

FISHERIES AND ENVIRONMENT CANADA  
ENVIRONMENTAL PROTECTION SERVICE  
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PACIFIC REGION

EVALUATION OF THE A-1 MEDIUM  
FOR RAPID RECOVERY OF FECAL COLIFORMS  
FROM MARINE WATERS

78-9

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# ABSTRACT

The efficiency of the A-1 medium in the recovery of fecal coliforms from marine waters was compared with the APHA Standard Method Test. The modified A-1 method, which included a 3 hour resuscitation period at 35°C, was found to be more productive in the recovery of E.coli from the marine environment than both the standard method or the A-1 method, and equally as productive for the recovery of fecal coliforms as the standard method. The A-1 method was slightly more selective for E.coli than was the modified A-1 method, with the standard method being the least selective.

Statistical analysis using the Analysis of Variance (F) test on 273 sample results demonstrated there was no significant difference in the results obtained for each method.

## Résumé

On a comparé l'efficacité du milieu A-1 à la méthode d'essai usuelle APHA pour récupérer les bactéries coliformes d'origine fécale contenues dans les eaux marines. La méthode A-1 modifiée, suivie d'une période de réanimation de trois heures à 35°C, s'est révélée, à cet égard, aussi productive que la méthode normale et plus productive que les méthodes normale et A-1 pour récupérer les Escherichia coli.

Les méthodes normale, A-1 modifiée et A-1 se sont révélées, dans l'ordre, plus sélectives pour l'E. coli.

L'analyse statistique de variance (F), appliquée à 273 résultats échantillonnés, démontre qu'il n'existe pas de différence significative entre les résultats obtenus pour chaque méthode.

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1 INTRODUCTION

During the 1971 National Shellfish Sanitation Workshop, the Microbiology Task Force recommended that an interagency laboratory program be established to review and evaluate rapid test procedures for the bacteriological examination of shellfish growing waters. The U.S. Food and Drug Administration was requested to develop and coordinate the program.

Four procedures of potential value were reviewed and are listed as follows:

- 1) A Single Medium for the Rapid Detection of Escherichia coli at 44°C, Mara, D.D., J. Hyg. Camb. (1973), 71, 783.
- 2) Rapid Recovery of Escherichia coli from Estuarine Water, Andrews, W.H. and Presnell, M.W., Applied Microbiology, March 1972. (A-1 Procedure)
- 3) a Membrane Filtration Technique for the Enumeration of Escherichia coli in Seawater, Halls, S. and Ayres, P.A., J. of Applied Bacteriology, 37, 1974.
- 4) L.E.S. (Lawrence Experiment Station) Two-Step, Two-Day Procedure for Fecal Coliforms in Estuarine Water [See: Measurement of Fecal Coliform in Estuarine Water - Presented at the Eighth National Shellfish Sanitation Workshop, New Orleans, Louisiana, January 1974].

Two of the procedures were membrane filtration procedures and were not further considered for several reasons. Firstly, the L.E.S. method, although showing considerable promise with regard to comparable recoveries to the Standard Method, was a 48 hour, rather than 24 hour test. Secondly, some types of samples cannot be filtered because of

the presence of high concentrations of suspended colloidal matter. Finally, since the fecal coliform standard has such a low median value (14 MPN) a comparatively large sample volume may be required for the MF test.

Of the remaining two procedures, the method of Andrews and Presnell was chosen for further investigation as the A-1 medium used in this method had been shown to give E.coli recovery in 24 hours comparable to that of the standard methods procedure of the APHA (96 hour test).

The purpose of this study was to evaluate the A-1 rapid method for the enumeration of fecal coliforms in shellfish growing area waters and to compare these with the conventional standard APHA method. The methods investigated were:

- (a) a 24 hour elevated temperature (44.5°C) test with A-1 medium,
- (b) A-1 modified method with preincubation of A-1 medium for three hours at 35°C, and
- (c) the 72 to 96 hr APHA standard method procedure.

The study consisted of three steps. Firstly, a small scale sampling program was initiated in a routine growing area to obtain preliminary test data. Secondly, a series of six split samples was sent to all participating laboratories for analysis via the three methods listed above. Thirdly, a more intensive study of one year's length was conducted by participating laboratories to obtain a large enough data block for statistical analysis.

The three part study began on May 20, 1975 and was completed on June 17, 1976.

## 2 MATERIALS AND METHODS

### 2.1 Sampling Procedure

All water samples were collected in sterile 200 ml wide mouth glass bottles, approximately 15 to 30 cm below the water surface by means of a rod sampling device. Samples were stored in coolers at 10°C and were analyzed by the EPS Regional Microbiology laboratory within two hours of collection.

### 2.2 Methods of Examination

2.2.1 Standard Method. The five-tube decimal dilution (MPN) method, as described in Part 908 of the 14th edition of Standard Methods for the Examination of Water and Wastewater (1) was used. Bacto-Lauryl Tryptose Broth was used as the presumptive test medium with incubation at  $35 \pm 0.5^{\circ}\text{C}$  for 24 and 48 hours, and positive tubes were transferred to Bacto-EC medium and incubated in a water bath at  $44.5 \pm 0.2^{\circ}\text{C}$  for 24 hours.

2.2.2 A-1 Method. Three decimal dilutions of water sample were pipetted into each of five tubes of A-1 medium. The A-1 medium was prepared according to the formula of Andrews and Presnell (2). The inoculated tubes were transferred immediately into a water bath maintained at temperature of  $44.5 \pm 0.2^{\circ}\text{C}$ . Tubes showing any amount of gas after 24 hours incubation were recorded as positive.

2.2.3 Modified A-1 Method. Water samples were pipetted directly into A-1 medium in three decimal dilutions using five tubes per dilution. The inoculated tubes were first incubated in an air incubator at 35°C for three hours before being transferred directly into a water bath at  $44.5 \pm 0.2^{\circ}\text{C}$  for 21 hours.



2.2.4 Differentiation of the Fecal Coliform Types. All positive tubes from representative test media were streaked on Levine eosin methylene blue (EMB) agar plates and incubated at  $35 \pm 0.5^{\circ}\text{C}$  for 24 hours. Each colony type was picked and transferred to lactose broth and incubated at  $35 \pm 0.5^{\circ}\text{C}$  for 24 to 48 hours. All cultures from gas-positive lactose tubes were subjected to the Indole, Methyl Red, Voges-Proskauer and Simmon's Citrate Agar tests.

### 3 RESULTS AND DISCUSSION

Results from each step of the study will be presented separately, followed by a general discussion.

#### 3.1 Step 1

Forty marine samples taken from the Cates Park-Deep Cove area were analyzed using the three different methods. IMViC analyses were performed on positive tubes from the first eight samples. One colony from each EMB plate was chosen. The MPN results are presented in Table 1. The data obtained indicated that the modified A-1 method gave results which were more compatible with those obtained using the standard method EC medium. The specificity of the A-1 medium for E.coli was superior to the EC medium with recoveries of IMViC type ++-- of 83%, 100% and 100% from the EC, A-1 and A-1 modified tests. The geometric means of the three tests were 15.8, 10.4 and 12.9 respectively, with the geometric mean of the A-1 method differing significantly from the geometric mean of the standard procedure ( $p < 0.01$ ).

When the data from all participating laboratories were examined and subjected to statistical analysis, it was found that, while results of both the A-1 and modified A-1 test showed good correlation with the standard method, a statistically significant difference existed between all three methods ( $p < 0.01$ ). Both the A-1 and modified A-1 tests showed a higher recovery of E.coli than the standard test. The data are plotted on log-probability paper in Figure 1. From this graph it can be demonstrated that the A-1 modified method shows comparable results with the standard method around an MPN of 14/100 ml (shellfish growing water standard), but tends to drop below the standard method result in the higher MPN ranges (approx.  $> 100/100$  ml). This however, would not appear to be a concern in the classification of shellfish growing waters, as the upper limit for the standard is an MPN of 43/100 ml. A complete summary report of the results from Step 1 of the evaluation is found in Appendix I.

