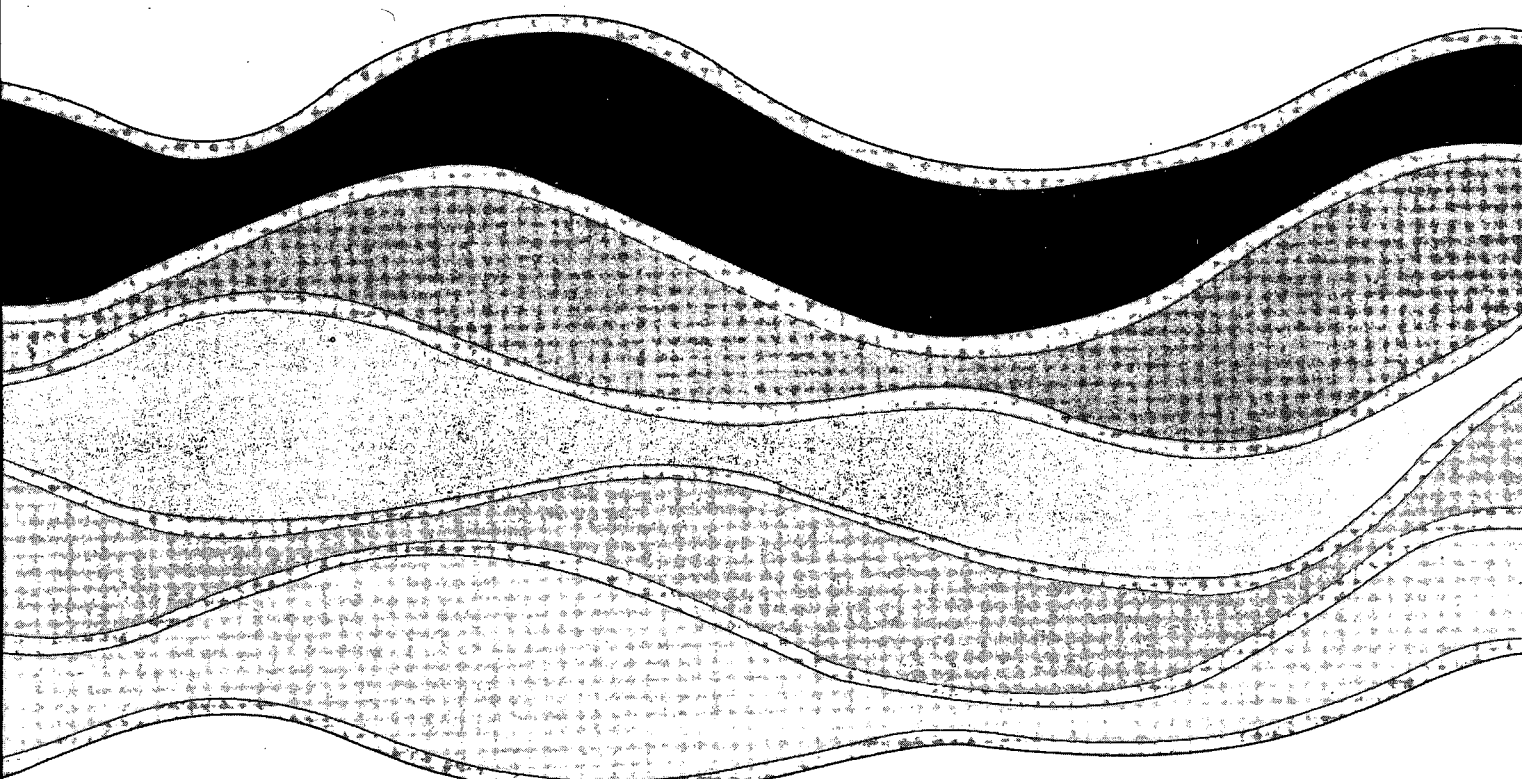


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CEPA NATIONAL INTERLABORATORY
COMPARISON STUDY (CP-4): ANALYSIS OF
DIOXINS AND FURANS IN SEDIMENT
EXTRACTS

W.C. Li and A.S.Y. Chau

NWRI CONTRIBUTION 94-84

**CEPA NATIONAL INTERLABORATORY COMPARISON STUDY
(CP-4): ANALYSIS OF DIOXINS AND FURANS IN
SEDIMENT EXTRACTS**

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NWRI Contribution No. 94-84

MANAGEMENT PERSPECTIVE

The highly chlorinated phenols used in pulp and paper industry across Canada contain dioxins as a manufacturing impurity. The most toxic form of dioxin is 2,3,7,8-TCDD. It has been detected in pulp and paper mill effluents. Two CEPA priority substances, dioxins and furans and effluents from pulp mills underwent CEPA assessments and were shown to be toxic as defined in the Act. These substances are presently undergoing a strategic options Process for possible regulatory control.

The successful implementation of the CEPA is dependent on the availability of reliable scientific data. To assist project managers and regulating bodies in ensuring the validity of analytical data under the Act, an interlaboratory study (CP-4) for the analysis of dioxins and furans in sediment extracts was designed and conducted. This study will help to establish the degree of comparability of dioxin and furan results among participating laboratories.

SOMMAIRE À L'INTENTION DE LA DIRECTION

Les phénols fortement chlorés utilisés par l'industrie des pâtes et papiers de tout le Canada contiennent des dioxines, à l'état d'impuretés de fabrication. La forme la plus toxique de dioxine est la 2,3,7,8-TCDD. On a décelé sa présence dans des effluents d'usines de pâtes et papiers. Deux substances de la liste prioritaire de la LCPE, les dioxines et les furanes, et des effluents des usines de pâtes, ont été évalués conformément à la LCPE, et il a été démontré que ces substances étaient toxiques aux termes de la Loi. Présentement, celles-ci font l'objet d'un processus d'options stratégiques pour d'éventuelles mesures réglementaires de limitation.

Le succès de la mise en oeuvre de la LCPE dépend de la disponibilité de données scientifiques fiables. Pour aider les gestionnaires de projets et les organismes de réglementation à garantir la validité des données analytiques conformément à la Loi, on a conçu et effectué une étude interlaboratoire (CP-4) pour l'analyse des dioxines et des furanes dans les extraits de sédiments. Cette étude contribuera à établir le degré de comparabilité des résultats des analyses de dioxines et de furanes entre les laboratoires participants.

ABSTRACT

As part of the quality assurance program under the auspices of the Canadian Environmental Protection Act (CEPA), an interlaboratory comparison study (CP-4) for analysis of dioxins and furans in sediment extracts was designed and conducted by the Quality Assurance Group at the National Water Research Institute. Eight laboratories were sent five test samples. Each laboratory was requested to analyze 2,3,7,8-TCDD and 2,3,7,8-TCDF, and tetra-, penta-, hexa-, hept-, and octachlorinated dibenzo-p-dioxins and dibenzofurans; each homologue group in total in all test samples. Surrogate recoveries were also requested. Seven out of eight laboratories submitted results.

Since design values of dioxins and furans in test samples were unknown, median values were used as target values for the evaluation of interlaboratory results. To estimate the quality of interlaboratory results generated by participating laboratories, comparison between means and medians for dioxins and furans in all five test samples was made. The majority of means and medians for dioxins and furans agreed with each other very well with the relative % difference within $\pm 25\%$.

For overall laboratory performance, five out of seven laboratories submitted satisfactory results for both of dioxins and furans in sediment extracts. Laboratory C030 had poor performance for both of dioxins and furans analyses.

RÉSUMÉ

Dans le cadre du programme d'assurance de qualité prévu par la Loi canadienne sur la protection de l'environnement (LCPE), le Groupe d'assurance de la qualité de l'Institut national de recherche sur les eaux a conçu et effectué une étude interlaboratoire comparative (CP-4) pour l'analyse des dioxines et des furanes dans des extraits de sédiments. On a envoyé cinq échantillons d'essai à huit laboratoires. Chaque laboratoire devait doser la 2,3,7,8-TCDD et le 2,3,7,8-TCDF, ainsi que les dibenzo-p-dioxines et les dibenzofuranes tétra-, penta-, hexa-, hepta- et octachlorés, soit chaque groupe d'homologues au complet dans tous les échantillons d'essai. On a également exigé les résultats des récupérations des substituts. Sept des huit laboratoires ont présenté des résultats.

Étant donné que les valeurs utilisées des dioxines et des furanes des échantillons d'essai étaient inconnues, on a pris des valeurs médianes comme valeurs cibles pour l'évaluation des résultats interlaboratoires. Afin d'évaluer la qualité de ces résultats fournis par les laboratoires participants, on a effectué des comparaisons entre les moyennes et les médianes des dioxines et des furanes pour chacun des cinq échantillons d'essai. Il y avait une très bonne concordance entre les moyennes et les médianes des dioxines et des furanes, avec une différence percentuelle relative inférieure à $\pm 25 \%$.

Pour ce qui du rendement de l'ensemble des laboratoires, cinq des sept laboratoires ont présenté des résultats satisfaisants tant pour les dioxines que pour les furanes dans les extraits de sédiments. Cependant, le rendement du laboratoire C030 laissait à désirer tant pour les analyses de dioxines que pour celles de furanes.

1 INTRODUCTION

The successful implementation of various aspects of the Canadian Environmental Protection Act (CEPA) is critically dependent on the availability of reliable scientific data. A key component of this CEPA QA program is to design and conduct, on a continual basis, a series of interlaboratory (Round Robin) studies for CEPA priority substances in a variety of matrices. These interlaboratory QA studies will assist CEPA managers and regulating bodies to ensure validity of analytical data.

