

Data Record on Sediment Attributes and Infaunal Diversity on Beam-Trawl and Trap-Line Transects in Clio Channel, British Columbia, Canada

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COLUMBIA, CANADA

By

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ABSTRACT

Trofte, P.M., Levings, C.D., Sutherland, T.F., Chang, H., Piercey, G.E., Petersen, S.A., Keong, V., Poon, P., McDermid, M., and Sunday, J. 2024. Data record on sediment attributes and Infaunal diversity on Beam-Trawl and Trap-Line transects in Clio Channel, British Columbia, Canada. *Can. Data Rep. Fish. Aquat. Sci.* 1388: viii + 32 p.

As part of a larger assessment project on the central coast of British Columbia investigating potential modification of marine ecosystems by shrimp trawling and trapping, we collected infauna and sediment samples before and after fishing with beam trawls and traps. Both Turnour Bay and Bones Bay, located in Clio Channel, were chosen to collect sediment and infaunal samples and carry out beam-trawling and trapping activities in January and March, 2002. Tabulated data are presented on both meiofauna and macrofauna abundance as well as sediment attributes including sediment grain size, porosity, carbon and nitrogen content, trace metal and chlorophyll concentrations.

RÉSUMÉ

Trofte, P.M., Levings, C.D., Sutherland, T.F., Chang, H., Piercey, G.E., Petersen, S.A., Keong, V., Poon, P., McDermid, M., and Sunday, J. 2024. Data record on sediment attributes and Infaunal diversity on Beam-Trawl and Trap-Line transects in Clio Channel, British Columbia, Canada. Can. Data Rep. Fish. Aquat. Sci. 1388: viii + 32 p.

Dans le cadre d'un projet d'étude des modifications des écosystèmes marins de la côte centrale de la Colombie-Britannique qui peuvent résulter de la pêche commerciale des crevettes au chalut et au casier, nous avons prélevé des échantillons d'endofaune et de sédiments avant et après les opérations de pêche. ainsi que deux baies du chenal Clio en janvier et mars 2002. Sont présentées dans ce rapport les données tabulées sur l'abondance de la macrofaune et de la méiofaune, ainsi que sur les caractéristiques des sédiments, y compris la taille des particules, la porosité et les teneurs en carbone, en azote, en métaux-traces et en chlorophylle.

1.0 INTRODUCTION

Sediment attributes and infaunal abundance (meiofauna, macrofauna) were characterized in Clio Channel, located off Johnstone Strait in the south central coast of British Columbia, Canada. Seafloor sampling was conducted before and after the deployment of two types of commercial shrimp fishing gear in two bays on Clio Channel: 1) beam trawl and trap-lines (Turnour Bay); and 2) traps (Bones Bay). Although the program's over-arching objective was to compare potential benthic impacts of two fishing methods on sediment characteristics and infaunal communities, this report is focused solely on data presentation before and after fishing activities. A parallel study took place in Simoom Sound, British Columbia, Canada and the data record for Simoom Sound can be found in Sutherland et al. 2024. Analysis of fishing gear impacts on sea whips (*Balticina willemoesi*) (Kölliker, 1870) (formerly *Halipterus willemoesi*) and sediments at Turnour Bay and Bones Bay was published in Troffe et al (2005). In this Data Report we present raw data that were given in summary form in Troffe et al (2005).

2.0 STUDY AREA

Clio Channel is a narrow marine waterway bordered by both Turnour Island and West Cracroft Island and oriented in a southwest-northeast bearing off Johnstone Strait. Turnour Bay has a length of 1.5 km, a width of 1.0 km, and is located on the southwest side of Turnour Island. The seafloor slopes from the northeast end of the bay (25 m) to the north-west entrance (40m). Sediment texture mainly consisted of sand mixed with small boulders, and sporadic rocky outcrops. Bones Bay is located on the northeast end of Clio Channel on the northwest side of West Cracroft Island. The bay has a length of 1.5 km, a width of 1.5 km, and a depth range of 64 m (northeast) and 74 m (southwest). Although the seafloor sloped across the bay, the sediment texture was consistent with muddy sediments. In the past, a salmon cannery, logging camp, and log storage were located in Bones Bay.