TABLE 1 SUMMARY OF SEAWATER SAMPLES ANALYSES - STEP 1  
(MPN/100 ml)

Date	Location	Standard Method			A-1		A-1 Plus	
		EC					Pre-incubation	
May 20	Cates Park	Stn # 1	5:1:0	33	4:0:0	13	4:2:0	22
		2	5:1:0	33	3:3:0	17	3:1:0	11
		3	5:1:0	33	4:3:0	27	3:0:0	8
		4	4:2:0	22	1:0:0	2	0:0:0*	<2
May 30	Deep Cove	Stn # 1	2:1:0	7	0:0:0*	<2	2:0:0	5
		2	4:1:0	17	4:1:0	17	4:1:0	17
		3	5:4:3	280	5:5:1	350	5:5:1	350
		4	1:0:0	2	2:0:0	5	2:0:0	5
May 21	Cates Park	Stn # 1	4:1:0	17	4:1:0	17	4:1:0	17
		2	5:1:0	33	4:0:0	13	3:1:0	11
		3	5:2:0	49	5:0:0	23	5:4:0	130
		4	5:0:0	23	3:0:0	8	5:1:0	33
May 21	Deep Cove	Stn # 1	4:1:0	17	1:1:0	4	4:0:0	13
		2	5:1:0	33	5:1:0	33	5:1:0	33
		3	5:5:0	240	5:2:1	70	5:2:2	94
		4	4:0:0	13	1:0:0	2	1:0:0	2
May 22	Cates Park	Stn # 1	2:0:0	5	0:0:0*	<2	3:1:0	11
		2	3:0:0	8	0:0:0*	<2	1:0:0	2
		3	5:1:0	33	5:4:0	130	5:1:0	33
		4	3:1:0	11	4:1:0	17	5:1:0	33

TABLE 1 SUMMARY OF SEAWATER SAMPLES ANALYSES - STEP 1 (cont.)  
(MPN/100 ml)

Date	Location	Standard Method			A-1		A-1 Plus	
		EC					Pre-incubation	
May 22	Deep Cove	Stn # 1	5:4:1	170	5:4:0	130	5:5:1	350
		2	2:0:0	5	2:0:0	5	1:0:0	2
		3	1:0:0	2	0:0:0*	< 2	1:0:0	2
		4	4:1:0	17	4:0:0	13	4:0:0	13
May 23	Cates Park	Stn # 1	5:1:1	46	4:2:0	22	5:2:0	49
		2	4:2:0	22	3:2:0	14	5:4:0	130
		3	4:1:0	17	5:0:0	23	5:1:0	33
		4	4:1:0	17	4:2:0	22	5:0:0	23
May 23	Deep Cove	Stn # 1	5:1:0	33	5:2:0	49	5:2:0	49
		2	2:1:0	7	3:0:0	8	1:1:0	4
		3	2:1:0	7	0:0:0*	< 2	0:0:0*	< 2
		4	4:0:0	13	2:0:0	5	3:0:0	8
May 26	Cates Park	Stn # 1	3:2:0	14	1:1:0	4	2:0:0	5
		2	4:1:0	17	4:1:0	17	4:1:0	17
		3	4:3:0	27	4:1:0	17	5:0:0	23
		4	4:0:0	13	3:1:0	11	1:0:0	2
May 26	Deep Cove	Stn # 1	2:0:0	5	3:0:0	8	2:0:0	5
		2	1:1:0	4	0:0:0*	< 2	1:0:0	2
		3	1:0:0	2	0:0:0*	< 2	4:0:0	13
		4	1:0:0	2	0:0:0*	< 2	0:1:0	2

\* These analyses, because of partial indeterminate results were not used in the evaluation of the method.

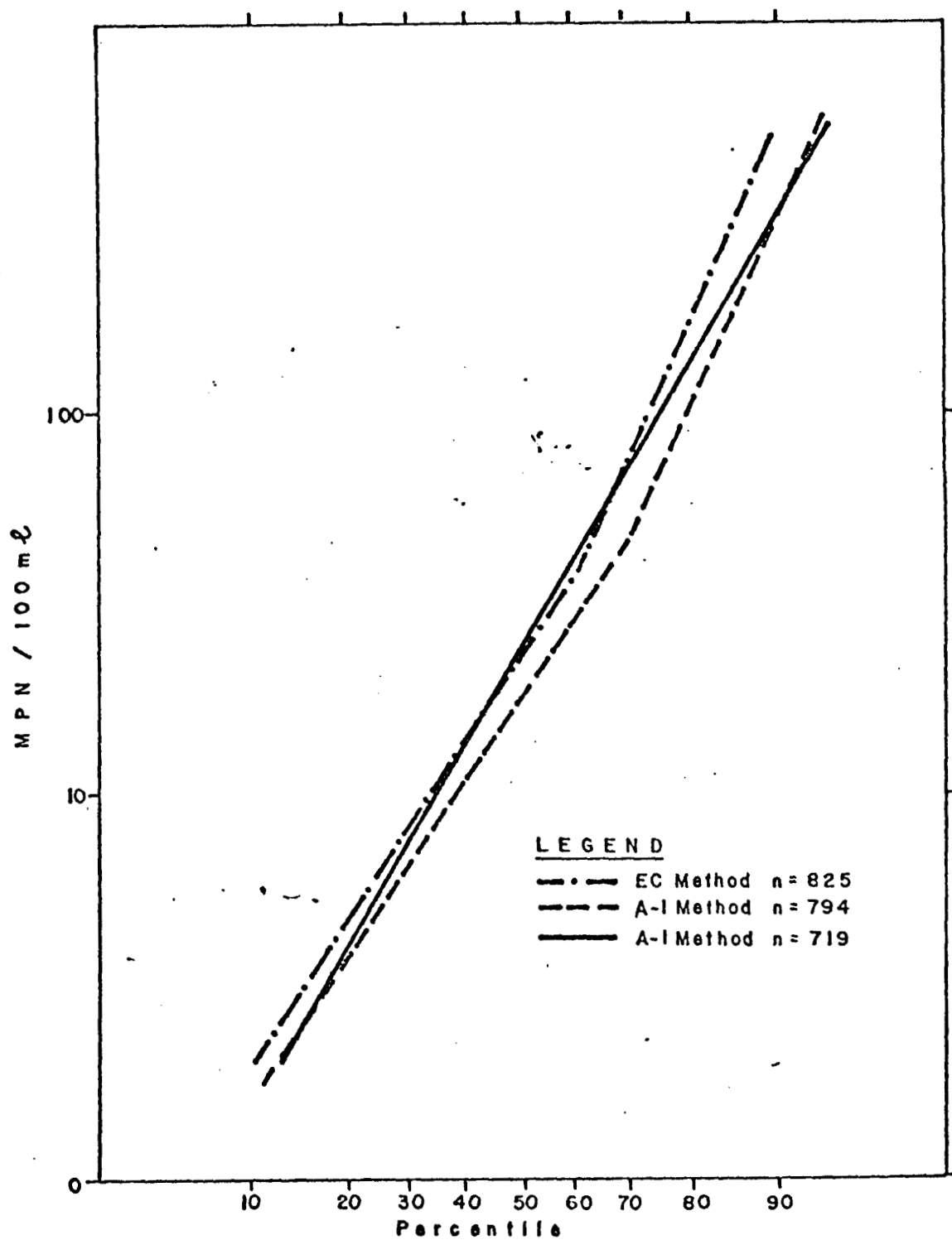


FIGURE I LOG PLOTS OF THE DATA FROM THE THREE METHODS - STEP I

3.2 Step 2

The second phase of the collaborative study was conducted to determine the comparative recovery of E. coli from a series of split artificial seawater samples using the standard fecal coliform test, the A-1 test and the modified A-1 test.

Salt was added to water to produce a salinity of 15 parts per thousand and peptone was added to a level of 20 mg/liter. The solution was divided into three equal volumes. They were subsequently spiked with a pure culture of E. coli to obtain expected recoveries of 0-10, 10-100, and 100-1000 organisms per 100 ml, respectively. Replicate subsamples of each of these solutions were sent to each of twenty-four laboratories from the Northeast Technical Services Unit (FDA) in Rhode Island. Three laboratories received their samples too late to be analyzed.

The results for the EPS-Pacific laboratory are presented in Table 2 and compared favourably with those obtained from all other

TABLE 2 BACTERIOLOGICAL DATA - SPLIT SAMPLE ANALYSIS

Sample Number	MPN per 100 ml		
	Standard Method	A-1	A-1 Modified
19	8	5	5
36	5	11	8
58	70	46	130
81	79	110	79
115	350	220	540
134	350	540	240
Geometric mean	55	57	61

participating laboratories. Table 3 summarizes geometric means for each method within each laboratory and across all laboratories.

