. 92-.

## DOCUMENTATION OF GOULDEN LARGE-SAMPLE EXTRACTOR (GLSE) TECHNOLOGY:

- Goulden, P.D. and Anthony, D.H.J. 1985. Design of a Large Sample Extractor for the Determination of Organics in Water. Environment Canada, National Water Research Institute Report 85-121, Burlington, Ontario, Canada.
- Goulden, P.D., and Anthony, D.H.J. 1986. A Modified Large-Sample Extractor for a 24 HR Sampling Period. Presented at CAPMON (Canadian Precipitation Monitoring Network) Workshop.
- Kuntz, K.W., Goulden, P.D., Biberhofer, H. and Anthony, D.H.J. 1986. Sampling and Extraction of Large Volume Water Samples for Trace Organic Analysis. Presented at American Water Resources Assoc. Symposium, Syracuse, NY, 17-20 May 1987. Presented at "Preconcentration of Trace Organic Compounds from Large Volumes of Water"; Workshop Proceedings, Canada Centre for Inland Waters, Burlington, Ontario, Canada, 23 June.
- Neilson, M., Stevens, R., Biberhofer, H., Goulden P.D. and Anthony, D.H.J. 1987. A Large Sample Extractor for Determining Organics in the Great Lakes. Environment Canada, Water Quality Branch, IWD Technical Bulletin #157.
- Anthony, D.H.J. and Goulden, P.D. 1987. Partition Coefficients of some Niagara River Organic Contaminants. Environment Canada, National Water Research Institute Report 86-222, Burlington, Ontario, Canada.
- Afghan, B.K., Carron, J., Goulden, P.D., Lawrence, J., Leger, D., Onuska, F., Sherry, J. and Wilkinson, R. 1987. Recent Advances in Ultratrace Analysis of Dioxins and Related Halogenated Hydrocarbons. Can. J. Chem. <u>65</u>, 1086-1097.
- Goulden, P.D. 1987. Liquid-Liquid Extraction. Proceedings of Workshop on Preconcentration of Trace Organic Compounds from Large Volumes of Water. Held at the Canada Centre for Inland Waters, Burlington, Ontario, 23 June.
- Analytical Protocol for Monitoring Ambient Water Quality at the Niagara-on-the-Lake and Fort Erie Stations. 1987. Environment Canada, National Water Quality Laboratory, Canada Centre for Inland Waters, 867 Lakeshore Rd., P.O. Box 5050, Burlington, Ontario, Canada, L7R 4A6.

**B2** 

- Afghan, B.K., Agemian, H., and Forbes, M.A. 1987. Validation of Analytical Protocol for Monitoring Ambient Water Quality at the Niagara-on-the-Lake and Fort Erie Stations. Environment Canada, National Water Quality Laboratory, Canada Centre for Inland Waters, 867 Lakeshore Rd., P.O. Box 5050, Burlington, Ontario, Canada, L7R 4A6.
- Niagara River Data Interpretation Group. 1988. A Joint Evaluation of Upstream/Downstream Niagara River Monitoring Data for the period April 1986 to March 1987. Environment Canada, Water Quality Branch, Inland Waters/Lands Directorate, Ontario Region, Canada Centre for Inland Waters, 867 Lakeshore Rd., P.O. Box 5050, Burlington, Ontario, Canada, L7R 4A6.
- Merriman, J.C. and Metcalfe, J.L. 1988. Pesticide Distribution, Lower Ottawa River, 1986.
  Environment Canada, Water Quality Branch, Inland Waters/Lands Directorate, Ontario Region, Technical Bulletin #160. Canada Centre for Inland Waters, 867 Lakeshore Rd., P.O. Box 5050, Burlington, Ontario, Canada, L7R 4A6.
- Anthony, D.H.J. 1988. Application of Continuous-Flow Liquid-Liquid Extraction Technology Technology to Great Lakes/Niagara River Toxic Organic Contaminant Monitoring Programs. Presented at the 31st Conference of the International Association for Great Lakes Research (IAGLR), 17-20 May, Hamilton, Ontario.
- Anthony, D.H.J. 1988. Large-Sample Solvent Extraction Technology for Improving the Detectability of Organic Contaminants in Water. Presented at the 3rd Chemical Congress of North America, 05-10 June, Toronto, Ontario.
- Mullin, M.D., Filkins, J.C., Koshko, C.A. and Utz, J.L. 1988. A Reconnaissance Survey of Green Bay for Congeneric PCBs. Presented at the 31st Conference of the IAGLR, 17-20 May, Hamilton, Ontario.
- Swackamer, D.L., Holmes, M.W. and Skoghind, R.S. 1988. Comparison of Methodologies for Isolating PCBs from Water. Presented at the 31st Conference of the IAGLR, 17-20 May, Hamilton, Ontario.
- Merriman, J.C. 1988. Distribution of Organic Contaminants in Water and Suspended Solids of the Rainy River. Water Poll. Res. J. Canada, 23, (4), 590-600.
- Green, D. 1988. Determination of Contaminant Concentrations Across the Niagara River Using Automatic In-Situ Water Samplers. Final Report for UPDP DSS Contract No. 02SE -KW405-7-9195, Seastar Instruments, Dartmouth, Nova Scotia.

 Neilson, M. and Stevens, R. 1988. Evaluation of a Large-Volume Extractor for Determining Trace Organic Contaminant Levels in the Great Lakes. Water. Poll. Res. J. Canada, <u>23</u>, (4), 578-588.