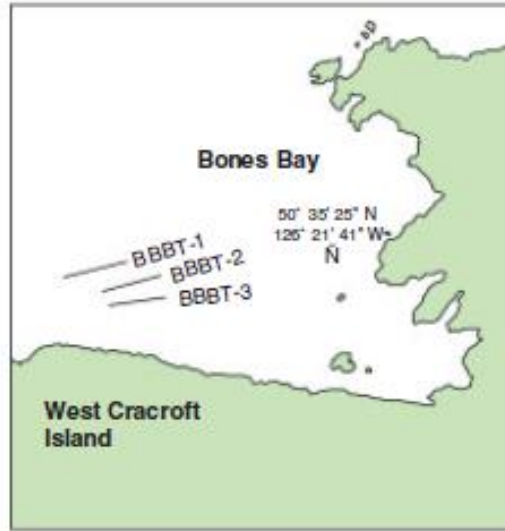
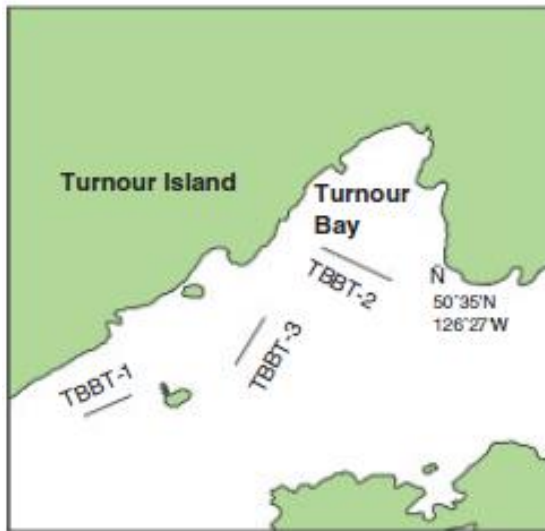
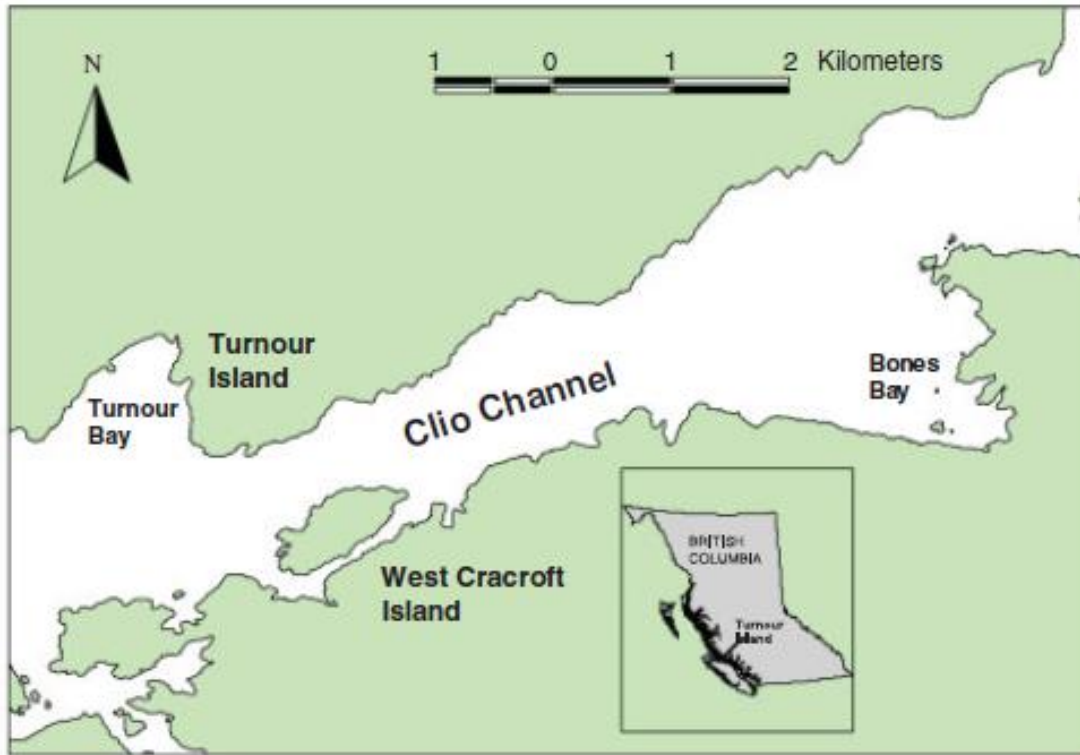


FIGURE 1: Map of the study areas and fishing lines in Clio Channel located on the central coast of British Columbia, Canada (from Troffe et al. 2005). TBBT = Turnour Bay Beam Trawl; BBBT = Bones Bay Beam Trawl; 1, 2, 3 = replicate trawl transects.

3.0 MATERIALS AND METHODS

The type of fishing gear and deployment methods used in this study were representative of fishing activities used at that time in the commercial shrimp fishery in British Columbia.

Beam trawling: A beam-trawl was towed once on each of three replicate transects (Figure 1) in Turnour Bay (January, 2002) and Bones Bay (October, 2001). The aluminium beam pole was 14 m long, while the net (codend size: 44 mm) was hung from rounded skids with a height of approximately 1.5 m. The footrope was attached about 30 cm from the bottom of the skids. The 11 m vessel (Amethyst II) towing the net was equipped with a 138 kw engine. Position information was obtained at 1-min intervals using a differential global positioning system in the wheelhouse of the vessel. The duration and length of the trawling transects ranged between 8 and 19 min and 311 to 419 m, respectively (Table 1).

Trapping: In March 2002, 40 baited traps were set and hauled five times by overlaying a trapline on each of the beam-trawling transects in Turnour Bay. The 12.6 m prawn vessel Eric Jane was deployed for this work. These repetitive trap deployments resulted in a total of 600 traps set and retrieved. The traps were set approximately 15 m apart on a rope groundline. Each semi-conical trap weighed 1.4 kg and measured 76.2 x 30.5 x 71.1 cm, with a stretch mesh size averaging 45 mm.

Fishing Gear Description: Complete details of fishing gear specifications and catches of shrimp and fish by species and abundance are described in Ong et al. (2002) and Troffe et al. (2003) In terms of this study, the three replicate transects to be fished within each gear block were plotted on *Nobeltec Visual Navigation Suite 5.0* before sample collection and the mid-point of each fishing transect was identified as the location for sediment sampling.

Sediment and infaunal sampling: A Van Veen grab (0.1 m²) was used to collect sediment samples for both sediment and infaunal samples. At Turnour Bay, four grab samples were obtained 3.5 h before and 23 h after the trawling. Grab sampling (up to four replicate grabs at each station) was conducted while the traps were deployed at Turnour Bay in March 2002. At Bones Bay, samples were only obtained after trawling; four grab samples were obtained in March 2002, about five months after beam trawling.

Grab deployment: The coordinates for each grab location were recorded from a *dGPS* unit (differential global positioning system) located in the vessel's wheelhouse. At Turnour Bay and Bones Bay, CCGS VECTOR provided a platform for grab sampling. On this vessel a hand held GPS (Garmin GPS 12) was used to obtain the position of the grab sample by holding the GPS unit close to the cable while the grab was being lowered into the water.