The results from the EPS Atlantic laboratory were noticeably different from those of the other labs. This laboratory had received the samples three or four days later than did the other laboratories, and the temperature of the samples was 25°C, five degrees higher than the highest temperature recorded by the other laboratories. Table 3 indicates that there was no consistent difference between methods; that is, no single method showed a consistently higher or lower recovery than any of the other methods in all laboratories.

An analysis of variance of the data (Table 4), showed no difference between method means ( $p > 0.50$ ). When EPS Atlantic results were included, the analysis of variance showed a significant difference between laboratories ( $p < 0.01$ ). However, when these results were excluded from the analysis, no significant difference ( $p > 0.50$ ) between laboratory mean recoveries was found. This analysis of variance had a replicate subsampling error variance of 0.063. The expected variability of the 5-tube 3-dilution MPN test itself, when 10-fold dilutions are used, is 0.060, indicating that the subsampling variability was totally accounted for by the variability of the MPN test.

It was concluded from the split sample data that all three methods were equally effective in recovering pure culture E. coli from a standardized seawater sample, and that all labs were comparable in their ability to recover these bacteria with the three methods employed.

### 3.3 Step 3

At the completion of Steps 1 and 2 of the A-1 media evaluation study, all data were reviewed by FDA and the following protocol was recommended for Step 3:

TABLE 3 GEOMETRIC MEANS - SPLIT SAMPLE ANALYSIS

Laboratory	Lab Code	Date Sent	Date Arrive	Time Arrived	Temp. °C	Geometric Mean Methods Within Each Lab. N=6 for Each Method		Geometric Mean Levels Within Each Lab. N=6 for Each Level			Geometric Means All Levels, All Methods-N=18/Lab	
						Std	A-1	A-1 Mod	Level 1	Level 2		Level 3
GCTSU-FDA	1	6-17	6-20	1355	5.3	48	52	88	8	50	578	60
New York	2	6-17	6-18	1400	5.1	38	58	68	5	57	568	53
Conn.	3	6-17	6-18	1145	15.0	84	57	65	8	55	680	68
Washington	4	6-17	6-20	0950	10.0	96	55	60	8	59	638	63
Louisiana	5	6-17	6-20	1230	18.0	69	59	56	6	70	554	61
Dartmouth-EPS	6	6-16	6-25	0845	25.0	2016	1998	2236	312	1731	16702	2081
Mass.	7	6-17	6-19	0805	7.0	71	70	34	6	59	502	55
Texas	8	6-17	6-19	1100	8.0	76	63	79	6	93	675	72
Alabama	9	6-17	6-20	1100	16.5	87	45	39	6	40	590	53
NETSU-FDA	10	6-17	6-18	1900	6.4	64	59	65	7	62	586	63
South Carolina	11	6-17	6-19	1555	9.5	50	53	76	8	67	402	59
Virginia	12											
N. British Columbia-EPS	13	6-16	6-17	1700	3.5	55	57	61	7	81	352	58
British Columbia-Fish	14	6-16	6-17	1620	3.0	58	46	48	4	60	527	50
Florida	15											
Quebec-Fish	17	6-16	6-19	0930	6.0	64	65	61	9	55	534	63
Longueil-EPS	18	6-16	6-19		9.0	59	67	88	6	90	658	70
Longueil-Fish	19	6-16	6-19		4.0	67	125	75	10	55	1110	85
NETSU-FDA-PR	20											
Maryland	21	6-17	6-19	1445	7.8	59	35	41	4	56	382	44
St. John's	22											
FDA-Wash.	23	6-17	6-18	1157	4.6	74	106	62	8	83	740	79



TABLE 4 ANALYSIS OF VARIANCE - SPLIT SAMPLES

	EFFECT	SS	DF	MS	SIGNIFICANCE
EXCLUDING DARTMOUTH EPS	LABS	1.278	19	0.067	N.S.
	METHODS	.1025	2	0.051	N.S.
	LEVELS	211.21	2	105.6	**
	LAB X METHODS	2.416	36	0.067	N.S.
	LEVELS X METHODS	0.211	4	0.053	N.S.
	LABS X LEVELS	1.892	36	0.053	N.S.
	LABS X LEVELS X METHODS	4.405	72	0.061	N.S.
	REPLICATE SUBSAMPLES	10.251	171	0.060	N.S.
INCLUDING DARTMOUTH EPS	LABS	41.45	20	2.073	**
	METHODS	0.0436	2	0.022	N.S.
	LEVELS	232.5	2	116.24	**
	LABS X METHODS	2.737	40	0.068	N.S.
	LEVELS X METHODS	0.226	4	0.057	N.S.
	LABS X LEVELS	2.431	40	0.061	N.S.
	LABS X LEVELS X METHODS	4.721	80	0.059	N.S.
	REPLICATE SAMPLES	11.925	189	0.063	N.S.

N.S. - not significant

\*\* - significant difference ( $p < 0.01$ )

1. Each participating laboratory was requested to analyze at least 10 samples per month from routine sampling stations for a period of 12 months.
2. The standard E.C. test, the A-1 and A-1 modified should be done on all samples.
3. IMViC tests should be performed on all EMB colony types arising from gas-positive tubes in each of the three tests.

Sampling and analysis in this laboratory began on September 22, 1976 and continued until June 17, 1977. During this period, 100 marine samples were analyzed, representing 295 MPN and 3,619 IMViC analyses. Sampling was carried out in two areas of the lower mainland, Cates Park and Sunset Beach (False Creek). The location of stations sampled is shown in Figure 2.

All of the gas positive tubes from each method were subjected to IMViC analysis, and on the basis of these data, the presence or absence of E. coli of either IMViC type (+ + - -) was determined. The tubes from which E. coli were not isolated were considered "false positives", in that the fermentation of lactose was due to an organism or organisms other than E. coli. The number of tubes from which E. coli was isolated, was calculated as a percentage of the total number of positive tubes in each sample, and this percentage represented the effective recovery of E. coli. Omitting the false positive tubes, the MPN/100 ml was determined (IMViC MPN), and this was compared with the MPN/100 ml resulting from the inclusion of all gas positive tubes, the figure which would normally be reported using that method (METHOD MPN).

In Appendix II, the effective recovery of E. coli, and the "Method MPN" vs. the "IMViC MPN" are summarized for each month. The percent recovery of E. coli, has also been averaged on a monthly basis,