In 1988, the Federal government initiated an emergency national sampling and analysis program [1] to monitor possible dioxins and furans contamination in the vicinity of Canadian pulp and paper mills using chlorine bleaching. To assist the managers in ensuring validity of analytical data, the Quality Assurance Group at the National Water Research Institute have designed and conducted several interlaboratory studies for analysis of dioxins and furans in sediments [2,3] and sediment extracts [4] to evaluate the comparability of the data generated by many different federal, provincial and private laboratories .

This CEPA interlaboratory study (No. CP-4) was distributed on February 17, 1994. The original deadline for reporting results was April 15, 1994. However, most laboratories were late in reporting, so the study was closed June 30, 1994. In August 26, 1994, a preliminary report was prepared and distributed to those participants which had submitted their results. The preliminary report allowed participants to compare their results with those of their peers. Thus any necessary corrective action could be taken in a timely manner. This final report provides information on the data summary as well as the data evaluation and laboratory performance of participants in this study.

2. STUDY DESIGN

This interlaboratory study (CP-4) for analysis of dioxins and furans in sediment

extracts was initiated in December, 1993. About 70 federal, provincial and private laboratories were invited to participate. From the returned questionnaires, eight laboratories expressed interest to participate in this study. By the time the study was closed, seven out of eight participants had submitted results. The list of participants is given in Table 1.

The study consists of five sediment extract samples for the analysis of dioxins and furans. The analytes of interest were 2,3,7,8-TCDD and 2,3,7,8-TCDF, and tetra-, penta-, hexa-, hepta-, and octachlorinated dibenzo-p-dioxins and dibenzofurans; each homologue group in total. Surrogates recoveries were also requested.

The identities and descriptions of the samples distributed in this study are given in Table 2. Briefly, the sediment extract SE-18 (samples #1) was prepared from freeze-dried sediment CRM EC-2 by soxhlet extraction using the method developed by Environment Canada [5]. This sample was used in the previous study CP-3 (samples #2 and #4). The recycle of this same sample in the present study allowed us for the evaluation of traceability of interlaboratory results in various studies and the stability of dioxins and furans in test samples. The sediment extracts SE-22 (samples #2 and 3) and SE-23 (samples #4 and 5) were prepared from bulk sediments EC-7 and EC-8a by the extraction procedure developed by Chau et. al. [6]. All the above extracts were sealed in ampules. Each ampule contained approximately 5 mL extracts in which one mL was equivalent to 1 g dry sediment. To assess reproducibility within the same laboratory, two pairs of blind duplicates were included as described earlier.

3. RESULTS AND DISCUSSION

3.1 Analytical Methodology

The participants were instructed to analyze the test samples using their in-house analytical procedure and calibration standards. The analytical procedures used by the participants in this study are presented in Table 3. All participants fortified or spiked

the extracts with various surrogate standards before cleanup procedures. In general, a wide variety of cleanup procedures and quantitation were used by different participants. For cleanup of raw extracts, column chromatography with silica gel, neutral or basic alumina, various carbon columns, or various combinations of these adsorbents were used. In all cases, the dioxins and furans fraction was evaporated to a small volume (10 to 20 μ L). Final analysis was performed by either GC/MSD or GC/MS (high resolution MS). For quantization of dioxins and furans in final extracts, either internal standard methods or external standard methods were used for calibrations. In most cases, correction for surrogate recoveries were made. See Table 3 for more details.

Reliable determination of dioxins and furans in environmental materials at trace and ultra-trace levels requires both high recoveries and final extracts that are free from any major interferences. In the report, "Internal Quality Assurance Requirements for the Analysis of Dioxins in Environmental Samples" [7], the Dioxin Quality Assurance Advisory Committee (DQAAC) recommended a sample size of 5 grams for dry sediment, soil, sludge or ash, and a final volume of 20 μ L for the final extract, in order to maximize capabilities for ultratrace analyses. Detection limits of dioxins and furans in sediment extracts for participating laboratories in this study are given in Table 4. In this table, the "Target MDLs" recommended by DQAAC are also included. These target method detection limits for low resolution mass spectrometry (LRMS) are based on an assumption of high surrogate recovery and final extracts are free from any major interferences (refer to reference 7 for further details). For those laboratories (C018, C020, C025 and C034) which employed high resolution MS for the detection of final extracts, their detection limits for their respective dioxins and furans were at least 10 times more sensitive, while the remaining laboratories which employed LRMS (MSD) for detection of respective dioxins and furans, their detection limit are in same order of magnitude as those of "Target MDLs".

Since sample size may be limited, the ability to analyze dioxins and furans at very low levels also requires that recoveries be as high as possible despite the need to employ

very stringent enrichment and cleanup steps to avoid major interferences for GC/MS analysis. The amount of analyte lost during cleanup as well as concentration steps may be reflected in the recoveries of the spiked surrogates. Thus results are usually corrected for surrogate recovery losses. A summary of surrogate recoveries reported by the participants for the five sediment extracts as well as their mean values for this study is given in Table 5. On the basis of the practical experience of several government and commercial laboratories, it was recommended that the acceptable range for surrogate recoveries from all matrices except biological tissues should be 30 - 130 % [8]. Beyond these limits, it was suggested that samples should be reprocessed and reanalyzed. As can be seen from Table 5, the majority of the reported surrogate recoveries were within this 30-130% range.

3.2 Data Evaluation

All data submitted by the participants for dioxins and furans in sediment extracts are summarized in Appendix I. One laboratory (C019) submitted data after the closing date. Their results were not included and evaluated in this final report but their data can be found in Appendix II as late results. All laboratories had the capability of analyzing all 2,3,7,8-TCDD and 2,3,7,8-TCDF congeners, and for each of the homologue group totals in all the samples. As shown in Appendix I, interlaboratory means and standard deviations as well as interlaboratory medians were calculated after outliers (marked with a *) were removed using Grubb's test [8]. With the rejections of these outliers, the majority of the means and medians for dioxins and furans agreed with each other very well. For the evaluation of interlaboratory results, medians were used as target values because true values were unknown and a panel of reference laboratories using proven bias-free methods was not available. A summary of interlaboratory median values for dioxins and furans in this study is given Table 6. To estimate the quality of the interlaboratory results generated by participating laboratories, comparison between means and medians was made for dioxins and furans for all five test samples. As can be seen from Fig. 1.1, the relative % differences between means and medians for the five test

samples and six dioxin parameters were within 25% except for samples #2, 3 and 4 for H6CDD which exceeded 25%. Where the relative % difference was expressed as $[(\text{mean} - \text{median})/(\text{mean} + \text{median})/2] \times 100$. Similarly, as shown in Fig. 1.2, the relative % difference between means and medians for the five test samples and six furan parameters were within 25% except for sample #2 for 2,3,7,8-TCDF and sample #1 for T4CDF which exceeded 25%. Overall, it indicated that comparable results for dioxins and furans had been generated by these participants in this study.

Interlaboratory precision for dioxins and furans, expressed as the relative standard deviation (RSD) is given in Table 7. For the analysis of dioxins and furans in sediment extracts at trace and ultratrace concentrations at ppt levels and in the presence of a large amount of co-extractives, the larger variations of analytical results were expected because of requiring to employ very stringent enrichment and cleanup steps to avoid major interferences for GC/MS analysis. Thus the interlaboratory results demonstrated favourable comparability among participating laboratories if the RSDs were within $\pm 50\%$. As can be seen from Fig. 2.1 for the interlaboratory precision of dioxins, only 3 out of 30 results (10%) had RSD exceeding $\pm 50\%$ (samples #1, 2 and 4 for H6CDD). Overall, the average values of RSDs for five out of six dioxin parameters were within $\pm 50\%$ except H6CDD. Furan results were less precise than the dioxin results. As can be seen from Fig. 2.2 for interlaboratory precision for furans, 9 out of 30 results (30.0%) had RSD exceeding $\pm 50\%$ (samples #2 for 2,3,7,8-TCDF, samples # 1, 3, 4 and 5 for T4CDF and P5CDF). Overall, the average values of RSDs for four out of six furan parameters were within $\pm 50\%$ with the exception of T4CDF and P5CDF which exceeded $\pm 50\%$. As compared with the previous study (CP-3) [4], the present study (CP-4) was less satisfactory with the higher RSDs for most of parameters of dioxins and furans.

Intralaboratory precision in this study was assessed by calculating RSD between the results provided by each participant for the two pairs of blind duplicates (i.e. SE-22 for samples #2 and #3 and SE-23 for samples #4 and #5). A summary of intralaboratory precision for participating laboratories is given in Table 8. The results show that four

laboratories (C018, C020, C025 and C034) had excellent precision for both of the two pairs of duplicate samples with RSDs less than $\pm 25\%$ for all dioxin and furan parameters. While three other laboratories (C003, C024 and C030) were less precise with some of dioxins and furan parameters exceeding $\pm 25\%$ RSD. In a few cases, the intralaboratory RSDs were higher than the interlaboratory RSDs from labs C003 and C024 as shown in Table 8. It is suggested that these above-mentioned two laboratories carefully review their internal QA/QC procedures.

3.3 Comparison of Laboratory Performance

For detailed data evaluation of each laboratory, submitted results were compared with the interlaboratory medians. As mentioned earlier, medians were used as target values because true values were unknown and results from a panel of reference laboratories using proven bias-free methods was not available. In addition, because of the small number of results available for this study, the Youden ranking technique [9] for the detection of bias as well as the computerize flagging procedure [10,11] were not used for data evaluation. Instead, a modified flagging procedure used in the national dioxin interlaboratory studies [2,3] and CEPA interlaboratory study [4] was employed in this study. This technique was a peer appraisal assessment, whereby the flags were assigned to the individual results when they deviated significantly from the interlaboratory median. Assuming that the medians had established the correct target values, the more accurate and comparable laboratories were therefore the ones with the least number of results flagged. Briefly, results within two-fold of the median for that particular parameter and sample, were deemed to be satisfactory and any values beyond this range were flagged. These ranges for the 'high' and 'low' flags were selected such that only the most extreme results would be flagged. Results recorded as "not detected" (ND) were not used for calculation of flags if the detection limits were higher than the medians. When the detection limits were lower than $1/2$ (Median), the ND results were flagged as low (L). Hence, the individual results were evaluated according to the following rating groups:

High (H)

 $x > 2$ (Median)

Satisfactory (no flags)

 $1/2$ (Median) $\leq x \leq 2$ (Median)

Low (L)

 $x < 1/2$ (Median)

The appraisal for flags for each individual result is listed in appendix I. Summaries of flags for dioxins and furans in sediment extracts for the study CP-4, obtained from appendix I, are given in Tables 9.1 and 9.2, respectively.