Sediment collected from the first two grabs in Turnour Bay was used for macrofaunal processing, while the sediment collected from the third and fourth grab deployments was reserved for the collection of meiofauna and sediment samples to be analysed at the laboratory. Grab samples were obtained during minimal ship-drift when the line used to lower the grab was vertical to obtain an accurate station position. Water depth was recorded from the ship's depth sounder. Prior to sample processing, a photograph of the labelled sediment surface was collected and a description of the grab sample (volume, colour) was documented.

Sediment sampling pre and post trawling: At Turnour Bay, before and after each trawl, four grab sediment samples were collected at the mid-point station on each transect. Multiple attempts of grab deployments were required at all the stations in Turnour Bay before sufficient sample volumes were obtained. Two grab deployments at each mid-transect supported macrofaunal samples, while the other two grab deployments were dedicated for meiofauna taxa and sediment attributes (Table 2). The grab samples were obtained after trawling activity on transects 1 and 3, and approximately 3.5 h after trawling on transect 2.

At Bones Bay, samples for sediment analyses were obtained in January 2002. The grab was deployed 19 times at this location. Sediments from the two macrofaunal grab samples were sieved through two stacked sieves (1.0 cm, 0.5 mm) using sea water. The retained sample on the sieve table was transferred to sample bottles and fixed with buffered formalin. Samples were scanned with a binocular dissecting microscope. Surface sediments from the other two grab samples were obtained to determine grain size. The samples were frozen and later submitted to an analytical laboratory and analyzed using standard methods

Water properties: Bottom-water properties within the two bays were determined from CCGS VECTOR with a rosette sampler. In September 2001, and January, May and December 2002, temperature, salinity, dissolved oxygen, and fluorescence were measured at two stations in Clio Channel, one near Turnour Bay and another near Bones Bay.

3.1 SEDIMENT ATTRIBUTES: MEIOFAUNA, CHLOROPHYLL AND GEOTECHNICAL ANALYSES

Once the grab sample had settled on the boat deck, temperature was measured with a hand-held thermometer inserted into the top 10 cm of the sediment and held in position until the temperature stabilized.

A syringe-core was used to obtain a vertical profile of chlorophyll concentration by carefully inserting the syringe-core into the intact sediment surface of the grab sample. Once the syringe-core was retracted from the sediment, it was capped, sealed, and kept upright in a cooler for storage. An additional syringe-core was also deployed into the grab for meiofauna sampling. Each core was sliced every 1 cm and placed into individual 50-ml jars filled to 1/3 of the jar volume with 4% buffered formalin. Surface sediments were also collected for geotechnical analyses (Table 4) prior to removing the meiofauna and chlorophyll syringe cores from the grab sediment. Sediment samples from the top 2-cm of the grab sediment were obtained using plastic putty knives. The first sample (500 ml) was reserved for grain size analysis, while the second sample (100 ml) was collected for porosity and organic content measurements, carbon and nitrogen content, and trace-element concentrations.

Sediment grain size analysis was conducted by Pacific Soil Analysis according to the size fractions in Table 3. Samples were wet sieved through a 63 μm mesh to separate coarse and fine fractions while percent silt and clay fractions were determined by pipette analysis as outlined by McKeague (1978). Porosity (% water content) was determined at the Pacific Science Enterprise Centre by weighing the sediments before and after drying at 55°C for 2 days and percent organic content was determined by comparing weights of dried and ashed sediment samples after they had been fired in a furnace at 550°C for two hours. Total Carbon, Total Nitrogen, and % Organic Carbon analyses were conducted at the Department of Earth and Ocean Sciences, University of British Columbia using a Carlo-Erba CHN analyser (precision \pm 1.2%) and the concentrations of semi-trace metals (strong acid

leachable) were conducted by Norwest Labs with ultrasonic nebulization and inductively coupled plasma optical emission spectroscopy.

The second syringe-core was removed from the grab sample following the collection of the surface sediments, sealed at the base using a plastic bag and electrician's tape, and stored vertically at -20°C. In the laboratory, chlorophyll concentration was determined with the acidification method as described by Arar and Collins (1997). Once in the lab, the chlorophyll syringe cores were partially thawed while clamp mounted to an apparatus stand. Starting with the surface layer, 0.5 cm increments were extruded with an inverted core plunger and sediments removed with a clean razor blade. Each 0.5 cm sediment increment was transferred to a scintillation vial with 10 ml of 90% acetone. These samples were stored in a dark refrigerator at 4°C for 24 hours. The acetone supernatant of each sample was transferred to a cuvette and the fluorescence readings recorded using Turner Designs 10-AU Fluorometer to measure chlorophyll a (F_a). Two drops of 10% HCL were then mixed into the aqueous extraction and the solution was re-assayed with the fluorometer to measure chlorophyll b (F_b).

3.2 MEIOFAUNA SAMPLING

Meiofauna samples were collected from two Turnour Bay grab samples deployed before and after fishing activity. A 60cc syringe-core was carefully inserted into the deepest portion of the grab sample before the grab was emptied. The syringe-core sample for meiofauna analysis was carefully removed from the grab sample to extrude sediment-core using the syringe plunger. The sediment-core was sliced to provide 1-cm depth intervals of the sediment profile. Each of the 1-cm (5 ml) sediment slices was placed in a labelled jar with 4% formalin. At the laboratory, the meiofauna samples were captured on a 63- μ m mesh to remove formalin and soaked overnight in an alcohol rose-bengal solution. Meiofauna were identified and sorted to 21 different taxonomic categories (Table 5). A binocular dissecting microscope (35-400 X) was used for taxa sorting.