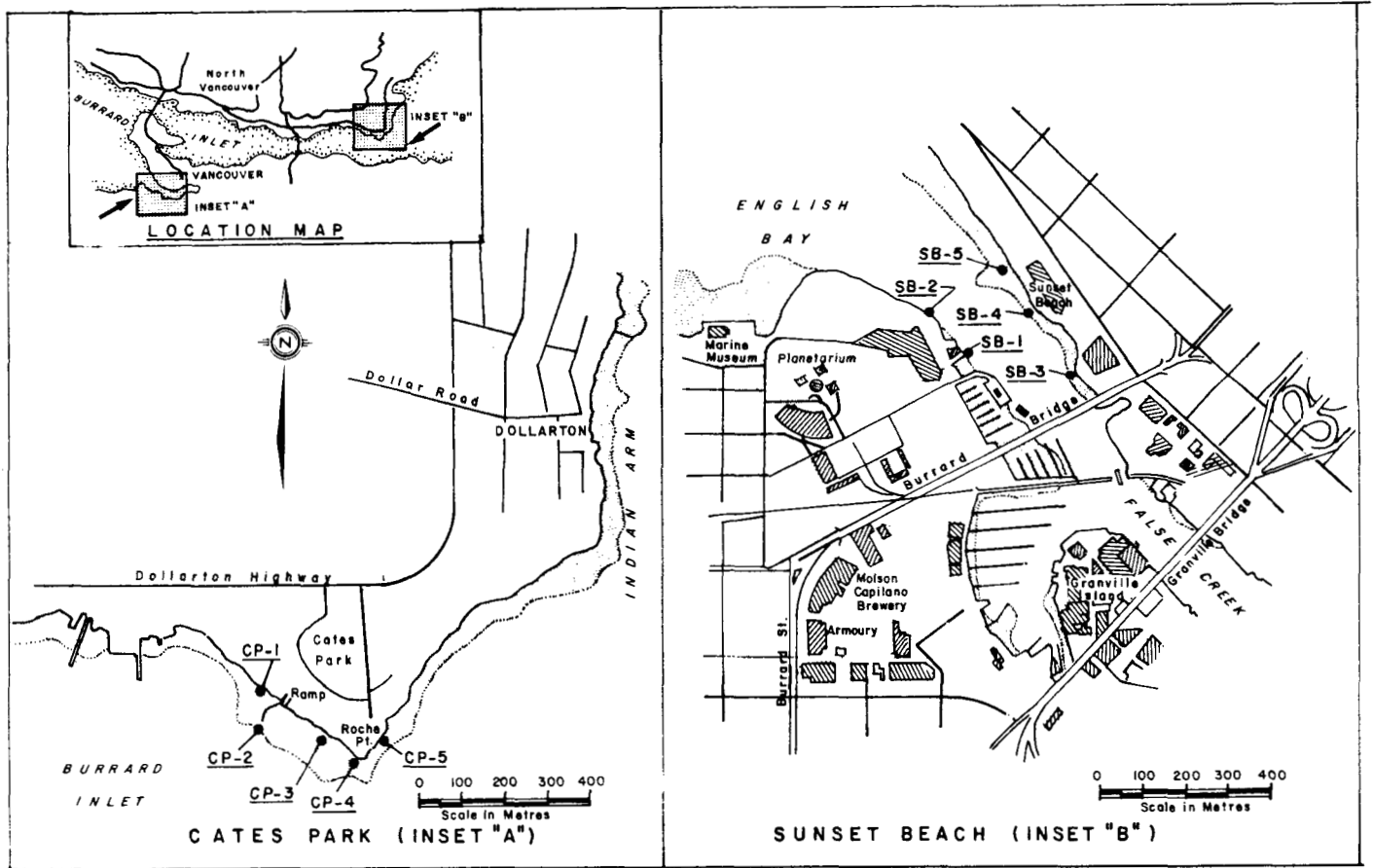


FIGURE 2 SAMPLE STATION LOCATIONS - A-I METHOD EVALUATION (STEP 3)

and this is presented on each page. The cumulative averages of recovery for each method, over 100 samples (95 in the standard method) are:

Standard Method: 90.4%  
A-1 Method: 96.8%  
Modified A-1 Method: 96.7%

The highest recovery of E. coli from gas positive tubes was achieved using the A-1 method. The modified A-1 Method gave slightly lower recovery while the standard method was the least effective in recovering E. coli.

These data, although indicating the increased selectivity of A-1 medium for E. coli, did not indicate which method gave the best recovery of E. coli from the marine environment, as opposed to recovery from gas-positive tubes (i.e., the productivity of the method).

The productivity of the methods was examined in two ways. Firstly, the productivity with respect to the recovery of E. coli was examined by totalling the number of E. coli positive tubes in each method. Secondly, the fecal coliform productivity for each method was determined by totalling the number of 44.5°C gas-positive tubes for each method. The results in Table 5 clearly demonstrate that the modified A-1 method was the most productive in recovering E. coli from the marine

TABLE 5\* PRODUCTIVITY OF THE THREE METHODS

Productivity	Number of positive tubes		
	Standard Method	A-1 Method	Modified A-1 Method
E. coli	744	757	799
Fecal coliforms	823	719	822

\*Results in this table do not include data for September 29/75 sampling.

environment and was comparable to the standard method in recovering fecal coliforms. The data suggest that the modified A-1 method has fewer false positive reactions than does the standard method.

The data were also examined statistically using the Analysis of Variance (ANOVA) test to determine whether there was any significant difference between the results obtained from each method. Both the method MPN's and the IMViC MPN's were compared between each method and across all three methods. IMViC MPN's were included in the ANOVA to compensate for any false positive reactions which occurred. The results are presented in Table 6. At the 95% level of confidence, there was no significant difference between methods or across all three methods. The

TABLE 6 ANALYSIS OF VARIANCE TEST RESULTS (STEP 3)

Methods compared	Degrees of Freedom	F value	Significance
X vs Y	181	0.505	not significant
X vs Z	181	0.5877	not significant
Y vs Z	181	0.0078	not significant
$X_1$ vs $Y_1$	181	2.6182	not significant
$X_1$ vs $Z_1$	181	2.2579	not significant
$Y_1$ vs $Z_1$	181	0.06	not significant
X vs Y vs Z	272	0.3433	not significant
$X_1$ vs $Y_1$ vs $Z_1$	272	1.7942	not significant

X = Standard Method IMViC MPN

Y = A-1 Method IMViC MPN

Z = Modified A-1 Method IMViC MPN

$X_1$  = Standard Method MPN

$Y_1$  = A-1 Method MPN

$Z_1$  = Modified A-1 Method MPN

F values for the first three comparisons indicate that, when the false positive reactions are corrected for by using the IMViC MPN's, the

methods are highly comparable. ANOVA testing was also performed on all data by FDA for each of the American regions, Canada, the seasons within regions, and the overall pooled data. The FDA interpretation of Canadian data indicated that there were significant differences between the three method means in winter but the summer data were comparable between all three methods. The winter mean MPN's for the Canadian data were 501.6/100 ml (Standard method), 356.5/100 ml (A-1 method) and 424.3/100 ml (Modified A-1 method). The higher counts provided by the standard method were perhaps due to false positive reactions which resulted in the significant difference in means. The higher incidence of false positive EC reactions during high precipitation (i.e., winter) conditions has been observed in this and other Canadian laboratories. The sanitary significance of coliform counts obtained under these conditions is therefore questionable. Bearing in mind the observed superior productivity of the A-1 medium in recovering E. coli from the marine environment, this further suggests that false positive reactions were the cause of the discrepancy between the means, as the modified A-1 test results would have approximated more closely the actual E. coli density in the sample, and the relative sanitary significance of the sample. Additional interpretation of the Canadian data was done by EPS, Atlantic Region, using the standard student's t test. At the 98% level of confidence, there was no significant difference between the standard method and the modified A-1 method for total, winter or summer data, although there was a significant difference between the standard method and the A-1 method.

4 SUMMARY

The evaluation of a new culture medium and method requires the consideration of several factors: accuracy, productivity, comparability to existing method and selectivity for the organism(s) being examined. The accuracy of A-1 medium was established by the collaborative split sample study (Step 2) which concluded that all three methods were equally proficient for the recovery of E. coli in pure culture. Productivity of the A-1 medium was examined in this laboratory and was found to be more productive in recovering E. coli from the marine environment than was the standard method, particularly when the 3 hour 35°C pre-incubation period was used. The A-1 methods were compared to the standard method using both the ANOVA and student's t tests. The results indicated no significant difference between the modified A-1 test and the standard method. The A-1 medium was found to be more selective for E. coli than was the standard methods media, the greatest selectivity being observed with the modified A-1 test method.

5 CONCLUSIONS

The modified A-1 test method was found to be superior in recovery and productivity for E. coli in sea water while producing results which were statistically comparable with the standard method.

In addition to its comparability with the standard method, the modified A-1 test method has several other practical advantages. The method requires less time, is more convenient and is less costly than the standard method. Also, by using this method, a more comprehensive shellfish water quality survey can be conducted, as it will permit the analysis of greater numbers of samples.

Based on the data presented in this and other reports (3), the modified A-1 test method is a viable alternative to the present method for the routine bacteriological examination of shellfish growing waters.



#### REFERENCES

1. "Standard Methods for the Examination of Water and Wastewater." 14th edition. American Public Health Association, Washington, D.C. (1975).
2. Andrews, W.H. and Presnell, M.W., "Rapid Recovery of Escherichia coli from Estuarine Water." Applied Microbiology, (March 1972).
3. Menon, A.S., Evaluation of A-1 Medium for the Rapid Recovery of Fecal Coliforms from Marine Waters. Environmental Protection Service, Atlantic Region. Technology Development Report [Draft], (1977).

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## APPENDICES

APPENDIX I

SUMMARY OF RESULTS OF DATA OF COMPARATIVE  
TESTS ON ROUTINE GROWING AREA SAMPLES

- Step 1

(U.S. Food and Drug Administration  
Memorandum to participating laboratories)

### Summary Of Results Of Data Comparative Test On Routine Growing Area Samples

Table 3 shows a comparison of method recoveries by the Standard, A-1, and A-1 Modified methods. The geometric means shown for each method are the means for the total number of water samples analyzed for each method. In 13 of the 20 laboratories, the method mean recoveries of fecal coliforms were significantly different from each other. The overall mean of 21.9 for the A-1 method and 26.7 for the A-1 Modified method were both significantly different from the Standard method. The ranking of the method recoveries shows that except for NETSU and three of the Canadian laboratories (Texas has too few analyses and did not use the A-1 Modified method), all laboratories recovered the highest number of fecal coliforms by the standard method, the next highest by the A-1 Modified method, and the lowest by the A-1 method.