To compare the overall laboratory performance in this study, the key step was the selection of an appropriate performance index. The performance index used for this report was the % flags within a study. This index provides a simple way to evaluate laboratory performance through acceptance criteria which are shown below:

| <u>Performance Index</u> | <u>Rating</u> |
|--------------------------|---------------|
| $\leq 25\%$ | Satisfactory |
| 26% - 50% | Moderate |
| $\geq 51\%$ | Poor |

The performance index for each individual laboratory in this study is given in Tables 9.1 and 9.2 for dioxins and furans in sediment extracts, respectively. Four laboratories (C018, C020, C025 and C034) demonstrated satisfactory performance for both of dioxins and furans in sediment extracts. While performances of the other three laboratories (C003, C024 and C30) were less satisfactory especially lab C030 had provided poor rating for both of dioxins and furans in sediment extracts.

3.4 Comparison of Results between Studies and Samples

Two pairs of duplicate samples were included in this interlaboratory study for assessing reproducibility within the same laboratory as described earlier. In addition, overall interlaboratory results from these duplicate samples would provide the additional information on the homogeneity and integrity of the test samples. Comparisons of interlaboratory median values between samples for the two pairs of blind duplicates (SE-22 for sample #2 and #3, and SE-23 for samples #4 and #5) are given in Tables 10.1 and 10.2. As can be seen from Table 10.1, the RSDs for samples #2 and #3 (SE-22) were within $\pm 25\%$ for all twelve parameters of dioxins and furans analyses. While the RSDs for samples #4 and #5 (SE-23) were within $\pm 25\%$ for eleven out of 12 parameters of dioxins and furans analyses (Table 10.2). Only one parameter (T4CDD) was with the RSD exceeding $\pm 25\%$. Overall, the agreement between duplicate samples was very good and this helped to verify the integrity of the test samples. In this study, sample #1 was recycled from the previous study CP-3 (Samples #2 and #4). A comparison of study to study interlaboratory median values is given in Table 10.3. Overall the agreement between studies and samples was very good (RSDs within $\pm 25\%$) for all twelve parameters of dioxins and furans analyses, and this helped to verify the stability of test samples. In conclusion, the interlaboratory results from this study also provided very valuable preliminary reference values for dioxins and furans in these sediment extract reference materials.

ACKNOWLEDGEMENT

The authors are grateful to the participating laboratories for the time and effort devoted to analyze the test samples and reporting the results. This interlaboratory study would not be successful without their active participation and cooperation.

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Table 1. List of participants in CEPA interlaboratory study (CP-4).

1. Axys Analytical Services Ltd.
Sidney, B. C.
2. Eli Eco Logic International Inc.
Rockwood, Ontario
3. Gouvernement du Quebec
Minnistere de l'Environnement
Laval, Quebec
4. Research Productivity Council
Fredericton, N. B.
5. NovaMann International
Lachine, Quebec
6. Wellington Environmental Laboratories
Guelph, Ontario
7. Zenon Environmental Labs.
Burnaby, B. C.

Table 2. Samples distributed in study CP-4.

| Sample No. | Identification Code | Description |
|------------|---------------------|---|
| 1 | SE-18 | Soxhlet extraction of EC-2 (1 mL in toluene is equivalent to 1 g dry sediment) |
| 2 | SE-22 | 1: 1 mixture of solvent extraction of EC-7 and EC-8a* (1 mL in toluene is equivalent to 1 g dry sediment) |
| 3 | SE-22 | Same as sample #2 |
| 4 | SE-23 | Solvent extraction of EC-8a* (1 mL in toluene is equivalent to 1 g dry sediment) |
| 5 | SE-23 | Same as sample #4 |

Note: * EC-8 has not yet processed and homogenized. Samples (EC-8a) used for the preparation of SE-22 and SE-23 for this study were different from those samples (EC-8) used for the preparation of SE-20 (sample # 3) for study CP-3.

Table 3. Analytical Methodology used by participating Laboratories.

| Lab No. | Sample pretreatment | Cleanup | Separation/measurement |
|---------|--|---|---|
| C003 | Sediment extracts fortified with surrogate standards; solvent exchange to hexane and concentrated to 5 mL. | Column chromatograph with double silica gel H2504, silica gel/ NaOH, silica gel/silver nitrate column; alumina column. | GC/MSD, 60 m x 0.25 mm i.d. DB-5 fused silica column; ISTD; corrected for recovery of the surrogate standards. |
| C018 | Sediment extracts spiked with ¹³ C-labelled PCDDs and PCDFs surrogates; solvent exchange into nonane. | Multilayer silica column (acid and base-treated silica); alumina column (basic); carbon column (PK21/celite). | HRC/HRMS (VG70-SE high resolution mass Spec. coupled to HP 5890 GC); 60 m x 0.25 mm i.d. x 0.25 µm film thickness DB-5 capillary column; ISTD; recoveries of surrogates were accounted for all samples. |
| C020 | Sediment extracts fortified with surrogates. | Acid/Base/silver nitrate/silica column; Basic alumina column. | GC/MS analysis with HRMS (10000 static resolution); corrected for recoveries of surrogate standards. |
| C024 | Sediment extracts spiked with surrogates; back extraction with KOH (3.6M), NaCl (0.9M), sulphuric acid and again NaCl. | Alumina/acid silica gel column; carbon/celite column. | HRC/LRMS (HP 5890 series II GC /HP 5971A MSD); SIM mode; 30 m x 0.25 mm i.d. x 0.25 µm film thickness SPB-5 capillary column; ISTD; corrected for recoveries of the surrogates standards. |
| C025 | Sediment extracts spiked with surrogate standards; solvent exchange to hexane. | alumina column; acid/base/ silver nitrate/silica column. | GC/MS, HP 5890 series II GC interfaced with VG AutoSpec HRMS; multi-group selected ion recording (SIR); J & V DB-5 60 m x 0.25 mm i.d. x 0.25 µm film thickness; ISTD. |
| C030 | Sediment extracts spiked with surrogates. | Acid-treated Biosil-A column; mixed bed silica column (acid, neutral, basic Biosil-A); celite/activated carbon column. | GC/MSD, SIM Mode; ESTD; 60 m x 0.25 mm i.d. DB-5 column; corrected for recoveries of the surrogate standards. |
| C034 | Sediment extracts spiked with surrogates; rinsed with hexane three times; washed with KOH; washed with H2504. | Silica gel column; alumina column; carbon/celite; alumina column. | GC/MS (VG 70 SE high resolution MS with HP 5890 GC); MID Mode; corrected for recoveries of the surrogate standards. |

Table 4. Detection limits (pg/mL) of dioxins and furans in sediment extracts.

| Lab No. | Dioxins | | | | | | Furans | | | | | |
|----------------------|-----------|-------|-------|-------|-------|-------|-----------|-------|-------|-------|-------|-------|
| | 2378-TCDD | T4CDD | P5CDD | H6CDD | H7CDD | O8CDD | 2378-TCDF | T4CDF | P5CDF | H6CDF | H7CDF | O8CDF |
| C003 | 15-29 | 15-29 | 25-41 | 38-70 | 40-63 | 44-80 | 11-57 | 11-57 | 22-48 | 39-47 | 31-80 | 51-70 |
| C018 | 0.2 | 0.2 | 0.3 | 0.2 | 0.2 | 0.7 | 0.2 | 0.2 | 0.2 | 0.2 | 0.3 | 0.8 |
| C020 | 1-2 | 1-2 | 1-2 | 1-3 | 9-20 | 2-3 | 1-2 | 1-2 | 1-2 | 2-5 | 2-5 | 2-3 |
| C024 | 30 | 30 | 60 | 60 | 30 | 30 | 30 | 30 | 60 | 60 | 30 | 30 |
| C025 | 1 | 1 | 1 | 8 | 7 | 7 | 2 | 2 | 2 | 5 | 5 | 9 |
| C030 | 20 | 20 | 40 | 40 | 60 | 100 | 15 | 15 | 20 | 30 | 40 | 100 |
| C034 | 0.9 | 0.9 | 0.9 | 1.3 | 1.3 | 1.1 | 1.1 | 1.1 | 0.9 | 1.8 | 2.0 | 2.0 |
| Target MDLs for LRMS | - | 12 | 24 | 24 | 36 | 48 | - | 12 | 24 | 24 | 36 | 48 |

Table 5. Sample size, final volume and surrogate recoveries for study CP-4.