3.3 MACROFAUNAL SAMPLING

Infaunal sieving was conducted using a stack of two sieves (1.0 cm, 0.5 mm mesh size; 70 x 70 cm frame dimensions) set on the deck. The contents of the grabs were transferred into a 65-Litre tote and then gently washed with seawater onto the sieves. Larger invertebrates retained on the 1.0 cm sieve were removed with forceps and put into labelled jars. The washed contents remaining on the 0.5 mm mesh were then transferred to a 0.5 mm Tyler sieve (approx. 20.5 cm diameter) and then into a sample jar and fixed with 10% formalin and seawater. Once in the laboratory, samples were washed through a 0.5 mm sieve, stained with an alcohol rose-bengal solution overnight, and preserved in 70% isopropanol. Macrofauna identification to the lowest taxonomic category possible was conducted under a binocular dissecting scope at 60-500 times resolving power. Damaged animals were only counted as individuals when the head or shell hinge (in the case of bivalves) were present and intact.

4.0 RESULTS

The result section consists of data tables associated with sediment attributes and the diversity of both meiofauna and macrofauna taxa along trawling and trapping transects in Clio Channel.

TABLE 1: Position data associated with beam-trawling at Turnour Bay, January 18-19, 2002. Start and end locations of each trawl-line are presented. PST = Pacific Standard Time; TBBT=Turnour Bay beam trawl; 1,2,3 = replicate trawl transect; * = starting position of trawling transect.

Date	Time (PST)	Trawl	Latitude (°N)	Longitude (°W)	Depth Range (m)	Transect
18-Jan-02	13:37	TBBT1*	50 34.570	126 28.834	40.8 - 46.8	Start*
18-Jan-02	13:47	TBBT1	50 34.465	126 29.098		End
19-Jan-02	10:15	TBBT2*	50 35.010	126 27.312	31.5 - 34.7	Start*
19-Jan-02	10:34	TBBT2	50 35.070	126 27.694		End
18-Jan-02	15:29	TBBT3*	50 34.887	126 27.866	40.6 - 43.3	Start*
18-Jan-02	15:37	TBBT3	50 34.743	126 28.009		End

TABLE 2: Position data for grab deployments at Turnour Bay, January 17-19, 2002. Sediment, meiofauna, and macrofauna samples were collected before (pre) and after (post) beam trawling. TBBT = Turnour Bay beam trawl, 1,2,3 = replicate trawl transects; sed = sediment.

Date	Time (PST)	Location	Sample type	Lat. (°N)	Long. (°W)	Depth (m)	% grab fullness	Temp (°C)	Comments
17-Jan-02	15:38	TBBT1-PRE	A / macrofauna	50 34.547	126 28.940	43.8	18	-	sandy mud, grey brown
17-Jan-02	16:00	TBBT1-PRE	B / macrofauna	50 34.517	126 28.926	41.3	25	-	shells softer mud, grey green.
17-Jan-02	16:06	TBBT1-PRE	C / meiofauna/sed	50 34.527	126 28.941	43.1	15	7	green brown mud
17-Jan-02	16:20	TBBT1-PRE	D / meiofauna/sed	50 34.518	126 28.916	42.1	20	7	sea whip
18-Jan-02	11:52	TBBT3-PRE	A / macrofauna	50 34.823	126 27.922	39.0	18	-	olive green, sea whip (0.914m)
18-Jan-02	12:01	TBBT3-PRE	B / macrofauna	50 34.819	126 27.928	39.9	20	-	olive green with some wood debris
18-Jan-02	12:15	TBBT3-PRE	C / meiofauna/sed	50 34.822	126 27.909	39	15	-	olive green with some slumping
18-Jan-02	12:23	TBBT3-PRE	D / meiofauna/sed	50 34.821	126 27.923	39.5	15	-	olive green with some slumping, only chlorophyll taken
18-Jan-02	18:01	TBBT2-PRE	A / macrofauna	50 35.043	126 27.526	33.4	15	-	sandy gravel and mud
18-Jan-02	18:07	TBBT2-PRE	B / macrofauna	50 35.037	126 27.526	32.6	12	-	grey sand and mud
18-Jan-02	18:11	TBBT2-PRE	C / meiofauna/sed	50 35.041	126 27.527	33.4	15	7	slightly muddy but dominated by sand; sea whip (length-0.94m)
18-Jan-02	18:23	TBBT- PRE	D / meiofauna/sed	50 35.041	126 27.527	32.7	10	7	Sand, gravel, wood bits
19-Jan-02	12:53	TBBT-1-POST	A / macrofauna	50 34.555	126 28.917	42.0	13	-	greyish sand and mud
19-Jan-02	13:02	TBBT-1-POST	B / macrofauna	50 34.544	126 28.935	43.0	18	-	brownish green mud
19-Jan-02	13:18	TBBT-1-POST	C / meiofauna/sed	50 34.572	126 28.952	42.5	18	8	greenish grey sand and mud
19-Jan-02	13:30	TBBT-1-POST	D / meiofauna/sed	50 34.523	126 28.948	41.5	15	7.5	green brown mud and sand
19-Jan-02	14:05	TBBT-2-POST	A / macrofauna	50 35.025	126 27.473	33.0	12	-	
19-Jan-02	14:11	TBBT-2-POST	B / macrofauna	50 35.014	126 27.488	32.0	18	-	Brownish green sandy mud
19-Jan-02	14:37	TBBT-2-POST	C / meiofauna/sed	50 35.032	126 27.543	33.6	14	7	Greenish grey sandy mud
19-Jan-02	14:56	TBBT-2-POST	D / meiofauna/sed	50 35.033	126 27.543	34.0	9	7	Sample kept.
19-Jan-02	15:16	TBBT-3-POST	A / macrofauna	50 34.835	126 27.925	38.6	12	-	greenish grey mud
19-Jan-02	15:21	TBBT-3-POST	B / macrofauna	50 34.830	126 27.928	40.4	12	-	greenish mud and sand
19-Jan-02	15:40	TBBT-3-POST	C / meiofauna/sed	50 34.836	126 27.861	39.8	9	7	greenish brown mud
19-Jan-02	15:50	TBBT-3-POST	D / meiofauna/sed	50 34.833	126 27.937	40.0	9	7	green grey muddy sand

TABLE 3: Percent proportions of sediment grain-size categories collected before (pre) and after (post) beam trawling in Turnour Bay, January 17-19, 2002. TBBT = Turnour Bay beam trawl; 1,2,3 = replicate trawl transects; C,D = sample type (see Table 2).