### Conclusion

The standard method recovered significantly higher numbers of "fecal coliforms" as defined by the standard procedure than either the A-1 or A-1 Modified procedure recovered as defined by gas fermentation in 24 hours.

Results for the A-1 and A-1 Modified methods, based upon the geometric mean of the MPN values were 75% and 91% respectively of the geometric mean of the recoveries by the standard method (Table 3). Results of both tests showed good correlation with the standard method although a statistically significant difference exists between all three methods. Both the A-1 and A-1 Modified tests showed a higher recovery of E. coli, the principal coliform organism, than the standard test (Table 6). The interaction of the analytical methods with the geographic and physical variables of the waters such as temperature, salinity, and turbidity, was not delineated by the data presented. Both the A-1 and A-1 Modified tests indicated differences in the sanitary quality of the waters tested which corresponded to the differences indicated by the standard methods (Graphs 1 & 2).

TABLE 3 COMPARISON OF METHODS USING DATA FROM ROUTINE GROWING AREA SAMPLES

Geometric Mean Of Method Recoveries Of Fecal Coliform							Ranking Of Method Recoveries <sup>2</sup>				
Laboratory	Lab Code	Stand.	n <sup>1</sup>	AI	n	AI-Mod	n	Sign	Stand.	AI	AI-Mod
Gulf Coast	1	48.5	42	29.3	42	34.9	42	**	1	3	2
New York	2	3.7	40	2.4	41	3.6	39	**	1	3	2
Conn.	3	181.4	32	88.5	35	125.6	34	**	1	3	2
Washington	4	67.3	50	36.8	50	45.7	50	**	1	3	2
Louisiana	5	27.7	43	23.6	43	26.2	43	N.S.	1	3	2
Dartmouth	6	37.0	42	28.3	42	36.6	42	*	2	3	2
Massachusetts	7	18.1	49	11.8	49	16.8	49	**	1	3	2
Texas	8	194.1	12	213.3	12			N.S.	3	1	2
Alabama	9	35.2	41	27.6	41	35.2	41	N.S.	1.5	3	1.5
NETSU	10	21.9	39	14.3	39	23.3	39	**	2	3	1
S. Carolina	11	63.7	40	36.6	40	36.1	40	**	1	2	3
Virginia	12	38.8	44	31.0	41	35.5	44	N.S.	1	3	2
N. Vancouver, B.C.	13	15.8	40	10.4	40	12.9	40	**	1	3	2
British Columbia	14	13.6	101	10.5	101	13.1	36	**	1	3	2
Florida	15	96.1	30	58.9	30	82.4	30	**	1	3	2
Provincial-Quebec	17	33.1	35	50.4	28	45.7	30	N.S.	3	1	2
Longueiel-EPS	18	52.6	50	52.8	25	49.8	26	N.S.	2	1	3
Fisheries-Longueiel	19	51.4	50	51.9	51	32.5	50	**	2	1	3
Parker River, Mass.	20	16.5	29	24.4	29	29.6	29	**	3	2	1
Maryland	21	10.2	58	8.7	57	10.1	57	N.S.	1	3	2
Over All States		29.3	867	21.9*	836	26.7*	761				

<sup>1</sup>no. of samples analyzed for each method<sup>2</sup><sub>1</sub> = method recovering highest no. of fecal coliforms<sub>2</sub> = method recovering second highest no. of fecal coliforms<sub>3</sub> = method recovering lowest no. of fecal coliform

\* - indicates methods are significantly different at p 0.01

N.S. - indicates no significant difference between methods

TABLE 4                      PERCENTILES - FECAL COLIFORM - 21 STATES

	Standard	A1	A1-Mod
10	2.0	2.0	2.0
20	4.5	2.0	4.0
30	7.8	6.8	7.8
40	17.0	11.0	14.0
50	23.0	17.0	23.0
60	33.0	33.0	46.0
70	79.0	49.0	79.0
80	170.0	110.0	130.0
90	540.0	350.0	350.0
100	4900.0	13,000.0	4900.0
n	825	794	719

TABLE 5                      COMPARISON OF GEOMETRIC MEAN OF EACH TEST AGAINST  
THE STANDARD PROCEDURE

Lab. No.	Standard	A-1	A-1 Mod.
1	48.5	29.3**	34.9**
2	3.7	2.4**	3.6 N.S.
3	181.4	88.5**	125.6 *
4	67.3	36.8**	45.7 *
5	27.7	23.6 N.S.	26.2 N.S.
6	37.0	28.3 *	36.6 N.S.
7	18.1	11.8**	16.8 N.S.
8	194.1	213.3 N.S.	
9	35.2	27.6 N.S.	35.2 N.S.
10	21.9	14.3**	23.3 N.S.
11	63.7	36.6**	36.1**
12	38.8	31.0 N.S.	35.5 N.S.
13	15.8	10.4**	12.9 N.S.
14	13.6	10.5 *	13.1 N.S.
15	96.1	58.9**	82.4 N.S.
17	33.1	50.4 N.S.	45.7 N.S.
18	52.6	52.8 N.S.	49.8 N.S.
19	51.4	51.9 N.S.	32.5 *
20	16.5	24.4 *	29.6**
21	10.2	8.7 *	10.1 N.S.

N.S. - no significant difference  $p < 0.05$

\* - significantly different from standard ( $p < 0.05$ )

\*\* - significantly different from standard ( $p < 0.01$ )