- Stevens, R.J.J. and Neilson, M.A. 1989. Inter- and Intralake Distributions of Trace Organic Contaminants in Surface Waters of the Great Lakes. J. Great Lakes Res, <u>15</u>, (3), 377-393.
- Gregor, D.J. and Gummer, W.D. 1989. Evidence of Atmospheric Transport and Deposition of Organochlornie Pesticides and Polychlornated Biphenyls in Canadian Arctic Snow. Environmental Science & Technology, 23, (5), 561-565.
- Gates, P.M. and Foster, G.D. 1989. Preconcentration of Pesticides from Water into Dichloromethane using the Goulden Large-Sample Extractor. Presented at the 31st Rocky Mountain Conference Program, 30 July - 04 August, 1989, Denver, Colorado, U.S.A. Abstract #205.
- \*Gates, P.M., \*Foster, G.D. and Rinella, F.A. 1989. Pesticide Occurrence and Transport in the Yakima River Basin, Washington. Presented at the 10th Annual Conference of the Society of Environmental Toxicology and Chemistry, Toronto, Ontario, Canada. \*U.S. Geological Survey, National Water Quality Laboratory, Arvada, CO, U.S.A., 80002.
- Frez, W.A., Lancaster, E.L., Kuroda, S.M., Fisher, J.E., Utz, J., \*Filkins, J.C. 1989. A Laboratory Evaluation of PCB Congener and Organochlorine Extraction Efficiences Using a Large Volume Continuous-Flow Extractor. Presented at the 10th Annual Conference of the Society of Environmental Toxicology and Chemistry, Toronto, Ontario, Canada. \*U.S. EPA, Large Lakes Research Station, Grosse Ile, MI, U.S.A.
- Tsanis, I.K., Biberhofer, J., Murthy, C.R. and Sylvestre, A. 1989. Loadings of Selected Chemicals into the St. Lawrence River System from Lake Ontario - 1986/87. Water Poll. Res. J. Canada, 24, (4), 589-608.
- Foster, G.D., Foreman, W.T. and Gates, P.M. 1990. Preconcentration of Pesticides from Stream Waters Using High Capacity C-18 Sorbent Cartridges and the Goulden Large-Sample Extractor. Presented at the 44th Mid-Atlantic Regional American Chemical Society Meeting, 21-25 May, 1990, Madison, N.J., U.S.A.

**B4** 

- Foster, G.D., Gates, P.M. and Foreman, W.T. 1990. Comparison of Large-Sample Liquid-Liquid and Solid-Phase Preconcentration Techniques to Lower Detection Limits for Pesticides in Water. Presented at the 32nd Rocky Mountain Conference Program, 29 July - 03 Aug., 1990, Denver, CO., U.S.A. Abstract #79.
- Gates, P.M., Foreman, W.T., McKenzie, S.W., Rinella, F.A. and Foster, G.D. 1990. Pesticide Occurrence and Transport in the Yakima River Basin, Washington. Presented at the 32nd Rocky Mountain Conference Program, 29 July - 03 August, Denver, CO., U.S.A. Abstract #80.
- Ibid. Presented at the 11th Annual Meeting of the Society of Environmental Toxicology and Chemistry (SETAC), 11-15 Nov., Arlington, VA, U.S.A. Abstract #P019.
- Anthony, D.H.J. 1990. Recommendations to the Prairie Provinces Water Board Committee on Water Quality Regarding the Application of Large-Sample Extraction Technology in the Development of a New Trans-Boundary Water Quality Program. Written version of presentation, 07 Nov. 1989, to PPWB Water Quality Monitoring Workshop, Winnipeg, Manitoba, 15 pp.
- Foster, G.D. and Rogerson, P.F. 1990. Enhanced Preconcentration of Pesticides from Water Using the Goulden Large-Sample Extractor. Intern. J. Environ. Anal. Chem., <u>41</u>, (3+4), 105-117.
- Foster, G.D., Foreman, W.T. and Gates, P.M. 1991. Performance of the Goulden Large-Sample Extractor in Multiclass Pesticide Isolation and Preconcentration from Stream Water. J. Agric. Food Chem., <u>39</u>, (9), 1618-1622.
- Merriman, J.C., Anthony, D.H.J., Kraft, J.A., and Wilkinson, R.J. 1991. Rainy River Water Quality in the Vicinity of Bleached Kraft Mills. Chemosphere, 23, (11/12), 1605-1615.
- Anthony, D.H.J. 1991. Preconcentration for Ultra-Trace Determination of Organic Contaminants in Aqueous Matrices using a Continuous-Flow Solvent Extraction Technique. Presented at the 74th Canadian Chemical Conference and Exhibition, 02-06 June, Hamilton, Ontario.

Anthony, D.H.J. 1992. An Environmentally Isolated Continuous-Flow Solvent Extraction Apparatus for Monitoring Ambient Levels of Hydrophobic Organic Contaminants in Freshwater Matrices. Presented at the 1992 Pittsburgh Conference on Analytical Chemistry and Applied Spectroscopy, 09-13 March, New Orleans, Louisiana, U.S.A. Abstract #145P.

- Afghan, B.K. and Wilkinson, R.J. 1992. Evaluation of the Method for the Determination of Dioxins and Furans in Pulp and Paper Industry Related Matrices. NWRI Contribution No. 92-10.
- Anthony, D.H.J. 1993. Incorporation of Goulden Large-Sample Extraction (GLSE) Technology in Water Quality Monitoring Programs. Preprint Extended Abstract #68, <u>31</u>, (1), 400-403. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, <u>28 March</u> - 02 April, 1993, Denver, CO, USA.
- Wiens, B. 1993. Quality Control During GLSE Extraction of Niagara River Water. Abstract #69. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, 28 March 02 April, 1993, Denver, CO, USA.
- Föster, G.D. 1993. Description of Extraction Theory for the Isolation of Trace Organics from Water in the Goulden Large-Sample Extractor. Preprint Extended Abstract #70, <u>31</u>, (1), 405-407. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, 28 March - 02 April, 1993, Denver, CO, USA.
- Anthony, D.H.J. 1993. Application of the Goulden Large-Sample Extraction (GLSE) Technique in Sampling "Difficult" Environmental Aqueous Matrices. Preprint Extended Abstract #71, <u>31</u>, (1), 408-411. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, 28 March - 02 April, 1993, Denver, CO, USA.
- Gates, P.M. and Foreman, W.T. 1993. The Operation of the Goulden Large-Sample Extractor in the Yakima River Basin, Washington. Preprint Extended Abstract #72, 31, (1), 412-414. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, 28 March - 02 April, 1993, Denver, CO, USA.