Date	Location	>2mm	<2mm	<1mm	<.5mm	<.25mm	<.10mm	<63 microns	<4 microns	<2 microns
17-Jan-02	TBBT-1 PRE-C	0	0	0.6	8.3	82.3	2.4	2.5	0.4	3.5
17-Jan-02	TBBT-1 PRE-D	0.2	0.2	1.1	17.8	73.0	1.0	2.5	0.4	3.8
18-Jan-02	TBBT-2 PRE-C	1.30	0.80	3.30	14.40	74.10	1.40	2.10	0.10	2.50
18-Jan-02	TBBT-2 PRE-D	0	1.0	3.4	17.1	72.2	1.6	1.9	0.1	2.7
18-Jan-02	TBBT-3 PRE-C	0.3	1.6	6.5	21	62.1	5.3	0.8	0.8	1.6
18-Jan-02	TBBT-3 PRE-D	0.2	1.5	6.5	18.8	60.7	7	1.6	0.8	2.9
19-Jan-02	TBBT-1 POST-C	0	0	0.6	13.6	79.3	2.2	1.7	0.7	1.9
19-Jan-02	TBBT-1 POST-D	0	0	8	18.0	73.1	1.5	2.3	0.2	4.1
19-Jan-02	TBBT-2 POST-C	7.90	2.60	6.10	17.10	61.20	0.90	1.60	0.20	2.40
19-Jan-02	TBBT-2 POST-D	0.6	1.0	4.9	17.0	69.2	3.2	1.4	0.1	2.6
19-Jan-02	TBBT-3 POST-C	0.6	2.3	8	20.0	59.9	4.6	1.2	1.2	2.2
19-Jan-02	TBBT-3 POST-D	0.1	0.5	2.3	10.7	80.9	1.7	1.3	0.1	2.4

TABLE 4: Water content and organic content of sediment samples collected before (pre) and after (post) beam trawling in Turnour Bay, January 17-19, 2002. TBBT=Turnour Bay beam trawl; 1,2,3 = replicate trawl transect; C,D = sediment sample type (see Table 2).

Date	Location	Organic content (%)	Water content (%)
17-Jan-02	TBBT-1 PRE-C	1.94	30.51
17-Jan-02	TBBT-1 PRE-D	1.90	33.27
18-Jan-02	TBBT-2 PRE-C	1.45	31.15
18-Jan-02	TBBT-2 PRE-D	1.39	32.29
18-Jan-02	TBBT-3 PRE-C	1.00	28.31
18-Jan-02	TBBT-3 PRE-D	1.33	26.71
19-Jan-02	TBBT-1 POST-C	1.01	26.00
19-Jan-02	TBBT-1 POST-D	1.66	32.02
19-Jan-02	TBBT-2 POST-C	1.53	27.97
19-Jan-02	TBBT-2 POST-D	1.24	25.84
19-Jan-02	TBBT-3 POST-C	0.94	24.73
19-Jan-02	TBBT-3 POST-D	0.97	24.33

TABLE 5: Macrofauna abundance from grab samples obtained before (pre) beam trawling in Turnour Bay, January 17-19, 2002. P = present. Note that macrofauna data are standardised per volume of sediment (dm³) with a standard surface area of collection (0.1 m² Van Veen grab). TBBT=Turnour Bay beam trawl; 1,2,3 = replicate trawl transect; A, B = macrofauna sample type (see Table 2).

Taxonomic Division	Genus/Species	17-Jan-02	17-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02
		TBBT-1 PREA ^{1,2}	TBBT-1 PREB	TBBT-2 PREA ¹	TBBT-2 PREB ^{1,3}	TBBT-3 PREA ^{1,4}	TBBT-3 PREB ^{1,5}
P. Annelida							
Ci. Polychaeta							
		3.11 [*]	3.41 [*]	7.15 [*]	9.50 [*]	0	6.83 [*]
	Ampharetidae	0.03	0.01	0.12	0	0	0.06
	Arabellidae	0	0	0	0	0	0.03
	Capitellidae	0.18 ⁶	0.15	1.26 ⁶	1.89 ⁶	0.01	0.2
	Chaetopteridae	0	0.01	0	0	0	0
	Cirratulidae	0.23	0.17	0.49	0.47	0.11	0.43
	Cossuridae	0	0.01	0.01	0.01	0	0
	Dorvilleidae	0.02	0	0	0	0	0
	Glyceridae	0.04	0.03	0.11	0.07	0.03	0.06
	Goniadae	0	0.01	0	0	0	0
	Goniadidae	0	0	0.01	0.01	0	0.01
	Lumbrinaridae	0.19	0.16	0.1	0.07	0.07	0.24
	Magelonidae	0	0	0	0	0	0.01
	Maldanidae	0.35	0.26	0.61	1.22	0.08	0.81
	Nephtyidae	0	0.07	0.20	0	0.03	0.12
	<i>Nephtys sp.</i>	0.07	0	0	0.24	0	0
	Nereidae	0.01	0.03	0.03	0	0	0.07
	Onuphidae	0.03	0.07	0.21	0.14	0.04	0.16
	Orbinidae	0.25	0.17	0.15	0.35	0.03	0.20
	Owenidae	0.68	0.27	3.05	6.83	0.046	1.42
	Paraonidae	0.08	0.33	0.06	0.11	0.12	0.50
	Pectinoridae	0	0	0	0.01	0	0.01
	Phyllodocidae	0.03	0.06	0.10	0.26	0.06	0.04