| Lab No. | Sample No. | Sample size (mL) | Final volume (uL) | ¹⁴ C-Dioxins | | | | | | ¹⁴ C-Furans | | | |
|---------|------------|------------------|-------------------|-------------------------|-------------|--------------|--------------|---------------|---------------|------------------------|-----------|-------------|---------------|
| | | | | 2376-TCDD | 12378-PSCDD | 123478-H4CDD | 123678-H6CDD | 1234789-H7CDD | 1234678-H7CDD | 08CDD | 2378-TCDF | 12378-PSCDF | 1234678-H7CDF |
| C003 | 1 | 2.9 | 20 | 100 | 100 | | 110 | | 110 | 100 | | | |
| | 2 | 4.8 | 20 | 120 | 110 | | 110 | | 120 | 110 | | | |
| | 3 | 4.8 | 20 | | 98 | | 99 | | 99 | 92 | | | |
| | 4 | 4.8 | 20 | 120 | 110 | | 120 | | 120 | 120 | | | |
| | 5 | 4.8 | 20 | 100 | 91 | | 96 | | 100 | 91 | | | |
| | Mean | | | 110 | 102 | | 107 | | 110 | 103 | | | |
| C018 | 1 | 2 | 40 | 82 | 92 | 84 | 75 | | 80 | 63 | 72 | 88 | 70 |
| | 2 | 2 | 40 | 82 | 86 | 95 | 84 | | 79 | 58 | 68 | 85 | 77 |
| | 3 | 2 | 40 | 84 | 82 | 87 | 86 | | 81 | 61 | 75 | 80 | 73 |
| | 4 | 2 | 40 | 89 | 97 | 91 | 86 | | 91 | 72 | 81 | 89 | 80 |
| | 5 | 2 | 40 | 87 | 94 | 95 | 86 | | 90 | 72 | 80 | 92 | 78 |
| | Mean | | | 85 | 90 | 90 | 83 | | 84 | 65 | 75 | 87 | 76 |
| C020 | 1 | | | 102 | 115 | | 108 | | 87 | 97 | 105 | 106 | 80 |
| | 2 | | | 103 | 107 | | 110 | | 89 | 93 | 105 | 103 | 86 |
| | 3 | | | 102 | 112 | | 110 | | 89 | 100 | 111 | 100 | 84 |
| | 4 | | | 97 | 115 | | 108 | | 92 | 102 | 95 | 107 | 87 |
| | 5 | | | 101 | 109 | | 120 | | 93 | 93 | 101 | 100 | 86 |
| | Mean | | | 101 | 112 | | 111 | | 90 | 97 | 103 | 103 | 85 |
| C024 | 1 | 4 | 20 | 86 | 45 | 22 | | | 83 | 78 | | | |
| | 2 | 4 | 20 | 76 | 82 | 64 | | | 70 | 68 | | | |
| | 3 | 4 | 20 | 78 | 48 | 39 | | | 80 | 73 | | | |
| | 4 | 4 | 20 | 85 | 82 | 42 | | | 86 | 75 | | | |
| | 5 | 4 | 20 | 67 | 27 | 17 | | | 63 | 60 | | | |
| | Mean | | | 78 | 57 | 37 | | | 76 | 71 | | | |

Table 5. Sample size, final volume and surrogate recoveries for study CP-4 (continued).

| Lab No. | Sample No. | Sample size (mL) | Final volume (mL) | ¹⁴ C-Dioxins | | | | | | ¹⁴ C-Furans | | | |
|---------|------------|------------------|-------------------|-------------------------|-------------|--------------|--------------|---------------|---------------|------------------------|-----------|-------------|---------------|
| | | | | 2378-TCDD | 12378-FSCDD | 123478-H6CDD | 123678-H6CDD | 1234789-H7CDD | 1234678-H7CDD | 08CDD | 2378-TCDF | 12378-FSCDF | 1234678-H7CDF |
| C025 | 1 | 1.0 | 20 | 92 | 91 | 94 | 95 | | 100 | 109 | 82 | 87 | 100 |
| | 2 | 1.0 | 20 | 95 | 104 | 93 | 91 | | 100 | 109 | 100 | 97 | 100 |
| | 3 | 1.0 | 20 | 88 | 97 | 93 | 86 | | 93 | 104 | 89 | 90 | 93 |
| | 4 | 1.0 | 20 | 94 | 105 | 106 | 97 | | 105 | 113 | 92 | 90 | 105 |
| | 5 | 1.0 | 20 | 99 | 99 | 110 | 98 | | 108 | 122 | 95 | 89 | 116 |
| | Mean | | | 94 | 99 | 99 | 93 | | 101 | 111 | 92 | 91 | 103 |
| C030 | 1 | 1 | 20 | 83 | 91 | | 72 | | 68 | 46 | 107 | | |
| | 2 | 1 | 20 | 70 | 82 | | 70 | | 67 | 45 | 69 | | |
| | 3 | 1 | 20 | 94 | 101 | | 80 | | 72 | 47 | 107 | | |
| | 4 | 1 | 20 | 78 | 89 | | 73 | | 66 | 46 | 94 | | |
| | 5 | 1 | 20 | 84 | 88 | | 73 | | 66 | 50 | 93 | | |
| | Mean | | | 82 | 90 | | 74 | | 68 | 47 | 94 | | |
| C034 | 1 | 1 | 20 | 113 | 112 | | 83 | | 102 | 82 | 97 | 95 | 82 |
| | 2 | 1 | 20 | 95 | 102 | | 70 | | 74 | 42 | 84 | 86 | 60 |
| | 3 | 1 | 20 | 107 | 115 | | 79 | | 82 | 56 | 88 | 100 | 63 |
| | 4 | 1 | 20 | 101 | 99 | | 80 | | 82 | 61 | 84 | 90 | 63 |
| | 5 | 1 | 20 | 104 | 107 | | 78 | | 87 | 67 | 85 | 94 | 68 |
| | Mean | | | 104 | 107 | | 78 | | 85 | 62 | 88 | 93 | 67 |

Table 6. Summary of interlaboratory median values for study CP-4

| Parameter | Concentration | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|---------------|-----------|-----------|-----------|-----------|-----------|
| Dioxins | | | | | | |
| 2378-TCDD | pg/mL | 281.5 | 50 | 47.9 | 95.7 | 100.5 |
| T4CDD | pg/mL | 422 | 83 | 93 | 155 | 225 |
| P8CDD | pg/mL | 195 | 69.5 | 61 | 130 | 126 |
| H6CDD | pg/mL | 715 | 245 | 229 | 468 | 451 |
| H7CDD | pg/mL | 1075 | 369.5 | 437 | 717.5 | 755 |
| O8CDD | pg/mL | 4025 | 1300 | 1300 | 2355 | 2400 |
| Furans | | | | | | |
| 2378-TCDF | pg/mL | 110 | 40 | 41.7 | 70.2 | 71 |
| T4CDF | pg/mL | 829 | 170 | 203 | 290 | 371.5 |
| P8CDF | pg/mL | 751 | 201 | 234 | 409 | 403 |
| H6CDF | pg/mL | 2250 | 615.5 | 625 | 1250 | 1250 |
| H7CDF | pg/mL | 4060 | 1225 | 1140 | 2280 | 2385 |
| O8CDF | pg/mL | 7650 | 1750 | 1740 | 3300 | 3450 |

Table 7. Summary of Interlaboratory precision (%RSD) for study CP-4.

| Parameter | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 | Average |
|-----------|-----------|-----------|-----------|-----------|-----------|---------|
| Dioxins: | | | | | | |
| 2378-TCDD | 21.2 | 10.1 | 13.2 | 12.6 | 14.5 | 14.3 |
| TCDD | 26.8 | 36.4 | 24.2 | 40.6 | 30.1 | 31.6 |
| PCDD | 46.8 | 22.9 | 25.8 | 37.3 | 24.1 | 31.4 |
| HxCDD | 50.8 | 79.1 | 48.1 | 75.7 | 31.2 | 57.0 |
| HxCDD | 24.2 | 28.8 | 21.7 | 21.8 | 33.2 | 25.9 |
| OCDD | 8.1 | 10.7 | 23.8 | 10.1 | 18.7 | 14.3 |
| Furans: | | | | | | |
| 2378-TCDF | 39.8 | 56.9 | 36.7 | 43.6 | 32.8 | 42.0 |
| TCDF | 56.0 | 42.4 | 63.8 | 65.5 | 57.6 | 57.1 |
| PCDF | 61.0 | 36.9 | 51.2 | 33.6 | 38.3 | 52.2 |
| HxCDF | 16.0 | 24.6 | 40.3 | 19.2 | 18.4 | 23.7 |
| HxCDF | 12.0 | 12.7 | 26.8 | 29.0 | 33.9 | 22.9 |
| OCDF | 10.1 | 18.2 | 13.1 | 12.1 | 10.7 | 12.8 |

Table 8. Summary of intralaboratory precision (%RSD) for study CP-4.