Foreman, W.T., Gates, P.M. and Foster, G.D. 1993. Performance of the Goulden Large-Sample Extractor for Preconcentration of Pesticides in Surface Water Samples from the Yakima River Basin, Washington. Preprint Abstract #73, 31, (1), 415-417. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, 28 March - 02 April, 1993, Denver, CO, USA.

- Headley, J.V., Swyngedouw, C., Crosley, R. and Whitley, G. 1993. Evaluation of the Goulden and Pressure-Can Large Volume Samplers for Acidic Compounds in Natural Waters. Abstract #89. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, 28 March - 02 April, 1993, Denver, CO, USA.
- Pearson, R.F., King, P.A., Holmes, M.W., Eisenreich, S.J. and Swackhamer, D.L. 1993. Continuous Flow Liquid-Liquid Extraction: Operational Variables Influencing the Recovery of PCBs. Preprint Extended Abstract #90, 31, (1), 441-444. Presented before the Division of Environmental Chemistry, 205th American Chemical Society National Meeting, 28 March - 02 April, 1993, Denver, CO, USA.



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emergency telephone no.	312/973-3600 (American Scientific Products)			MATERIAL SAFETY	
chemtrec telephone no	800/424-930	0		DATA SHEET	
information telephone no.	616/726-317	1 (American Burdick & Jacks	on)		
I. Identification					
chemical name	Methylene C	hloride molecular weight	84.93	ETHYLENE CHLORI	
chemical family	Chlorinated		12012		
synonyms	Dichloromet	hane, Methylene Dichloride		2-DICHLOROMETHA	
DOT proper shipping name.	Ada Abadama C	hloride or Dichloromethane	(-/	2-DICHLOROMETRA	
DOT hazard class	ORM-A	······································			
DOT identification no.	UN1593	CAS no75-09-2	· ·		
	minal Nata				
II. Physical and Che	39.75°C	freezing point	nin and a sta	(ether=1) ca 0.7	
boiling point, 760mm Hg vapor pressure at 20°C		vapor density (air = 1) 2.9		A 2000 1 001	
		specific gravity ( $H_2O = 1$ ) $O = 1$	solubility in water		
% volatiles by volume			•33stability	Stanie	
hazardous polymerization		Not expected to occur.			
appearance and odor		Clear, colorless liquid with a			
conditions to avoid	· · · · · · · · · · · · · · · · · · ·	Heat, sparks, open flame, open	en containers, poo	r ventilation, and	
		moisture.			
	<u></u>	Active metals and strong alk	-line colutions	<b>`</b>	
materials to avoid		Active metals and strong ark	anne solutions.		
				· · · · · · · · · · · · · · · · · · ·	
hazardous decomposition pr		Phosgene, hydrogen chloride,	and chlorine.		
macarooos accomposition pr	ــــــــــــــــــــــــــــــــــــــ				

III Fire and Evaluation Harard Data

III. Fire and Explosion Mazard Data				
flash point, (test method)	None (closed cup)	auto ignition temperature	556°C	
flammable limits in air % by volume: lower limit.	12.0	upper limit	19.0	
unusual fire and explosion hazards	Concentrated vapors can be ignited by high intensity heat source or			
	flame. Toxic and corrosive gases are formed on contact with flame			
	or hot glowing surfa	ces.		
extinguishing media	Non-flammable mat	erial. Use dry chemical, o	carbon dioxide, foam,	
· · · · · · · · · · · · · · · · · · ·	water spray as appropriate for surrounding fire and materials.			
special fire fighting procedures	Non-flammable material. Wear full protective clothing and self-			
	contained breathing apparatus. Heat will build pressure and may			
	rupture closed stora	e containers. Keep fire-	exposed containers	
	cool with water spra	y		
IV. Hazardous Components	•		· · · · · · · · · · · · · · · · · · ·	
Methylene Chloride	%C	a 100 TLV 100 ppm	1 CAS no 75-09-2	

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#### V. Health Hazards\_

Occupational Exposure Limits			<u>Concentration Immediately Dangerous</u> <u>to Health</u>	
OSHA	8-hour PEL Ceiling Peak	– 500 ppm – 1000 ppm – 2000 ppm	OSHA/NIOSH	5000 ppm
ACGIH	TLV-TWA TLV-STEL (15-min)	– 100 ppm	Odor Threshold	
		– 500 ppm	OHS	200 ppm
NIOSH	TLV-TWA TLV-C (1-hour)	– 75 ppm – 500 ppm	NSC	200 ppm

## Carcinogenic, Mutagenic, and Teratogenic Data

Indefinite animal carcinogen (IARC) Experimental carcinogen (NTP) Positive mutagen (RTEC)

## Primary Routes of Entry

Methylene chloride may exert its effects through inhalation, skin absorption, and ingestion.

# Industrial Exposure: Route of Exposure/Signs and Symptoms

Inhalation: Exposure can cause light-headedness, vertigo, drowsiness, narcosis, headache and dizziness, unconsciousness, and even death in extreme cases. Exposure to vapors can elevate carboxyhemoglobin levels in the cardiovascular system.

Eye Contact: Liquid or high vapor concentration can cause pain and irritation with slight corneal injury possible.

Skin Contact: Prolonged or repeated skin contact can cause irritation and dermatitis through defatting of skin.

Ingestion: Can cause burning of throat and mouth.

## Effects of Overexposure

Acute inhalation or ingestion causes mild central nervous system depression. The primary toxic effect is narcosis. Other toxic effects are pulmonary edema, encephalopathy, and hemolysis. Methylene chloride irritates the eyes, skin and respire ory tract. No systemic effects have been reported in humans, although excess ve concentrations have caused liver and kidney damage in animals.