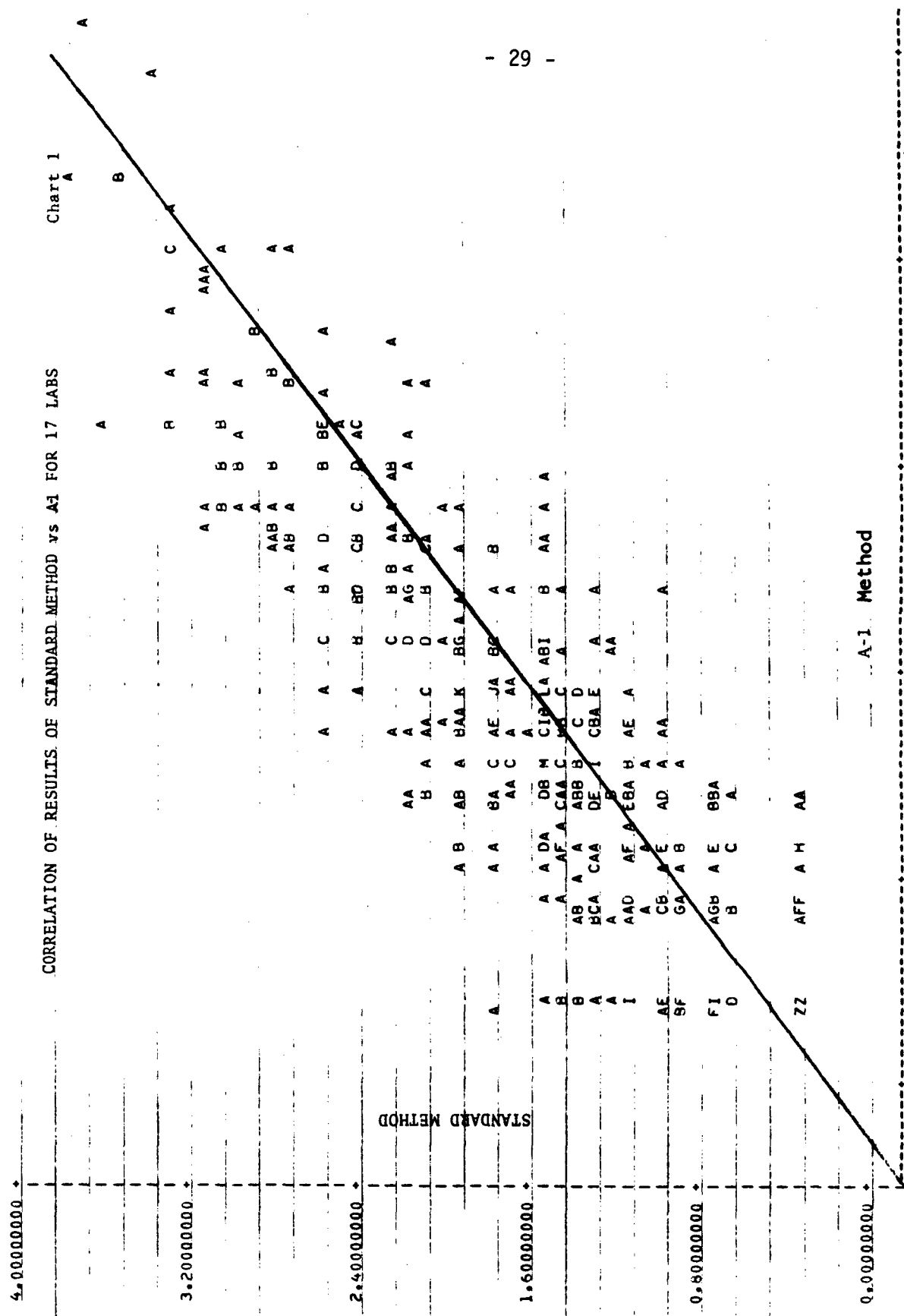
TABLE 6 SUMMARY OF IMViC DATA

Lab Code #	TUBES			TUBES			TUBES		
	Standard			A-1			A1-Modified		
	# positive	total	%	# pos.	total	%	# pos.	total	%
21	141	165	85.5	146	158	92.4	146	163	89.6
9	25	39	64.1	28	32	87.5	28	32	87.5
19	11	11	100	12	12	100	12	12	100
17	n.r.	n.r.	82	n.r.	n.r.	92	n.r.	n.r.	91
14	n.r.	n.r.	83	n.r.	n.r.	75	n.r.	n.r.	100
18	55	70	78.6	42	56	82.1	26	26	100
20	39	50	78.0	32	39	80	42	43	97.7
6	34	40	85	68	85	97.5	69	71	97.2
3				115	118	92	118	118	100
1	207	260	79	198	215	98.7	210	233	90
5	247	259	95	236	239		246	251	98

#positive - No. tubes positive for E. coli IMViC ++-- or -+--

total - total number of gas positive tubes IMViC tested

n.r. - not reported

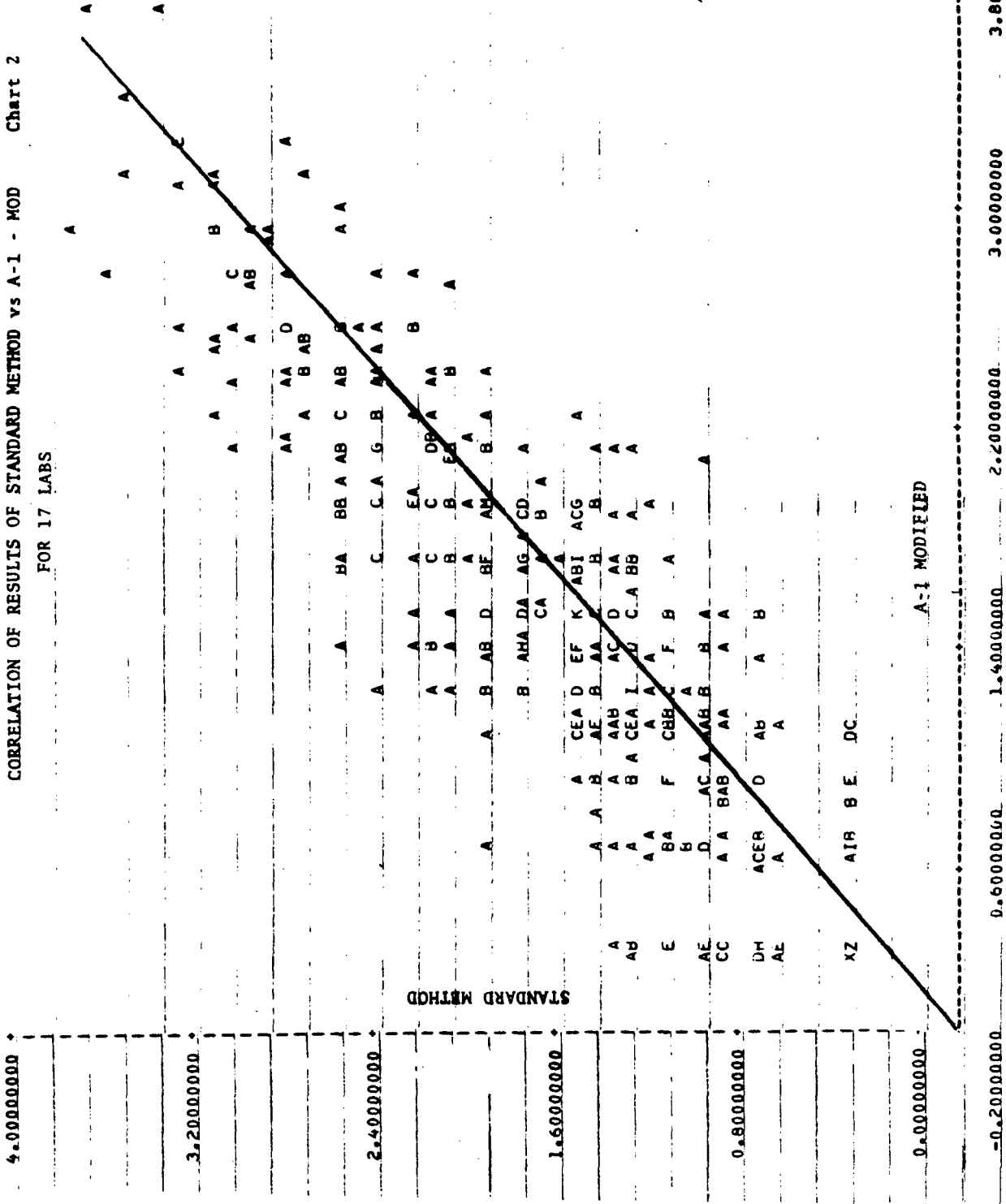


0.00000000 0.80000000 1.60000000 2.40000000 3.20000000 4.00000000

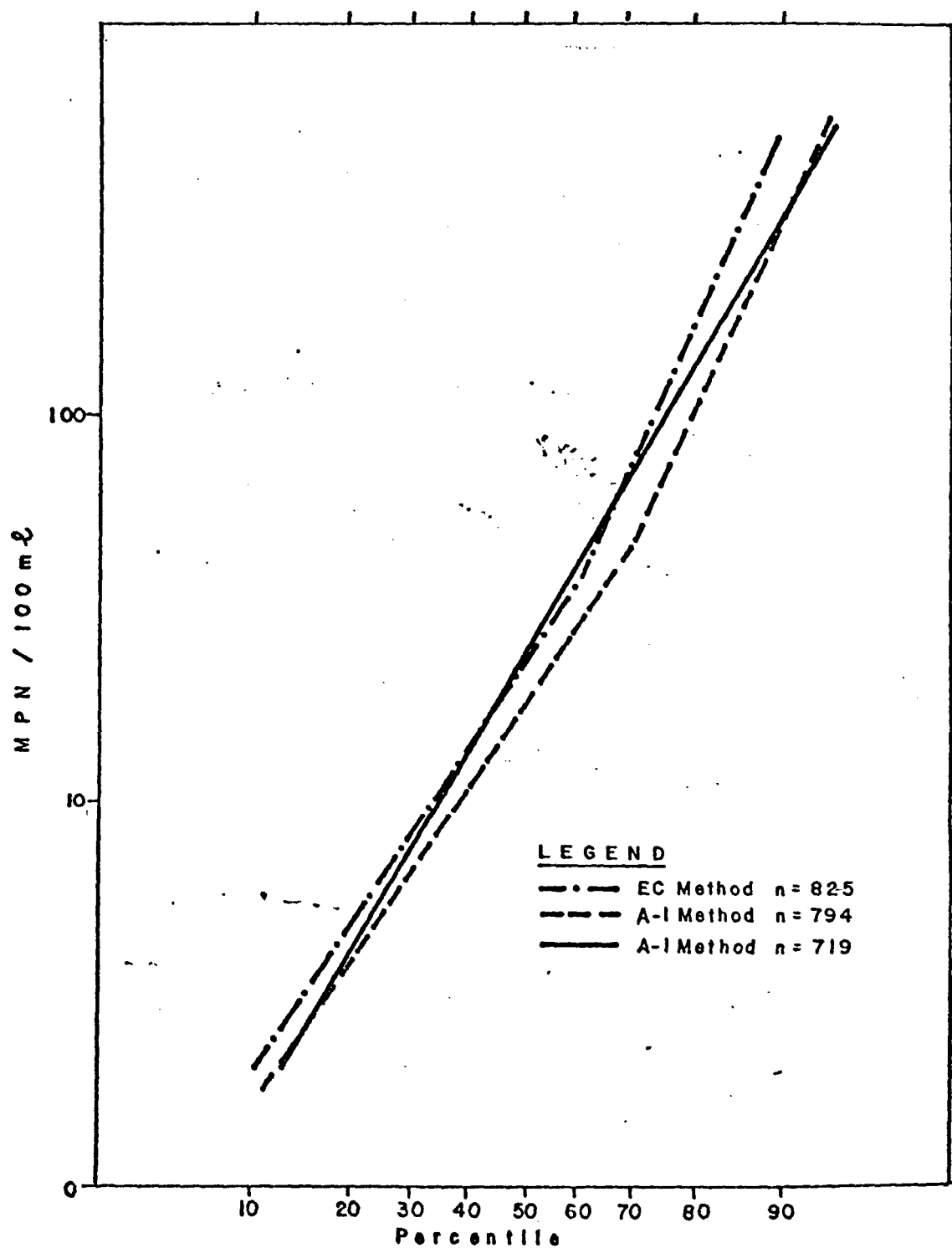
LEGEND: A = 1 OBS, B = 2 OBS, ETC.