| Parameter | C003 | | | C018 | | | C020 | | | C024 | | | C028 | | | C030 | | | C034 | | |
|-----------|-------|-------|-------|-------|-------|------|-------|-------|------|-------|-------|-------|-------|-------|------|-------|-------|------|-------|-------|------|
| | SE-23 | SE-23 | AVG. | SE-23 | SE-23 | AVG. | SE-23 | SE-23 | AVG. | SE-23 | SE-23 | AVG. | SE-23 | SE-23 | AVG. | SE-23 | SE-23 | AVG. | SE-23 | SE-23 | AVG. |
| Dioxins: | | | | | | | | | | | | | | | | | | | | | |
| 2378-TCDD | 141.4 | 38.8 | 98.1 | 0 | 4.8 | 2.4 | 0 | 2.3 | 1.1 | 12.9 | 6.1 | 9.5 | 5.0 | 1.6 | 3.3 | 5.7 | 9.4 | 7.5 | 4.4 | 0.7 | 2.5 |
| T4CDD | 141.4 | 83.9 | 112.7 | 4.1 | 6.3 | 5.3 | 8.1 | 0 | 4.1 | 12.9 | 6.1 | 9.5 | 10.7 | 1.9 | 6.3 | 5.7 | 8.3 | 7.0 | 9.4 | 1.6 | 5.5 |
| P5CDD | 141.4 | 141.4 | 141.4 | 6.1 | 7.6 | 6.8 | 5.7 | 0 | 2.8 | - | 141.4 | 141.4 | 3.4 | 2.9 | 3.1 | 5.1 | 141.4 | 75.3 | 5.3 | 0 | 2.6 |
| H6CDD | 0 | 58.4 | 29.2 | 0 | 0.6 | 0.3 | 3.0 | 1.7 | 2.3 | 47.1 | 7.4 | 27.3 | 6.0 | 1.6 | 3.8 | 32.6 | 29.8 | 31.2 | 6.0 | 0.8 | 3.4 |
| H7CDD | 11.3 | 14.6 | 12.9 | 1.0 | 1.5 | 1.2 | 3.8 | 4.1 | 4.0 | 22.5 | 48.4 | 33.9 | 6.3 | 2.7 | 4.5 | 5.7 | 17.7 | 11.7 | 2.1 | 1.1 | 1.6 |
| O8CDD | 0 | 6.4 | 3.3 | 0 | 0.6 | 0.3 | 5.7 | 0.6 | 3.1 | 12.9 | 16.3 | 14.5 | 4.4 | 1.0 | 2.7 | 0 | 10.1 | 5.1 | 3.0 | 3.3 | 3.2 |
| Furans: | | | | | | | | | | | | | | | | | | | | | |
| 2378-TCDF | 39.5 | 61.8 | 90.6 | 8.1 | 1.3 | 4.7 | 0 | 6.8 | 3.4 | 141.4 | 0 | 70.7 | 4.9 | 0 | 2.5 | 27.7 | 0 | 13.9 | 0.9 | 1.0 | 1.0 |
| T4CDF | 64.5 | 93.6 | 79.0 | 2.5 | 4.6 | 3.6 | 0 | 1.3 | 0.6 | 141.4 | 0 | 70.7 | 5.9 | 6.4 | 6.3 | 17.2 | 32.6 | 24.9 | 0 | 0 | 0 |
| P5CDF | 27.8 | 3.1 | 15.5 | 8.5 | 4.1 | 6.3 | 4.6 | 12.9 | 8.7 | - | 76.1 | 76.1 | 10.4 | 1.0 | 5.7 | 39.0 | 33.6 | 36.3 | 1.1 | 1.7 | 1.4 |
| H6CDF | 7.5 | 5.7 | 6.6 | 3.5 | 0.5 | 2.0 | 8.9 | 1.1 | 5.0 | 141.4 | 110.6 | 124.0 | 3.8 | 1.9 | 2.9 | 5.7 | 34.6 | 20.1 | 2.9 | 0 | 1.5 |
| H7CDF | 21.8 | 13.5 | 17.6 | 4.1 | 3.9 | 4.0 | 10.4 | 5.8 | 8.1 | 42.8 | 24.6 | 33.7 | 3.3 | 1.0 | 2.3 | 0 | 10.3 | 5.1 | 2.8 | 2.9 | 2.9 |
| O8CDF | 3.8 | 4.3 | 4.1 | 8.4 | 1.8 | 5.1 | 6.8 | 0 | 3.4 | 4.0 | 2.0 | 3.0 | 4.5 | 2.4 | 3.5 | 1.9 | 9.2 | 5.6 | 2.3 | 2.3 | 2.3 |

Table 9.1 **Performance of individual laboratory for dioxins in sediment extracts.**

| Lab No. | Total No. of Results Reported | No. of Results "not Detected" | No. of Results Ranked | No. of Results Flagged | | % Flags (Performance Index) | Comment |
|---------|-------------------------------|-------------------------------|-----------------------|------------------------|---|-----------------------------|--------------|
| | | | | H | L | | |
| C003 | 30 | 4 | 28 | 5 | 3 | 28.6 | Moderate |
| C018 | 30 | 0 | 30 | 0 | 0 | 0 | Satisfactory |
| C020 | 30 | 0 | 30 | 0 | 0 | 0 | Satisfactory |
| C024 | 30 | 4 | 28 | 2 | 2 | 14.3 | Satisfactory |
| C025 | 30 | 0 | 30 | 0 | 0 | 0 | Satisfactory |
| C030 | 30 | 1 | 30 | 16 | 1 | 56.7 | Poor |
| C034 | 30 | 0 | 30 | 0 | 0 | 0 | Satisfactory |

Table 9.2 Performance of individual laboratory for furans in sediment extracts.

| Lab No. | Total No. of Results Reported | No. of Results "not Detected" | No. of Results Ranked | No. of Results Flagged | | | % Flags (Performance Index) | Comment |
|---------|-------------------------------|-------------------------------|-----------------------|------------------------|----|---|-----------------------------|--------------|
| | | | | H | L | L | | |
| C003 | 30 | 0 | 30 | 0 | 4 | | 13.3 | Satisfactory |
| C018 | 30 | 0 | 30 | 0 | 0 | | 0 | Satisfactory |
| C020 | 30 | 0 | 30 | 0 | 0 | | 0 | Satisfactory |
| C024 | 30 | 6 | 29 | 1 | 16 | | 58.6 | Poor |
| C025 | 30 | 0 | 30 | 0 | 1 | | 3.3 | Satisfactory |
| C030 | 30 | 0 | 30 | 19 | 0 | | 63.3 | Poor |
| C034 | 30 | 0 | 30 | 0 | 0 | | 0 | Satisfactory |

Table 10.1. Comparison of interlaboratory median values between duplicate samples (SE-22).

| Parameter | Concentration | Sample No. | | Average Median | S.D. | RSD, % |
|-----------|---------------|------------|------|----------------|------|--------|
| | | 2 | 3 | | | |
| Dioxins: | | | | | | |
| 2378-TCDD | pg/mL | 50 | 47.9 | 49.0 | 1.5 | 3.0 |
| T4CDD | pg/mL | 83 | 93 | 88 | 7.1 | 8.0 |
| P5CDD | pg/mL | 69.5 | 61 | 65.3 | 6.0 | 9.2 |
| H6CDD | pg/mL | 245 | 229 | 237 | 11.3 | 4.8 |
| H7CDD | pg/mL | 369.5 | 437 | 403.3 | 47.7 | 11.8 |
| O8CDD | pg/mL | 1300 | 1300 | 1300 | 0 | 0 |
| Furans: | | | | | | |
| 2378-TCDF | pg/mL | 40 | 41.7 | 40.9 | 1.2 | 2.9 |
| T4CDF | pg/mL | 170 | 203 | 186.5 | 23.3 | 12.5 |
| P5CDF | pg/mL | 201 | 234 | 217.5 | 23.3 | 10.7 |
| H6CDF | pg/mL | 615.5 | 625 | 620.3 | 6.7 | 1.1 |
| H7CDF | pg/mL | 1225 | 1140 | 1182.5 | 60.1 | 5.1 |
| O8CDF | pg/mL | 1750 | 1740 | 1745 | 7.1 | 0.4 |

Table 10.2. Comparison of interlaboratory median values between duplicate samples (SE-23).

| Parameter | Concentration | Sample No. | | Average Median | S.D. | RSD, % |
|-----------|---------------|------------|-------|----------------|-------|--------|
| | | 4 | 5 | | | |
| Dioxins: | | | | | | |
| 2378-TCDD | pg/mL | 95.7 | 100.5 | 98.1 | 3.4 | 3.5 |
| T4CDD | pg/mL | 155 | 225 | 190 | 49.5 | 26.1 |
| P5CDD | pg/mL | 130 | 126 | 128 | 2.8 | 2.2 |
| H6CDD | pg/mL | 468 | 451 | 459.5 | 12.0 | 2.6 |
| H7CDD | pg/mL | 717.5 | 755 | 736.3 | 26.5 | 3.6 |
| O8CDD | pg/mL | 2355 | 2400 | 2377.5 | 31.8 | 1.3 |
| Furans: | | | | | | |
| 2378-TCDF | pg/mL | 70.2 | 71 | 70.6 | 0.6 | 0.8 |
| T4CDF | pg/mL | 290 | 371.5 | 330.8 | 57.6 | 17.4 |
| P5CDF | pg/mL | 409 | 403 | 406 | 4.2 | 1.0 |
| H6CDF | pg/mL | 1250 | 1250 | 1250 | 0 | 0 |
| H7CDF | pg/mL | 2280 | 2385 | 2332.5 | 74.2 | 3.2 |
| O8CDF | pg/mL | 3300 | 3450 | 3375 | 106.1 | 3.1 |

Table 10.3. Comparison of interlaboratory median values between studies (SE-18).

| Parameter | Study No. | CP-3 | CP-3 | CP-4 | Average Median | S.D. | RSD, % |
|-----------|------------|-------|------|-------|----------------|-------|--------|
| | Sample No. | 2 | 4 | 1 | | | |
| Dioxins: | | | | | | | |
| 2378-TCDD | pg/mL | 290 | 285 | 281.5 | 285.5 | 4.3 | 1.5 |
| T4CDD | pg/mL | 396 | 380 | 422 | 399.3 | 21.2 | 5.3 |
| P5CDD | pg/mL | 202 | 241 | 195 | 212.7 | 24.8 | 11.7 |
| H6CDD | pg/mL | 730 | 760 | 715 | 735 | 22.9 | 3.1 |
| H7CDD | pg/mL | 1200 | 1150 | 1075 | 1141.7 | 62.9 | 5.5 |
| O8CDD | pg/mL | 3800 | 4000 | 4025 | 3941.7 | 123.3 | 3.1 |
| Furans: | | | | | | | |
| 2378-TCDF | pg/mL | 100 | 110 | 110 | 106.7 | 5.8 | 5.4 |
| T4CDF | pg/mL | 640 | 634 | 829 | 701 | 110.9 | 15.8 |
| P5CDF | pg/mL | 778.5 | 1050 | 751 | 859.8 | 165.3 | 19.2 |
| H6CDF | pg/mL | 2146 | 2051 | 2250 | 2149 | 99.5 | 4.6 |
| H7CDF | pg/mL | 3460 | 3400 | 4060 | 3640 | 365.0 | 10.0 |
| O8CDF | pg/mL | 6250 | 7000 | 7650 | 6966.7 | 700.6 | 10.1 |