### Medical Condition Aggravated by Exposure

Preclude from exposure individuals with diseases of liver, kidneys; cardiovascular and central nervous systems, and heavy smokers. Simultaneous exposure to methylene chloride and alcohol can increase the toxic hazards of methylene chloride.

#### Emergency First Aid

Inhalation: Immediately remove to fresh air. If not breathing, administer mouth-to-mouth resuscitation. If there is no pulse, administer cardiopulmonary resuscitation (CPR). Contact physician immediately.

Eye Contact: Rinse with copious amounts of water for at least 15 minutes. Get emergency medical assistance.

Skin Contact: Flush thoroughly for at least 15 minutes. Wash affected skin with soap and water. Remove contaminated clothing and shoes. Wash clothing before re-use and discard shoes. Get emergency medical assistance.

Ingestion: Call local Poison Control Center for assistance. Contact a physician immediately. Never induce vomiting or give anything by mouth to a victim unconscious or having convulsions.

#### VI. Safety Measures and Equipment.

Ventilation:

Adequate ventilation is required to protect personnel from exposure to chemical vapors exceeding the PEL. The choice of ventilation equipment, either local or general, will depend on conditions of use, quantity of material, and other operating parameters.

Respiratory:

Skin:

Use approved respirator equipment. Follow NIOSH and equipment manufacturer's recommendations to determine appropriate equipment (air-purifying, air-supplied, or self-contained breathing apparatus).

Eyes: Safety glasses are considered minimum protection. Goggles or face shield may be necessary depending on quantity of material and conditions of use.

Protective gloves and clothing are recommended. The choice of material must be based on chemical resistance and other user requirements. Generally, neoprene or Buna-N offers acceptable chemical resistance. Individuals who are acutely and specifically sensitive to methylene chloride may require additional protective equipment. Storage:

Methylene chloride should be protected from moisture, temperature extremes, and direct sunlight. Proper storage of methylene chloride must be determined based on other materials stored and their hazards and potential chemical incompatibility. In general, methylene chloride should be stored in a cool, well ventilated and secure toxic storage room.

Other:

Emergency eye wash fountains and safety showers should be available in the vicinity of any potential exposure.

### VII. Spill and Disposal Data\_

Wear protective clothing and use approved respirator equipment. **Spill Control:** Absorb spilled material in an absorbent recommended for solvent spills and remove to a safe location for disposal by approved methods. If released to the environment, comply with all regulatory notification requirements.

#### Waste **Disposal:**

Dispose of methylene chloride as an EPA hazardous waste. Hazardous waste number - U080(Toxic).