¹ Unknown egg cases present

² Bryozoa present on shell

³ Unknown organisms (possibly *Urochordata*) present

⁴ Sponge bits present

⁵ *Hydroidea polyps* present

⁶ Miscellaneous pieces

⁶ Possibly *Notomastus tenuis*

TABLE 5: (continued)

Taxonomic Division	Genus/Species	17-Jan-02	17-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02
		TBBT1-A PRE-A	TBBT-1 PRE-B	TBBT-2 PRE-A	TBBT-2 PRE-B	TBBT-3 PRE-A	TBBT-3 PRE-B
Polydoridae		0	0	0	0.10	0	0
Polynoidae		0.10	0.16	0.38	0.39	0.20	0.67
Sabellariidae		0	0.01	0	0.01	0	0
Sabellidae		0.40	0.23	0.50	0.76	0.05	0.46
Scalabregmidae		0.06	0.15	0.01	0	0	0.17
Siglionidae		0	0.01	0	0	0	0
Sphaerodoridae		0.21	0.17	0.31	0.15	0.09	0.24
Spionidae		1.27	0.77	2.19	1.47	0.14	1.5
Syllidae		1.55	0.69	2.89	5.24	0.37	3.59
Terrebelidae		0.02	0.01	0	0.03	0.01	0.01
Trichobranchidae		0	0.01	0.01	0	0	0.04
OTHER		0.01 ¹	0	0.21 ¹	0	0	0
Cl. Oligocheata		0	0	0	0.08	0	0.04
P. Mollusca							
Cl. Gastropoda							
F. Cerithiidae	<i>Bittium sp.</i>	0.04	0.01	0.03	0.06	0.01	0.04
	<i>Bittium munitum</i>	0	0.01	0	0	0	0
F. Columbelloidea	<i>Astyris gausapata</i>	0.07	0.01	0.05	0.10	0.03	0.04
F. Cylichnidae	<i>Cylichna attonsa</i>	0.03	0.01	0	0.06	0.04	0
	<i>Cylichna alba</i>	0	0	0.04	0	0	0
F. Hydatinidae	<i>Parvamplustrum sp.</i>	0.18	0	0	0	0.01	0
F. Muricidae	<i>Ocenebra sp.</i>	0.01 ²	0	0.01	0	0	0
F. Naticidae	<i>Euspira pallida</i>	0	0.01	0	0	0	0
F. Philinidae							
	<i>Philine sp.</i>	0	0	0.01	0	0	0
F. Pyramelloidea	<i>Turbonilla sp.</i>	0.01	0.01	0.06	0.01	0.03	0.09
	<i>Odostomia sp.</i>	0	0	0.07	0	0	0.04

¹ Too small or damaged to positively identify² Possibly *O. orpheus*

TABLE 5: (continued)

Taxonomic Division	Genus/Species	17-Jan-02	17-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02
		TBBT-1 PRE-A	TBBT-1 PRE-B	TBBT-2 PRE-A	TBBT-2 PRE-B	TBBT-3 PRE-A	TBBT-3 PRE-B
F. Rossoidae	<i>Alvania rosana</i>	0.07	0.09	0.05	0.11	0.07	0.09
	<i>Alvania sp.</i>	0	0	0.01	0	0	0.08
F. Trichotropidae	<i>Trichotropis concellata</i>	0	0	0.02	0	0	0
F. Trochidae	<i>Solariella vancouverensis</i>	0.08	0.01	0.05	0.03	0.02	0
F. Turridae	<i>Oenopota sp.</i>	0.01	0	0	0	0	0
OTHER ¹		0.08	0.04	0.26	0.5	0.08	0.48
Cl. Bivalvia							
F. Astartidae	<i>Astarte esquimalti</i>	0	0.01	0	0	0	0
F. Cardiidae	<i>Clinocardium sp.</i>	0	0	0	0.03	0.01	0.01
	<i>Clinocardium nutellii</i>	0	0	0.01	0	0	0
	<i>Nemocardium centifilosum</i>	0	0	0	0.01	0	0
F. Cuspidariidae	<i>Cardiomya pectinata</i>	0.01	0	0.03	0.04	0	0.01
F. Lasaeidae	<i>Rochefortia tumida</i>	0	0.03	0	0	0	0.03
F. Lucinidae	<i>Parvilucina tenuisculpta</i>	1.16	1.14	0.77	0.63	0.72	1.12
	<i>Lucinoma annulatum</i>	0	0.01	0	0	0	0.02
F. Lyonsidae	<i>Lyonsia arenosa</i>	0.01	0	0.02	0.03	0	0
F. Mactridae	<i>Mactrometris polynema</i>	0	0.01	0	0.01	0	0
F. Mytilidae	<i>Mytilus sp.</i>	0.01	0	0	0	0	0
	<i>Solamen columbianum</i>	0	0.01	0.03	0.08	0.01	0.04
	<i>Vilasina seminuda</i>	0	0	0	0.03	0	0
F. Nuculanidae	<i>Nuculana minuta</i>	0.03	0.01	0.02	0.08	0.04	0.06
F. Nuculidae	<i>Eunucula tenuis</i>	0.54	0.43	0.69	0.76	0.31	0.77
	<i>Acila castrensis</i>	0	0.01	0.28	0	0.5	0.57
	<i>Lucinoma annulatum</i>	0	0	0	0	0.04	0
	unknown	0	0	0.63	0	0	0.47
F. Pandoridae	<i>Pandora bilirata</i>	0.03	0.03	0.02	0	0.01	0
F. Pectinidae	<i>Cyclopecten bistriatus</i>	0	0	0.01	0	0	0