Fig.1.1. Comparison between Mean and Median for Dioxins

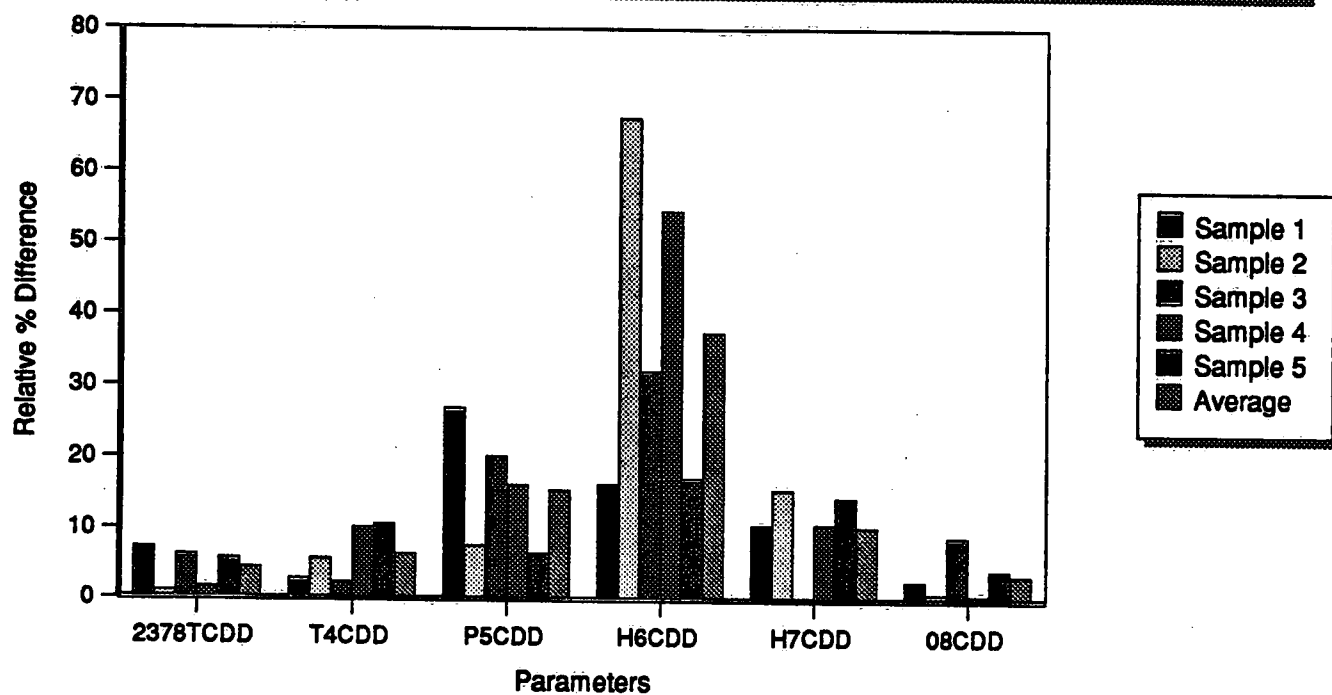


Fig. 1.2. Comparison between Mean and Median for Furans

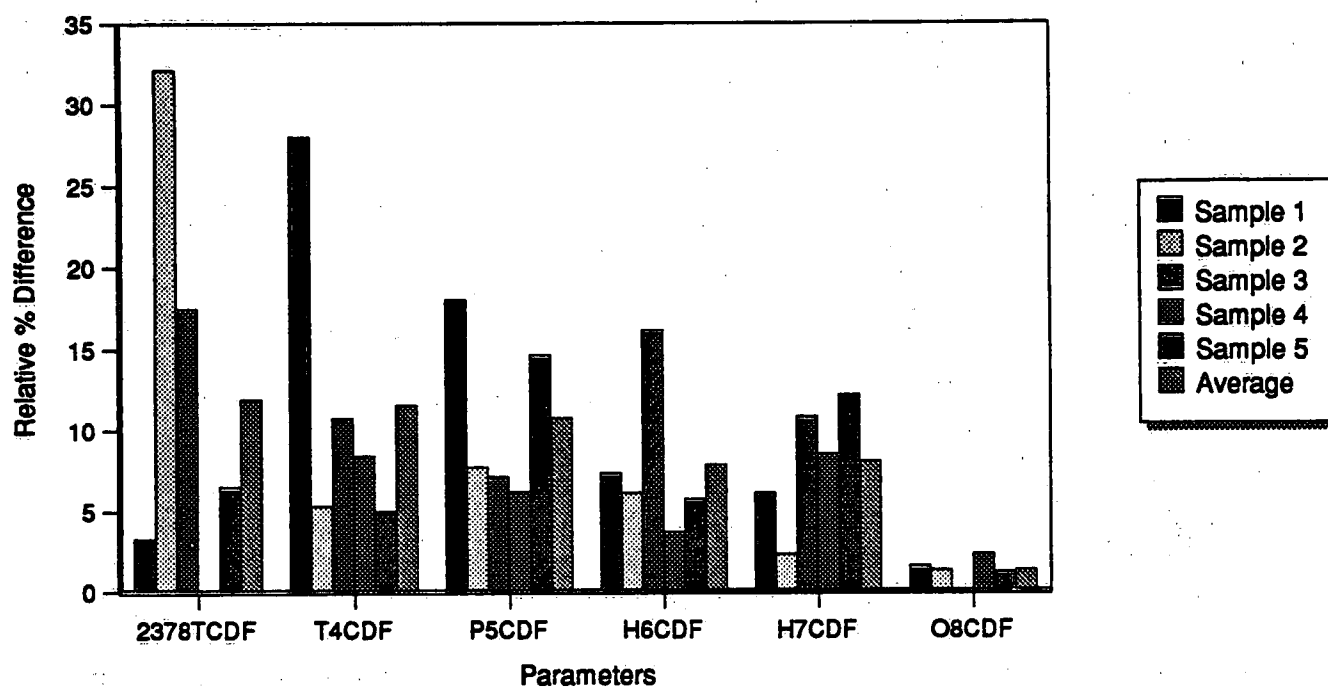


Fig. 2.1. Interlaboratory Precision for Dioxins

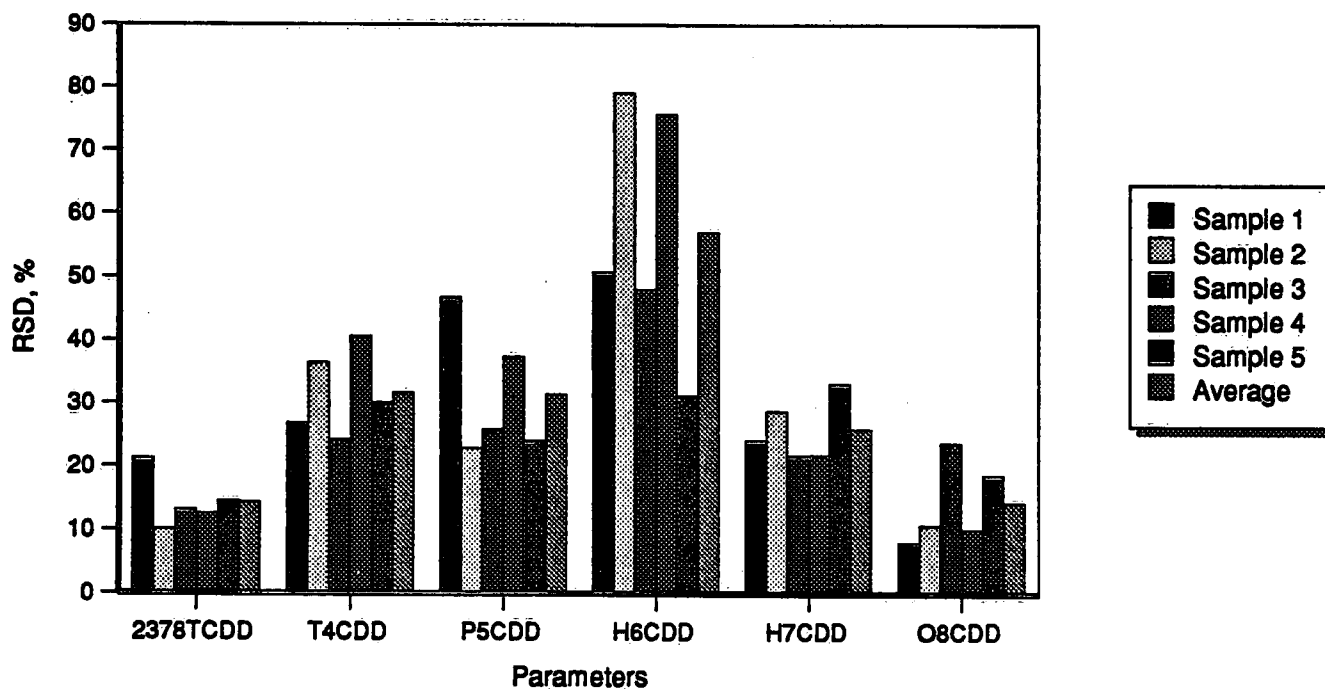
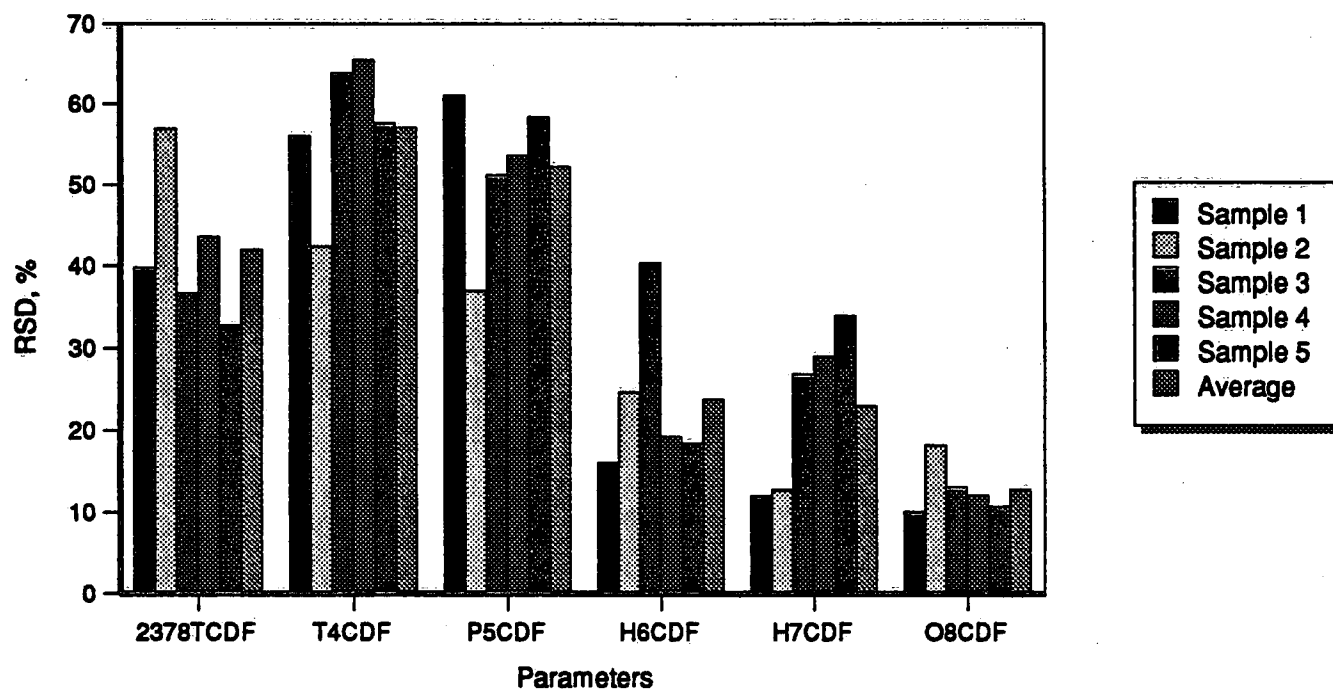