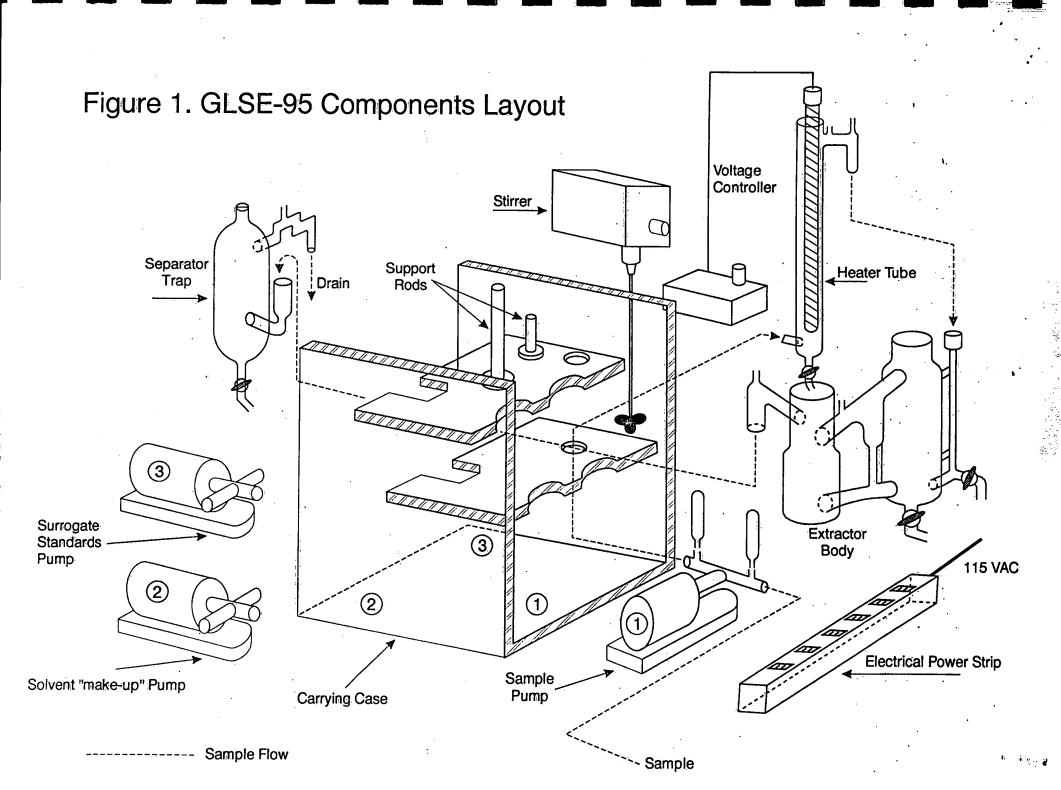
**Revision Date: 1/85** 

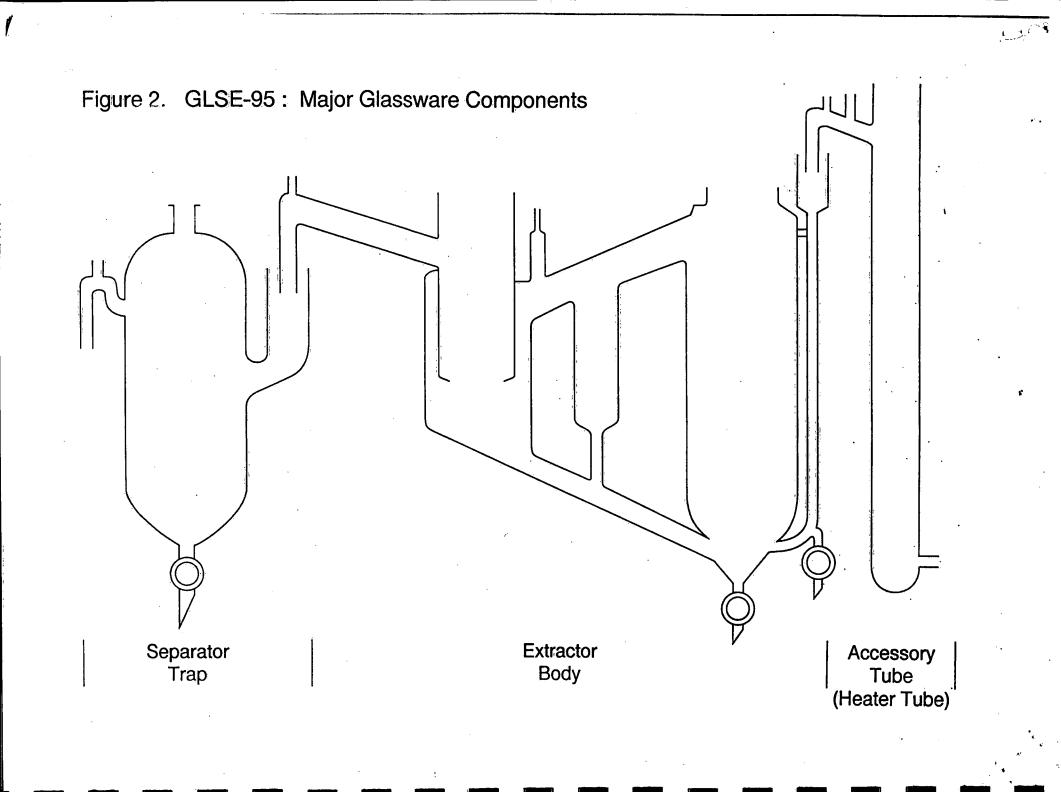
KEY

C3

ca na C	Approximately Not applicable Ceiling	STEL TLV TWA BuAc	Short Term Exposure Level Threshold Limit Value Time Weighted Average Butyl Acetate
PEL	Permissable Exposure Level	BuAc	Butyl Acetate

NSC National Safety Council ("Fundamentals of Industrial Hygiene", 1983) OHS Occupational Health Services ("Hazardline")





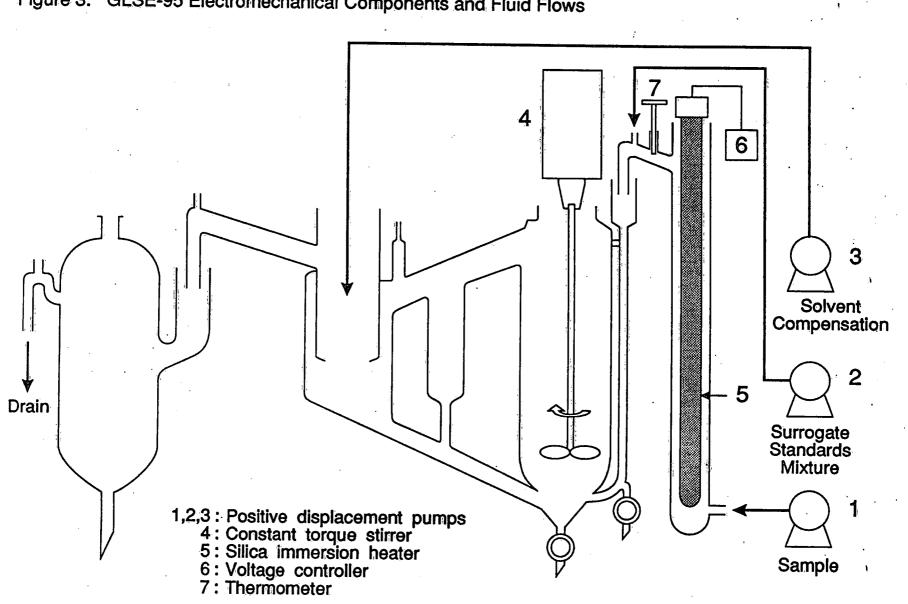
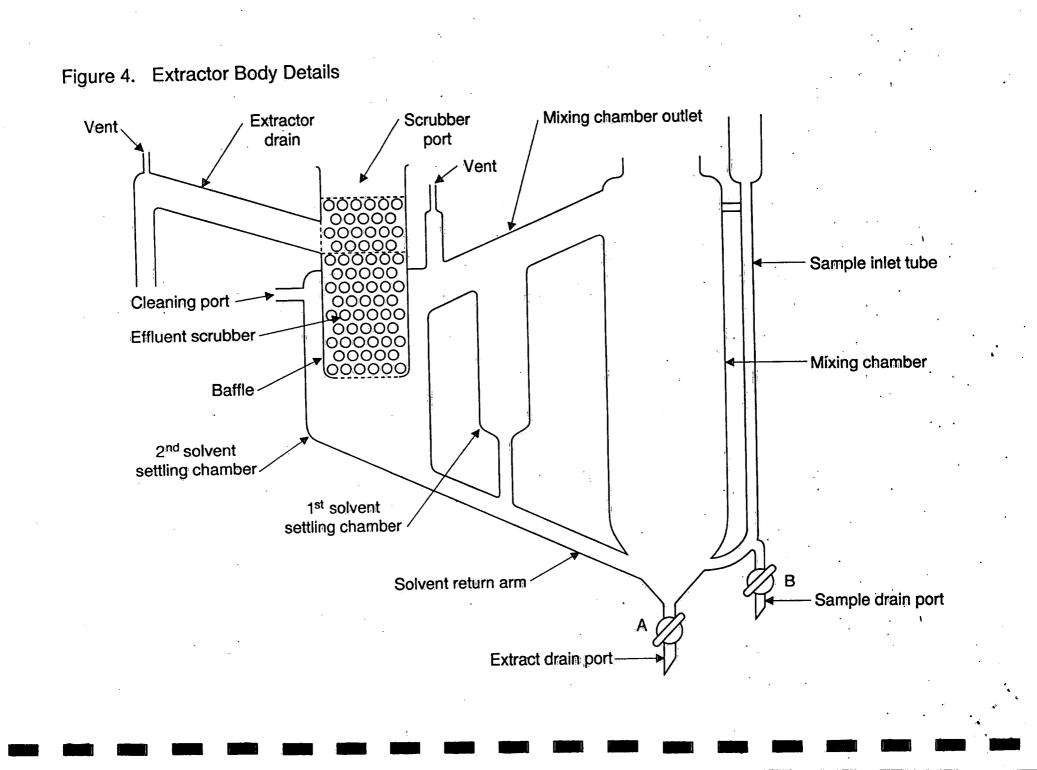
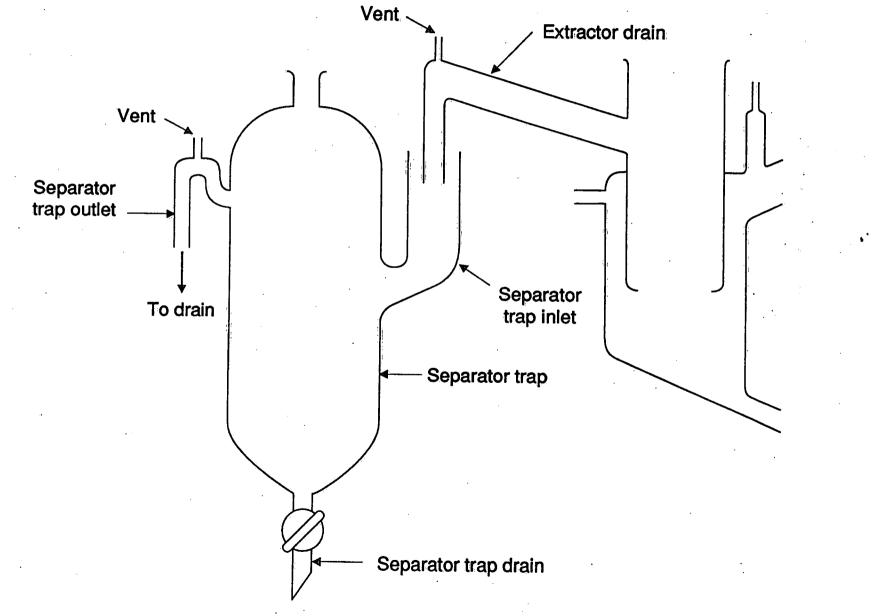