¹ Too small or damaged to positively identify

TABLE 5: (continued)

Taxonomic Division	Genus/Species	17-Jan-02	17-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02
		TBBT-1 PRE-A	TBBT-1 PRE-B	TBBT-2 PRE-A	TBBT-2 PRE-B	TBBT-3 PRE-A	TBBT-3 PRE-B
F. Prsiglomidae	<i>Pseudoglomus fragilis</i>	0	0	0	0.03	0	0
F. Tellinidae	<i>Macoma sp.</i>	0.07	0.05	0.93	0.29	0.08	0.62
	<i>Macoma elinata</i>	0.06	0.04	0	0.06	0.12	0.02
	<i>Macoma nasuta</i>	0	0.15	0	0	0	0
F. Thyasiridae	<i>Thyasira flexuosa</i>	0.02	0.01	0	0.01	0	0.01
	<i>Axinopsida serricata</i>	0	0.01	0.05	0	0	0.03
	<i>Nuculana pernula</i>	0	0.01	0	0	0	0
F. Veneridae	<i>Nutricula lordi</i>	0.72	0.33	0.36	0.04	0.17	0.37
	<i>Compsomyax subdiaphana</i>	0.02	0	0	0	0.01	0.01
F. Yoldidae	<i>Yoldia sp.</i>	0.02	0	0	0	0	0.01
	<i>Yoldia seminuda</i>	0.01	0	0	0	0	0.01
OTHER ¹		0.48	0.15	0.52	1.43	2.32	0.46
Cl. Scaphopoda		0	0	0.07	0.17	0.06	0.08
F. Pulsellidae	<i>Pulsellum salishorum</i>	0.34	0.26	0	0	0	0
P. Nematoda		0.29	0.46	0.43	1.21	0.33	2.41
P. Arthropoda							
Cl. Copepoda							
O. Harpacticoida		0.06	0.01	0.01	0.06	0	0
O. Calanoida		0.04	0.01	0.04 ²	0.01	0.01	0
Cl. Ostracoda		0.37	0.12	0.88	0.78	0.09	0.49
Cl. Malacostraca							
O. Amphipoda	S.O. Gammaridea	0.78 ³	0.39	1.27	1.50	0.38	0.58
O. Cumacea		0.25	0.07	0.08	0.31	0.03	0.1
O. Isopoda							
F. Munnidae		0.08	0.01	0.23	0.14	0.01	0.16
F. Asellota		0.01	0	0	0	0	0

¹ Too small or damaged to positively identify² Very damaged³ + 3 embryonic juveniles

TABLE 5: (continued)

Taxonomic Division	Genus/Species	17-Jan-02	17-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02	18-Jan-02
		TBBT-1	TBBT-1	TBBT-2	TBBT-2	TBBT-3	TBBT-3
		PRE-A	PRE-B	PRE-A	PRE-B	PRE-A	PRE-B
Other		0.01 ¹	0	0.12	0.04	0.01	0.01
O. Tanaidacea		0.04	0.01	0.26	0.24	0.02	0.12
Cl. Cirrropedia	<i>Balanus sp.</i>	0.08	0	0.01	0	0	0
P. Nemertea		0.05	0.02	0.04	0.15	0.01	0.04
	parts	0	0	0.06	0	0	0.04
P. Sipuncula		0.01	0.01	0.09	0.14	0.01	0.06
P. Echinodermata							
Cl. Ophiuroidea		0.01	0	0.02	0	0.01	0.01
Cl. Holothuroidea		0	0	0.02	0.01	0	0
P. Cnidaria							
O. Pennatulacea	Juvenile	0.08	0.01	0.07	0.1	0.05	0.17
	Adult	0	0	0	0	0	0.01
	Totals	15.20	11.49	29.32	39.26	7.67	27.92

¹ Too small or damaged to positively identify

TABLE 6: Macrofaunal abundance from grab samples obtained after (post) beam trawling in Turnour Bay, January 19, 2002. P= present. Note that macrofauna data are standardised per volume of sediment (dm³) with a standard surface area of collection (0.1 m² van Veen grab). TBBT=Turnour Bay beam trawl; 1,2,3, = replicate trawl transect; A and B = sediment sample type (see Table 2).

Taxonomic Division	Genus/Species	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02
		TBBT-1 POST-A	TBBT-1 POST-B	TBBT-2 POST-A	TBBT-2 POST-B ¹	TBBT-3 POST-A ²	TBBT-3 POST-B ³
P. Annelida							
Cl. Polychaeta							
		3.28 ³	39.74 ⁴	32.29 ⁴	122.14 ⁴	14.76 ⁴	28.89 ⁴
	Ampharetidae	0.01	0.04	0.03	0.15	0	0.03
	Arabellidae	0	0	0	0	0	0.01
	Capitellidae	0.08	0.21	0.05	0.87	0.02	0.01
	Cirratulidae	0.31	0.25	0.18	0.58	0.12	0.15
	Dorvilleidae	0	0	0	0.01	0	0
	Glyceridae	0.03	0.03	0.05	0.1	0.01	0.02
	Goniadae	0	0	0	0.02	0	0
	Goniadidae	0	0	0.01	0	0	0.01
	Lumbrinaridae	0.23	0.25	0.01	0.16	0.06	0.13
	Magelonidae	0	0	0.01	0.01	0	0.01
	Maldanidae ⁴	0.5	0.56	0.24	2.86	0.07	0.19
	Nephtyidae	0.04	0	0.11	0.08	0.08	0.03
	<i>Nephtys sp.</i>	0	0.09	0	0	0	0
	Nereidae	0.01	0.02	0.01	0.01	0.03	0.02
	Onuphidae	0.04	0.04	0.09	0.12	0.03	0.04
	Orbinidae	0.22	0.35	0.11	0.04	0.1	0.12
	Owenidae	0.62	0.55	1.81	4.76	0.24	0.5
	Paraonidae	0.3	0.25	0.1	0.39	0.1	0.09
	Pectinoridae	0	0	0	0.03	0	0
	Phyllodocidae	0.04	0.03	0.09	0.1	0.02	0.03
	Polynoidae	0.13	0.21	0.11	0.41	0.17	0.24
	Sabellidae	0.39	0.54	0.32	0.32	0.17	0.31
	Scalabregmidae	0.47	0.18	0	0.04	0.19	0.15
	Serpulidae	0.01	0	0	0	0	0