Fig. 2.2. Interlaboratory Precision for Furans



APPENDIX I
DATA SUMMARY

Table I-1. Results for 2378-TCDD in sediment extracts (pg/mL).

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 430 | 52 | ND | 74 | 130 |
| C018 | 273 | 47.9 | 47.9 | 94.4 | 101 |
| C020 | 290 | 56 | 56 | 97 | 100 |
| C024 | 300 | 50 | 60 | 110 | 120 |
| C025 | 254 | 41 | 44 | 90 | 88 |
| C030 | * 620 H | * 130 H | * 120 H | * 210 H | * 240 H |
| C034 | 270 | 50 | 47 | 99 | 100 |
| MEAN | 302.8 | 49.5 | 51.0 | 94.1 | 106.5 |
| S.D. | 64.3 | 5.0 | 6.7 | 11.9 | 15.4 |
| MEDIAN | 281.5 | 50 | 47.9 | 95.7 | 100.5 |

Table I-2. Results for T4CDD in sediment extracts (pg/mL).

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | * 960 H | 52 | ND L | 74 L | 290 |
| C018 | 464 | 107 | 101 | 215 | 235 |
| C020 | 380 | 83 | 74 | 155 | 155 |
| C024 | 300 | 50 | 60 | 110 | 120 |
| C025 | 347 | 73 | 85 | 148 | 152 |
| C030 | 620 | 130 | 120 | 270 | 240 |
| C034 | 495 | 120 | 105 | 230 | 225 |
| MEAN | 434.3 | 87.9 | 90.8 | 171.7 | 202.4 |
| S.D. | 116.3 | 32.0 | 22.0 | 69.7 | 60.9 |
| MEDIAN | 422 | 83 | 93 | 155 | 225 |

Table I-3. Results for P5CDD in sediment extracts (pg/mL) .

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 450 H | 74 | ND | 250 | ND L |
| C018 | 155 | 55.8 | 60.8 | 95.2 | 106 |
| C020 | 195 | 65 | 60 | 130 | 130 |
| C024 | ND L | ND | ND | ND L | 120 |
| C025 | 188 | 64 | 61 | 121 | 126 |
| C030 | * 820 H | 100 | 93 | 130 | ND L |
| C034 | 290 | 91 | 98 | 190 | 190 |
| MEAN | 255.6 | 75.0 | 74.6 | 152.7 | 134.4 |
| S.D. | 119.7 | 17.2 | 19.2 | 56.9 | 32.4 |
| MEDIAN | 195 | 69.5 | 61 | 130 | 126 |

Table I-4. Results for H6CDD in sediment extracts (pg/mL) .

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 1700 H | 470 | 470 H | 1900 H | 790 |
| C018 | 753 | 228 | 228 | 468 | 472 |
| C020 | 700 | 240 | 230 | 420 | 430 |
| C024 | 730 | 1100 H | 550 H | 630 | 700 |
| C025 | 570 | 181 | 197 | 385 | 394 |
| C030 | * 3100 H | 1000 H | * 1600 H | 1500 H | * 2300 H |
| C034 | 595 | 245 | 225 | 420 | 425 |
| MEAN | 841.3 | 494.9 | 316.7 | 817.6 | 535.2 |
| S.D. | 427.0 | 391.4 | 152.3 | 618.9 | 166.9 |
| MEDIAN | 715 | 245 | 229 | 468 | 451 |

Table I-5. Results for H7CDD in sediment extracts (pg/mL) .

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 1600 | 610 | 520 | 960 | 780 |
| C018 | 1100 | 369 | 364 | 715 | 730 |
| C020 | 1520 | 570 | 540 | 1070 | 1010 |
| C024 | 950 | 370 | 510 | 720 | 1400 |
| C025 | 1050 | 327 | 357 | 711 | 684 |
| C030 | * 3200 H | * 1200 H | * 1300 H | * 2100 H | * 2700 H |
| C034 | 955 | 345 | 335 | 620 | 630 |
| MEAN | 1195.8 | 431.8 | 437.7 | 799.3 | 872.3 |
| S.D. | 288.9 | 124.2 | 94.8 | 174.6 | 290.0 |
| MEDIAN | 1075 | 369.5 | 437 | 717.5 | 755 |

Table I-6. Results for O8CDD in sediment extracts (pg/mL) .

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 4100 | 1300 | 1300 | 2100 | 2300 |
| C018 | 4100 | 1350 | 1350 | 2510 | 2530 |
| C020 | 3950 | 1300 | 1200 | 2520 | 2500 |
| C024 | 4300 | 1500 | 1800 | 2700 | 3400 |
| C025 | 3640 | 1090 | 1160 | 2180 | 2210 |
| C030 | * 5500 | * 2000 | 2000 | * 3900 | * 4500 |
| C034 | 3450 | 1200 | 1150 | 2200 | 2100 |
| MEAN | 3923.3 | 1290 | 1422.9 | 2368.3 | 2506.7 |
| S.D. | 319.2 | 138.6 | 338.9 | 240.4 | 467.9 |
| MEDIAN | 4025 | 1300 | 1300 | 2355 | 2400 |

Table I-7. Results for 2378-TCDF in sediment extracts (pg/mL).

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 130 | 40 | 71 | 29 L | 74 |
| C018 | 113 | 36.9 | 41.4 | 70.2 | 71.4 |
| C020 | 110 | 42 | 42 | 76 | 69 |
| C024 | 60 | 90 H | ND | 60 | 60 |
| C025 | 73 | 28 | 30 | 55 | 55 |
| C030 | 200 | 110 H | 74 | 130 | 130 |
| C034 | 110 | 40 | 39.5 | 72 | 71 |
| MEAN | 113.7 | 55.3 | 49.7 | 70.3 | 75.8 |
| S.D. | 45.2 | 31.4 | 18.2 | 30.7 | 24.9 |
| MEDIAN | 110 | 40 | 41.7 | 70.2 | 71 |

Table I-8. Results for T4CDF in sediment extracts (pg/mL).

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 1000 | 190 | 71 L | 120 L | 590 |
| C018 | 829 | 247 | 256 | 429 | 458 |
| C020 | 420 | 150 | 150 | 290 | 285 |
| C024 | 60 L | 90 | ND L | 60 L | 60 L |
| C025 | 343 L | 114 | 124 | 189 | 207 |
| C030 | 880 | * 600 H | 470 H | 600 H | 960 H |
| C034 | 845 | 285 | 285 | 520 | 520 |
| MEAN | 625.3 | 179.3 | 226 | 315.4 | 353.3 |
| S.D. | 350.3 | 76.1 | 144.3 | 206.5 | 203.4 |
| MEDIAN | 829 | 170 | 203 | 290 | 371.5 |

Table I-9. Results for P5CDF in sediment extracts (pg/mL).

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 1100 | 140 | 94 L | 230 | 220 |
| C018 | 751 | 265 | 235 | 447 | 474 |
| C020 | 560 | 150 | 160 | 300 | 360 |
| C024 | 150 L | ND L | ND L | 60 L | 200 L |
| C025 | 736 | 201 | 233 | 409 | 403 |
| C030 | 1900 H | * 810 H | 460 | 610 | 990 H |
| C034 | 1100 | 330 | 325 | 635 | 620 |
| MEAN | 899.6 | 217.2 | 251.2 | 384.4 | 466.7 |
| S.D. | 548.8 | 80.2 | 128.6 | 205.9 | 272.3 |
| MEDIAN | 751 | 201 | 234 | 409 | 403 |

Table I-10. Results for H6CDF in sediment extracts (pg/mL) .