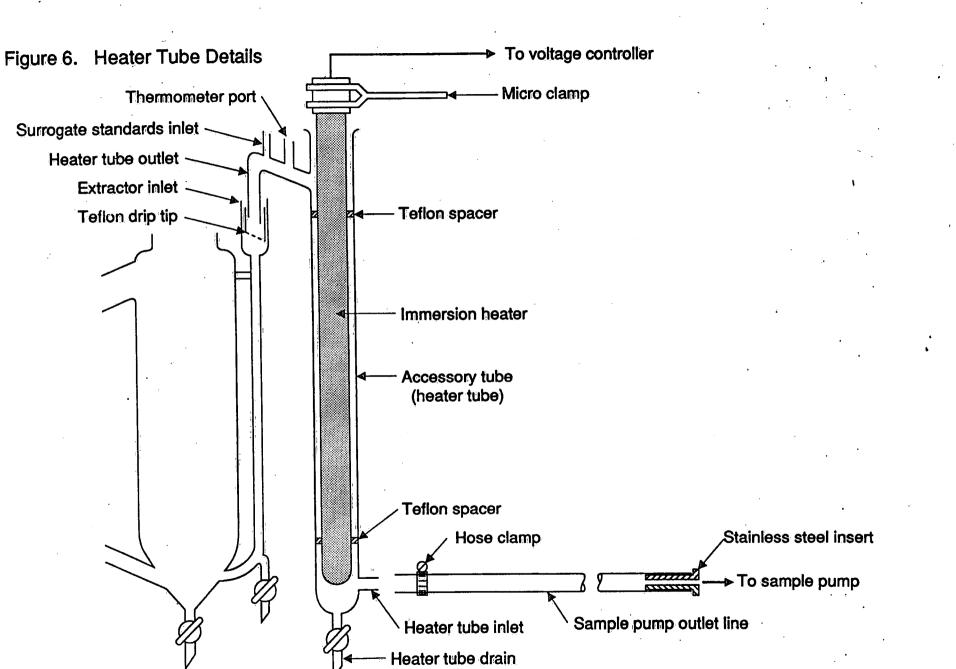
Figure 3. GLSE-95 Electromechanical Components and Fluid Flows