CORRELATION OF RESULTS OF STANDARD METHOD vs A-1 - MOD Chart 2

FOR 17 LABS



LEGEND: A = 1 OBS , B = 2 OBS , ETC.



LOG PLOTS OF THE DATA FROM THE  
THREE METHODS - STEP I

APPENDIX II

A-1 MEDIA EVALUATION - DATA SURVEY

- Step 3

APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY

Date	Sample	Standard Method					A-1 Method					Modified A-1 Method					
		#E. coli/ Total #		%	Method	IMViC MPN/ 100 ml	#E. coli/ Total #		%	Method	IMViC MPN/ 100 ml	#E. coli/ Total #		%	Method	IMViC MPN/ 100 ml	
		Tubes	Recovery	100 ml	100 ml	Tubes	Recovery	100 ml	100 ml	Tubes	Recovery	100 ml	100 ml	Tubes	Recovery	100 ml	100 ml
Sept.	CP1	5/5	100	23	23	1/1	100	2	2	3/3	100	8	8	3/3	100	8	
22/75	CP2	2/2	100	5	5	3/3	100	8	8	2/2	100	5	5	2/2	100	5	
	CP3	1/1	100	2	2	2/2	100	5	5	1/1	100	2	2	1/1	100	2	
	CP4	3/3	100	8	8	1/1	100	2	2	2/2	100	7	7	2/2	100	7	
	CP5	2/2	100	5	5	4/4	100	13	13	2/3	66.6	4	4	2/3	66.6	4	
Sept.	SB1	-	-	79	-	7/8	87	70	46	9/9	100	140	94*	9/9	100	140	
29/75	SB2	-	-	170	-	5/6	83	33	17	8/8	100	79	79	8/8	100	79	
	SB3	-	-	350	-	9/10	90	240	41	8/10	80	240	25*	8/10	80	240	
	SB4	-	-	920	-	9/9	100	130	130	9/12	75	540	28	9/12	75	540	
	SB5	-	-	23	-	8/9	89	110	33	6/6	100	49	22*	6/6	100	49	
Average % Recovery		100		94.9		92.2											

\* One or more tubes lack IMViC data.

APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method				A-1 Method				Modified A-1 Method			
		#E. coli/		Method	IMViC	#E. coli/		Method	IMViC	#E. coli/		Method	IMViC
		Total #	%	MPN/ 100 ml	MPN/ 100 ml	Total #	%	MPN/ 100 ml	MPN/ 100 ml	Total #	%	MPN/ 100 ml	MPN/ 100 ml
		Tubes	Recovery	100 ml	100 ml	Tubes	Recovery	100 ml	100 ml	Tubes	Recovery	100 ml	100 ml
Oct. 6/75	SB1	8/9	39	110	33	11/11	100	350	350	10/10	100	240	240
	SB2	10/10	100	170	170	9/9	100	130	130	10/10	100	240	240
	SB3	10/10	100	240	240	9/9	100	110	110	11/11	100	350	350
	SB4	6/7	86	49	33	7/7	100	49	49	8/8	100	79	79
	SB5	8/9	89	130	79	8/8	100	79	79	10/10	100	240	240
Oct. 13/75	SB1	14/14	100	1600	1600	13/13	100	920	920	12/13	92	920	64
	SB2	10/12	83	540	48	14/14	100	1600	1600	11/12	92	920	180*
	SB3	14/14	100	1600	1600	11/12	92	540	350	14/15	93	1600	1600
	SB4	13/14	93	1600	350	13/13	100	920	920	12/13	92	1600	280*
	SB5	11/11	100	350	350	12/12	100	540	540	10/12	83	540	47
Average % Recovery		94.0				99.2				95.2			

\* One or more tubes lack IMViC data

APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method						A-1 Method						Modified A-1 Method					
		#E. coli/			Method			#E. coli/			Method			#E. coli/			Method		
		Total #	%	Recovery	MPN/	100 ml	IMViC	Total #	%	Recovery	MPN/	100 ml	IMViC	Total #	%	Recovery	MPN/	100 ml	IMViC
		Tubes			MPN/	100 ml	100 ml	Tubes			MPN/	100 ml	100 ml	Tubes			MPN/	100 ml	100 ml
Oct. 27/75	SB1	10/11	91		3500	1700	14/14	100		100	1600	1600	1600	11/12	92		540	540	56
	SB2	10/11	91		3500	1700	9/9	100		100	920	920	920	11/11	100		350	350	350
	SB3	7/8	87		790	270	10/11	91		91	350	350	240	13/13	100		920	920	920
	SB4	8/8	100		790	790	11/11	100		100	240	240	240	12/12	100		540	540	540
	SB5	6/7	86		460	210	11/11	100		100	350	350	350	10/10	100		240	240	240
Nov. 3/75	SB1	9/13	69		920	28	10/11	91		91	220	220	47	10/10	100		240	240	240
	SB2	7/11	67		350	21	11/12	92		92	540	540	56	9/10	90		240	240	41
	SB3	12/13	92		920	64	14/14	100		100	1600	1600	1600	13/13	100		920	920	920
	SB4	11/12	92		540	350	9/11	82		82	540	540	40*	10/11	91		540	540	48
	SB5	9/11	82		220	39	10/11	91		91	220	220	56	12/13	92		920	920	540
Average % Recovery		85.7									94.7						96.5		

\*One or more tubes lack IMViC data



APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method					A-1 Method					Modified A-1 Method				
		#E. coli/		Method		IMViC	#E. coli/		Method		IMViC	#E. coli/		Method		IMViC
		Total #	%	MPN/	100 ml		Total #	%	MPN/	100 ml		Total #	%	MPN/	100 ml	
		Tubes	Recovery	100 ml	100 ml		Tubes	Recovery	100 ml	100 ml		Tubes	Recovery	100 ml	100 ml	
Nov. 25/75	SB1	8/10	80	2400	340		8/8	100	790	790		9/9	100	1300	1300	
	SB2	7/7	100	490	490		10/10	100	1700	1700		8/8	100	790	790	
	SB3	7/7	100	490	490		7/8	87	790	490		9/9	100	1300	1300	
	SB4	9/9	100	1700	1700		6/6	100	330	330		8/8	100	700	700	
	SB5	6/6	100	330	330		8/8	100	790	790		10/10	100	1700	1700	
Dec. 2/75	SB1	8/10	80	2400	340		9/9	100	1300	1300		9/10	90	1700	1100	
	SB2	10/10	100	2400	2400		10/10	100	1400	1400		10/10	100	2400	2400	
	SB3	10/10	100	2400	2400		9/9	100	1300	1300		10/11	91	3500	480	
	SB4	8/10	80	2400	250		8/8	100	790	790		9/9	100	1300	1300	
	SB5	8/9	89	1300	790		12/12	100	5400	5400		10/10	100	1400	1400	
Average % Recovery			92.9					98.7					98.1			

APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method					A-1 Method					Modified A-1 Method				
		#E. coli/ Total #	Tubes	Recovery %	Method MPN/ 100 ml	IMViC MPN/ 100 ml	#E. coli/ Total #	Tubes	Recovery %	Method MPN/ 100 ml	IMViC MPN/ 100 ml	#E. coli/ Total #	Tubes	Recovery %	Method MPN/ 100 ml	IMViC MPN/ 100 ml
Jan. 12/76	SB1	13/14	93		1600	920	13/14	93		1600	350	11/11	100		350	350
	SB2	13/15	87		2400	340	14/14	100		1600	1600	13/13	100		920	920
	SB3	13/14	93		1600	350	16/16	100		2200	2200	14/14	100		1600	1600
	SB4	14/15	93		1700	400	12/12	100		540	540	14/14	100		1100	1100
	SB5	15/16	94		2200	470	14/14	100		1600	1600	15/15	100		1400	1400
Jan. 19/76	SB1	11/11	100		350	350	10/10	100		170	170	10/11	91		350	48
	SB2	10/10	100		240	240	10/11	91		350	170	12/12	100		280	280
	SB3	12/12	100		540	540	12/12	100		350	350	13/14	93		1600	920
	SB4	14/14	100		1600	1600	12/12	100		540	540	13/13	100		920	920
	SB5	11/11	100		350	350	10/10	100		240	240	12/12	100		540	540
Average % Recovery		96.0					98.4					98.4				

## APPENDIX II

## A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method				A-1 Method				Modified A-1 Method			
		#E. coli/ Total #		Method		#E. coli/ Total #		Method		#E. coli/ Total #		Method	
		Tubes	% Recovery	MPN/ 100 ml	IMViC 100 ml	Tubes	% Recovery	MPN/ 100 ml	IMViC 100 ml	Tubes	% Recovery	MPN/ 100 ml	IMViC 100 ml
Feb.	CP1	10/10	100	170	170	7/7	100	49	49	9/9	100	110	110
2/76	CP2	7/7	100	49	49	7/7	100	49	49	9/9	100	130	130
	CP3	8/8	100	79	79	6/6	100	33	33	7/7	100	46	46
	CP4	7/10	70	240	27	11/11	100	350	350	10/10	100	140	140
	CP5	6/8	75	70	17	6/7	86	49	33	7/9	78	130	49
Feb.	CP1	9/9	100	110	110	8/8	100	33	33	8/8	100	70	70
16/76	CP2	10/10	100	240	240	8/8	100	95	95	8/8	100	79	79
	CP3	9/10	90	170	110	8/8	100	79	79	8/8	100	79	79
	CP4	10/10	100	140	140	7/7	100	27	27	7/7	100	49	49
	CP5	8/9	89	130	34	7/7	100	49	49	7/7	100	49	49
Average % Recovery		92.4				98.6				97.8			

## APPENDIX II

Date	Sample	Standard Method				A-1 Method				Modified A-1 Method			
		#E. coli/		Method	IMViC	#E. coli/		Method	IMViC	#E. coli/		Method	IMViC
		Total #	%	MPN/	100 ml	Total #	%	MPN/	100 ml	Total #	%	MPN/	100 ml
		Tubes	Recovery	100 ml	100 ml	Tubes	Recovery	100 ml	100 ml	Tubes	Recovery	100 ml	100 ml
March	CP1	7/7	100	49	49	4/4	100	11	11	8/9	88.8	130	79
2/76	CP2	5/6	83.3	33	17	5/6	83.3	22	14	5/5	100	23	23
	CP3	6/6	100	33	33	5/5	100	14	14	5/5	100	23	23
	CP4	7/7	100	46	46	7/7	100	49	49	6/6	100	33	33
	CP5	5/5	100	23	23	7/7	100	49	49	5/5	100	23	23
March	CP1	8/8	100	79	79	6/6	100	33	33	5/6	83.3	22	14
8/76	CP2	6/6	100	33	33	6/6	100	33	33	7/8	87.5	79	27
	CP3	6/6	100	33	33	4/4	100	11	11	5/5	100	17	17
	CP4	5/5	100	17	17	4/4	100	13	13	4/4	100	13	13
	CP5	4/4	100	11	11	5/5	100	17	17	3/3	100	8	8
Average % Recovery			98.3				98.3				95.9		

APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method						A-1 Method						Modified A-1 Method					
		#E. coli/			Method			#E. coli/			Method			#E. coli/			Method		
		Total #	%	Recovery	MPN/	100 ml	100 ml	Total #	%	Recovery	MPN/	100 ml	100 ml	Total #	%	Recovery	MPN/	100 ml	100 ml
		Tubes						Tubes						Tubes					
April	CP1	6/6	100		31		31	5/5	100		23		23	6/6	100		33		23
20/76	CP2	7/8	87.5		79		27	6/6	100		22		22	5/5	100		17		17
	CP3	5/5	100		23		23	3/3	100		7		7	7/7	100		49		49
	CP4	7/7	100		49		49	5/5	100		23		23	6/6	100		33		33
	CP5	7/7	100		46		46	7/7	100		46		46	9/9	100		130		130
April	CP1	6/9	66.6		130		17	6/6	100		17		17	4/4	100		9		9
27/76	CP2	6/6	100		17		17	2/2	100		5		5	3/4	75		11		8
	CP3	6/6	100		33		33	2/2	100		5		5	3/4	75		13		8
	CP4	8/8	100		33		33	10/10	100		170		170	10/10	100		170		170
	CP5	1/2	50		4		2	-	-		2		-	-	-		2		-
Average % Recovery			90.4						100								94.4		

APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method						A-1 Method						Modified A-1 Method					
		#E. coli/			Method			#E. coli/			Method			#E. coli/			Method		
		Total #	%	Recovery	MPN/	100 ml	100 ml	Total #	%	Recovery	MPN/	100 ml	100 ml	Total #	%	Recovery	MPN/	100 ml	100 ml
		Tubes			MPN/	100 ml	100 ml	Tubes			MPN/	100 ml	100 ml	Tubes			MPN/	100 ml	100 ml
May	SB1	5/8	62.5		70	12	10/10	10/10	100		240	240	240	10/10	100		240	240	240
3/76	SB2	4/6	66.6		330	110	8/9	88.8			130	34	34	10/10	100		240	240	240
	SB3	3/7	42.9		490	80	10/10	100			240	240	240	11/11	100		350	350	350
	SB4	2/8	25		700	40	10/10	100			170	170	170	10/10	100		170	170	170
	SB5	3/6	50		220	70	11/11	100			350	350	350	12/12	100		540	540	540
May	SB1	7/9	77.7		130	27	8/8	100			79	79	79	5/6	83.3		31	17	17
10/76	SB2	7/7	100		46	46	6/6	100			33	33	33	7/7	100		49	33	33
	SB3	5/6	83.3		330	170	5/5	100			230	230	230	6/6	100		330	330	330
	SB4	4/10	40		140	8	8/8	100			79	79	79	8/8	100		79	79	79
	SB5	6/7	85.7		49	22	4/4	100			13	13	13	3/3	100		7	7	7
Average % Recovery		63.4				98.8				98.3				98.3					

APPENDIX II A-1 MEDIA EVALUATION - DATA SURVEY (continued)

Date	Sample	Standard Method					A-1 Method					Modified A-1 Method				
		#E. coli/		Method		IMViC	#E. coli/		Method		IMViC	#E. coli/		Method		IMViC
		Total #	%	MPN/	100 ml		Total #	%	MPN/	100 ml		Total #	%	MPN/	100 ml	
		Tubes	Recovery	100 ml	100 ml		Tubes	Recovery	100 ml	100 ml		Tubes	Recovery	100 ml	100 ml	
May	SB1	5/6	83.3	33	17	17	5/6	83.3	33	17	17	5/5	100	23	23	
31/76	SB2	5/6	83.3	33	23	23	6/6	100	33	33	33	7/7	100	49	49	
	SB3	7/7	100	49	49	49	6/6	100	33	33	33	8/8	100	79	79	
	SB4	9/10	90	240	48	48	4/4	100	13	13	13	5/5	100	23	23	
	SB5	8/8	100	70	70	70	2/2	100	5	5	5	4/4	100	13	13	
June	SB1	8/10	80	2400	790	790	12/12	100	540	540	540	10/10	100	240	240	
7/76	SB2	11/11	100	350	350	350	9/9	100	110	110	110	11/11	100	350	350	
	SB3	9/11	81.8	220	39	39	11/12	91.6	540	350	350	13/13	100	920	920	
	SB4	6/8	75	79	33	33	7/8	87.5	70	26	26	10/10	100	240	240	
	SB5	4/9	44.4	130	9	9	9/10	90	240	41	41	10/10	100	240	240	
Average % Recovery			83.5					95.2								100