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 2400 | 710 | 790 | 1200 | 1300 |
| C018 | 2250 | 641 | 610 | 1420 | 1430 |
| C020 | 2310 | 720 | 635 | 1300 | 1320 |
| C024 | ND L | 380 | ND L | * 110 L | 900 |
| C025 | 1600 | 434 | 458 | 960 | 934 |
| C030 | * 5800 H | * 1200 | 1300 H | 1700 | * 2800 H |
| C034 | 1900 | 590 | 615 | 1200 | 1200 |
| MEAN | 2092 | 579.2 | 734.7 | 1296.7 | 1180.7 |
| S.D. | 334.0 | 142.6 | 296.3 | 249.0 | 217.2 |
| MEDIAN | 2250 | 615.5 | 625 | 1250 | 1250 |

Table I-11. Results for H7CDF in sediment extracts (pg/mL) .

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 4100 | 1200 | 880 | 1900 | 2300 |
| C018 | 4070 | 1250 | 1180 | 2450 | 2590 |
| C020 | 4150 | 1390 | 1200 | 2680 | 2470 |
| C024 | * 560 L | 990 | 530 L | 980 L | 690 L |
| C025 | 3540 | 1050 | 1100 | 2160 | 2130 |
| C030 | 3000 | * 3000 H | * 3000 H | * 5800 H | * 6700 H |
| C034 | 4050 | 1300 | 1250 | 2400 | 2500 |
| MEAN | 3818.3 | 1196.7 | 1023.3 | 2095 | 2113.3 |
| S.D. | 459.0 | 151.7 | 274.6 | 607.7 | 716.2 |
| MEDIAN | 4060 | 1225 | 1140 | 2280 | 2385 |

Table I-12. Results for O8CDF in sediment extracts (pg/mL) .

| Lab No. | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 |
|----------------|------------------|------------------|------------------|------------------|------------------|
| C003 | 7900 | 1800 | 1900 | 3200 | 3400 |
| C018 | 8610 | 2330 | 2070 | 4080 | 3980 |
| C020 | 7500 | 1850 | 1680 | 3550 | 3550 |
| C024 | 7800 | 1700 | 1800 | 3400 | 3500 |
| C025 | 6590 | 1360 | 1450 | 2930 | 3030 |
| C030 | * 15000 | * 3700 H | * 3800 H | * 7900 H | * 9000 H |
| C034 | 6750 | 1600 | 1550 | 3100 | 3000 |
| MEAN | 7525 | 1773.3 | 1741.7 | 3376.7 | 3410 |
| S.D. | 757.5 | 323.3 | 228.9 | 408.1 | 364.7 |
| MEDIAN | 7650 | 1750 | 1740 | 3300 | 3450 |

APPENDIX II

Late Data Submitted by Laboratory C019

Results Report Form

CEPA National Interlaboratory Study No. CP-4

| Parameter | Sediment Extracts (pg/mL) | | | | | |
|--------------|---------------------------|-----------|-----------|-----------|-----------|-------------------------|
| | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 | Average Detection Limit |
| 2,3,7,8-TCDD | 380 | 48 | 56 | 100 | 95 | 1.6 |
| T4CDD | 620 | 120 | 110 | 240 | 230 | 1.6 |
| P5CDD | 370 | 110 | 63 | 200 | 270 | 1.2 |
| H6CDD | 880 | 310 | 300 | 590 | 540 | 3.2 |
| H7CDD | 1800 | 520 | 510 | 1040 | 1050 | 5.8 |
| O8CDD | 5300 | 1700 | 1500 | 2900 | 3200 | 6.9 |

Results Report Form

CEPA National Interlaboratory Study No. CP-4

| Parameter | Sediment Extracts (pg/mL) | | | | | |
|--------------|---------------------------|-----------|-----------|-----------|-----------|-------------------------|
| | Sample #1 | Sample #2 | Sample #3 | Sample #4 | Sample #5 | Average Detection Limit |
| 2,3,7,8-TCDF | 98 | 32 | 31 | 56 | 55 | 9.2 |
| T4CDF | 970 | 270 | 260 | 440 | 330 | 9.2 |
| P5CDF | 1400 | 390 | 330 | 630 | 790 | 4.2 |
| H6CDF | 2300 | 650 | 650 | 1300 | 1200 | 15 |
| H7CDF | 4300 | 1300 | 1200 | 2300 | 2400 | 9.8 |
| O8CDF | 9100 | 1900 | 1900 | 3700 | 4000 | 4.9 |

Results Report form

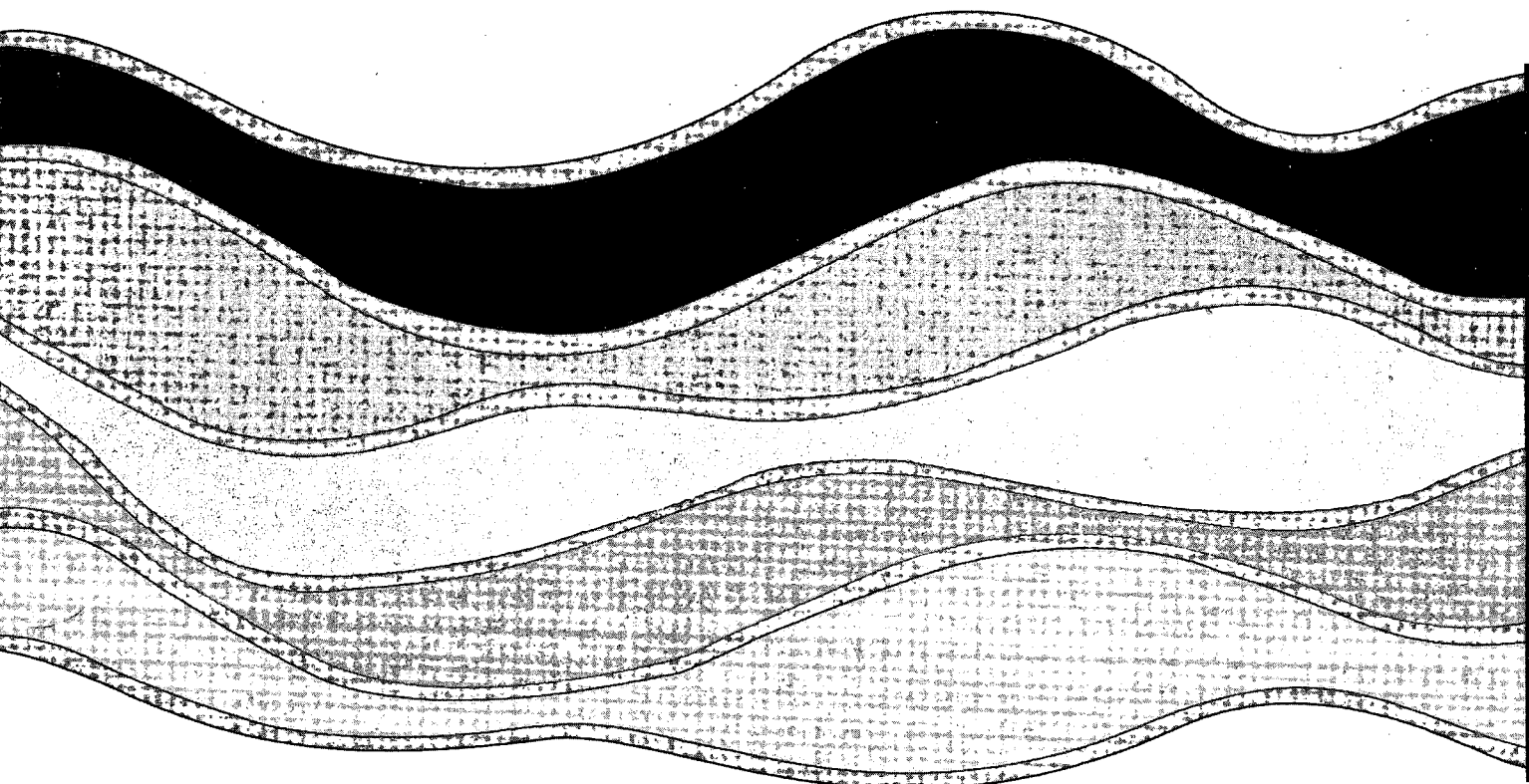
CEPA National Interlaboratory Study No. CP-4

| Sediment extract | Sample size (mL) | Final volume (μL) | Surrogate recoveries (%) | | | | | | | | | |
|------------------|------------------|-------------------|--------------------------|-------------|--------------|--------------|---------------|------------------------|-----------|-------------|---------------|--|
| | | | ¹³ C-Dioxins | | | | | ¹³ C-Furans | | | | |
| | | | 2378-TCDD | 12378-P5CDD | 123478-H6CDD | 123678-H6CDD | 1234678-H7CDD | 08CDD | 2378-TCDF | 12378-P5CDF | 1234678-H7CDF | |
| sample #1 | 2.0 | 20.0 | 76 | 91 | 103 | 85 | 62 | 45 | 79 | 94 | 84 | |
| sample #2 | 2.0 | 20.0 | 93 | 107 | 115 | 98 | 78 | 54 | 95 | 96 | 88 | |
| sample #3 | 2.0 | 20.0 | 86 | 86 | 100 | 94 | 80 | 57 | 88 | 93 | 89 | |
| sample #4 | 2.0 | 20.0 | 92 | 105 | 108 | 100 | 73 | 56 | 95 | 107 | 98 | |
| sample #5 | 2.0 | 20.0 | 85 | 65 | 119 | 98 | 77 | 58 | 82 | 92 | 93 | |

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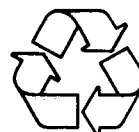
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