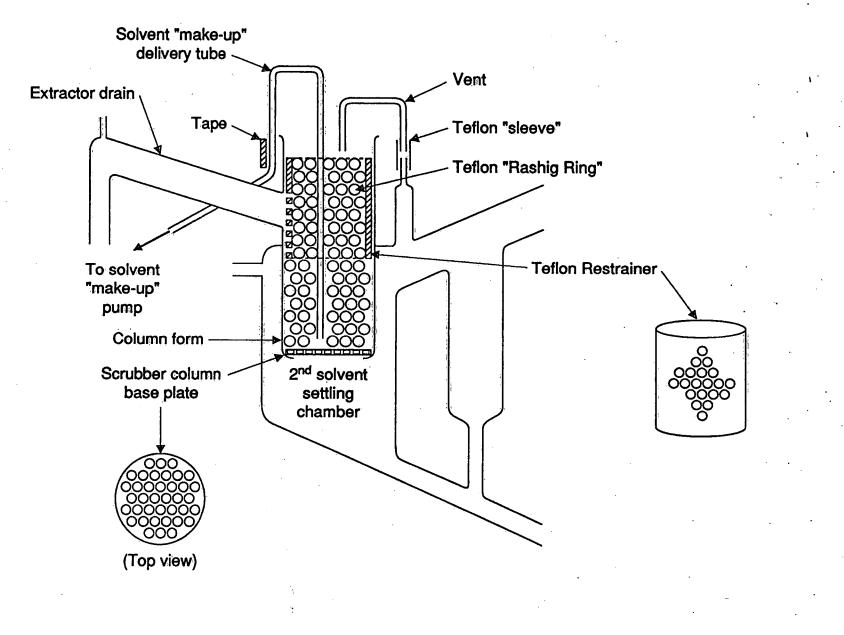


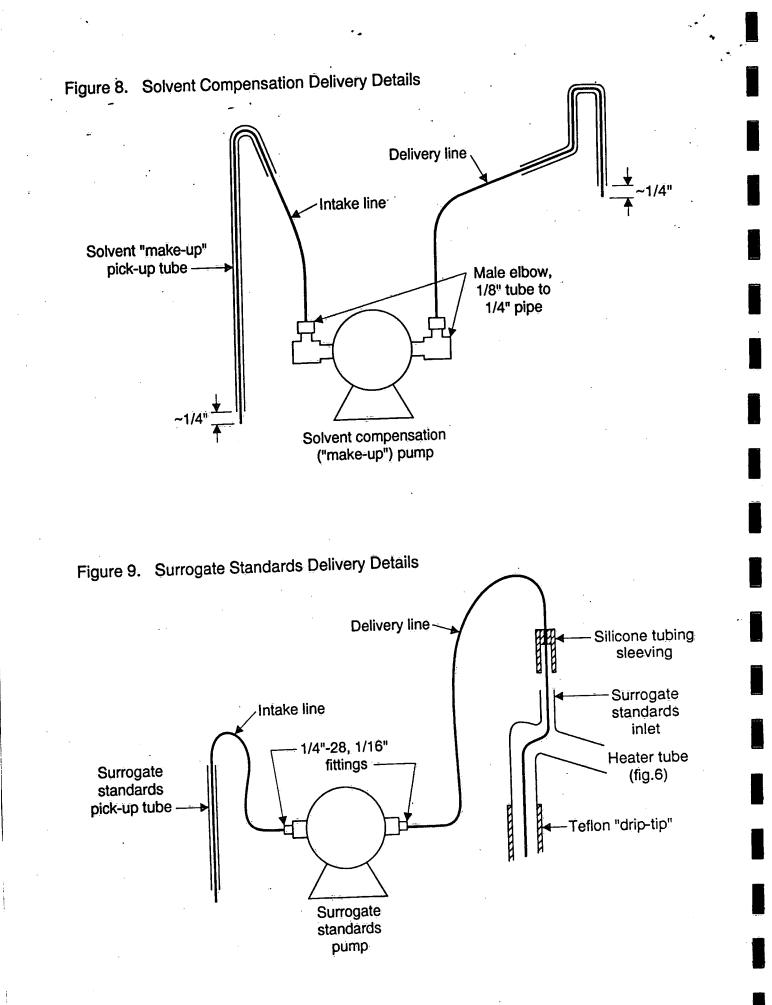
 $C_1 C_2 = 2$ 



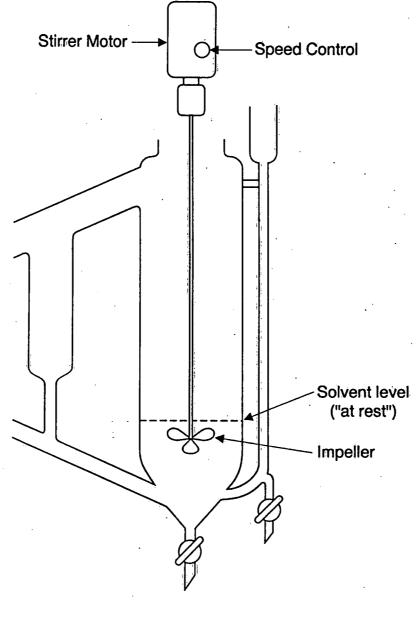
<u>é 51. 2</u>



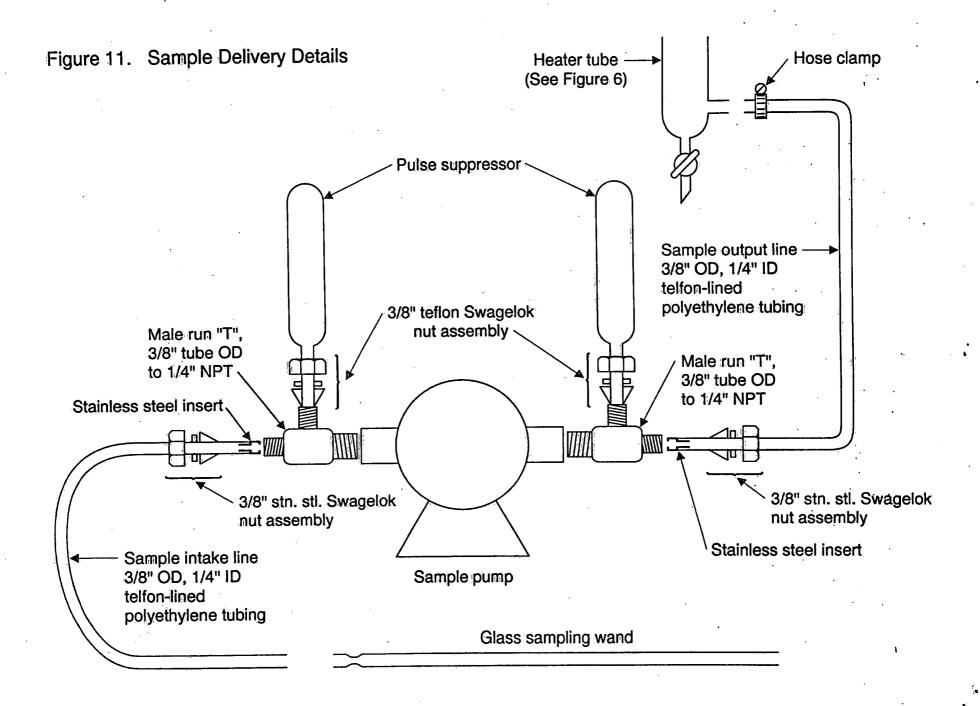


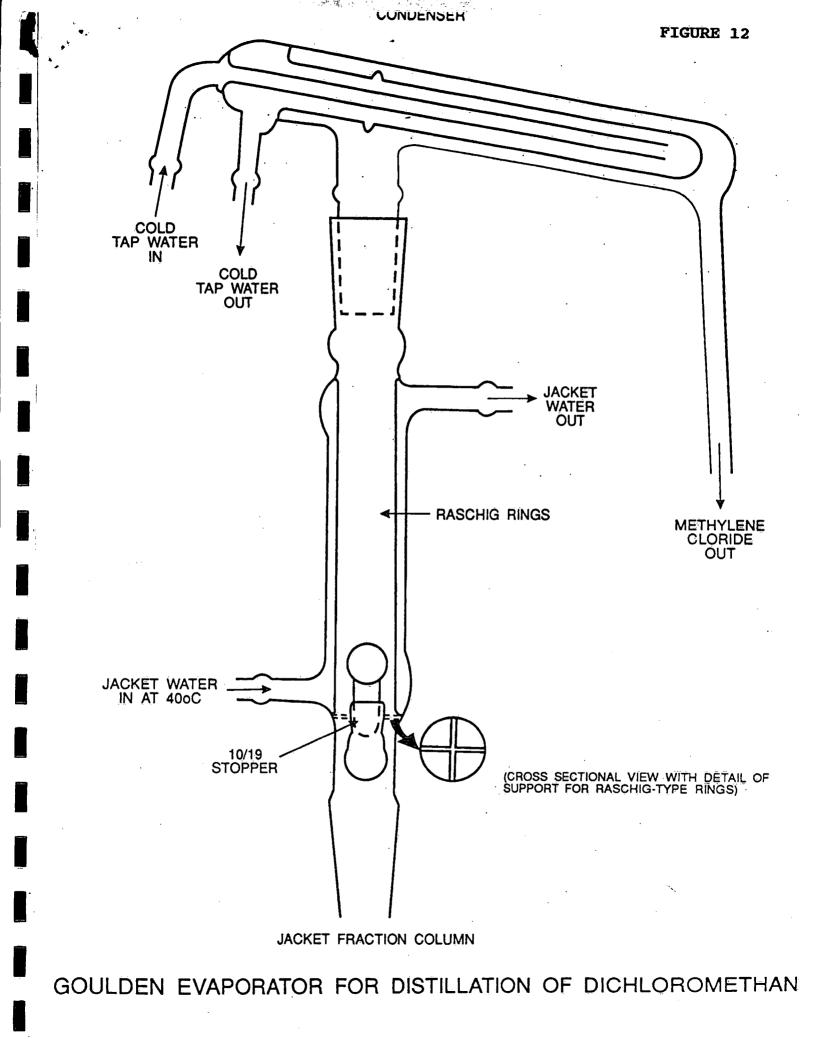


## Figure 10. Impeller Positioning

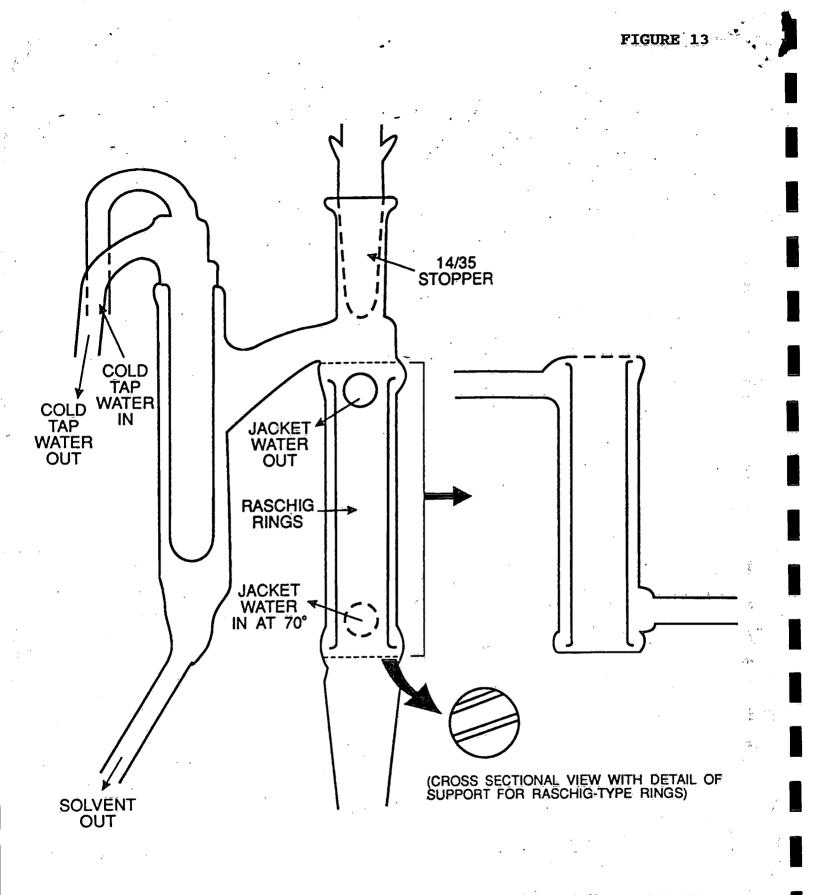


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GOULDEN EVAPORATOR FOR DISTILLATION OF HEXANE