¹ *P. Entoprocta*, *P. Bryozoa*, and *P. Porphyra* present. Misc. egg cases and unknown tissue (possibly *Urochordata*) also present.

² Unknown egg cases present

³ Miscellaneous pieces

⁴ May include *Chaetopteridae*

TABLE 6: (continued)

Taxonomic Division	Genus/Species	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02
		TBBT-1 POST-A	TBBT-1 POST-B	TBBT-2 POST-A	TBBT-2 POST-B	TBBT-3 POST-A	TBBT-3 POST-B
Sphaerodoridae		0.17	0.2	0.09	0.13	0.11	0.25
Spionidae		0.95	2.02	0.87	2.36	0.39	0.5
Syllidae		1.03	1.03	1.07	4.46	0.5	0.83
Terrebelidae		0	0	0.01	0.48	0	0.01
Trichobranchidae		0	0	0	0.06	0	0
OTHER		0.01	0	0	0.08	0	0
Cl. Oligocheata		0	0	0.03	0.04	0	0
P. Mollusca							
Cl. Gastropoda							
F. Atyidae	<i>Hamanoea sp.</i>	0	0	0.01	0	0	0
F. Cerithiidae	<i>Bittium sp.</i>	0.06	0.07	0.05	0	0.01	0.01
F. Columbelloidea	<i>Astyris gausapata</i>	0.11	0.05	0.11	0.03	0.05	0.04
F. Cylichnidae	<i>Cylichna attonsa</i>	0.02	0.01	0	0	0	0.01
	<i>Cylichna alba</i>	0	0.02	0.01	0	0.05	0
	<i>Cylichna sp.</i>	0	0	0.01	0	0	0
F. Eulimidae	<i>Balchis micans</i>	0	0	0	0.01	0	0
F. Naticidae							
	<i>Natica clausa</i>	0	0.01	0	0	0	0
F. Philinidae		0.01 ¹	0	0	0	0	0
F. Pyramelloidea	<i>Turbonilla sp.</i>	0.01	0.02	0	0.01	0	0.01
	<i>Odostomia sp.</i>	0	0.01	0.21	0.07	0.08	0
F. Rossoidea	<i>Alvania rosana</i>	0.08	0.07	0.05	0.01	0.13	0.12
	<i>Alvania sp.</i>	0	0	0	0	0.02	0
F. Trochidae	<i>Solarialla vancouverensis</i>	0.01	0.02	0.01	0	0.01	0.02
OTHER ²		0.04	0.03	0.02	0.11	0.01	0.13
Cl. Bivalvia							

¹ Species uncertain

² Too small/damaged to positively identify.

TABLE 6: (continued)

Taxonomic Division	Genus/Species	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02
		TBBT1 POST-A	TBBT1 POST-B	TBBT2 POST-A	TBBT2 POST-B	TBBT3 POST-A	TBBT3 POST-B
F. Cardiidae	<i>Nemocardium centifilosum</i>	0	0	0	0.01	0	0
F. Cuspidariidae	<i>Cardiomya pectinata</i>	0	0.01	0.01	10.01	0	0
F. Lasaeidae	<i>Rochefortia tumida</i>	0.01	0.02	0	0	0	0.01
F. Lucinidae	<i>Parvilucina tenuisculpta</i>	1.29	1.37	0.39	0.15	0.8	1.1
	<i>Lucinoma annulatum</i>	0	0	0	0.02	0	0
F. Lyonsidae	<i>Lyonsia arenosa</i>	0	0.01	0.01	0	0.01	0
F. Mytilidae:	<i>Mytilus sp.</i>	0	0	0	0.03	0	0
	<i>Solamen columbianum</i>	0.01	0	0.01	0.04	0.01	0.01
	<i>Musculus sp.</i>				0.01		
F. Nuculanidae	<i>Nuculana minuta</i>	0.01	0	0.01	0.04	0.01	0.05
	<i>Nuculana pernula</i>	0.01	0	0	0	0.01	0
F. Nuculidae	<i>Eunucula tenuis</i>	0.48	0.67	0.23	0.08	0.31	0.59
	<i>Acila castrensis</i>	0.01	0.1	0.31	0.24	0.33	0.57
	unknown	0.13	0	0.17	0.4 ¹	0.2	0.12
	<i>Pandora bilirata</i>	0.01	0.04	0.03	0	0.01	0.01
F. Prsiglomidae	<i>Axinopsida serricata</i>	0	0	0.01	0	0	0
F. Tellinidae	<i>Macoma sp.</i>	0.01	0.06	0.25	0.19	0.37	0.08
	<i>Macoma carlottenses</i>	0.01	0.1	0.01	0	0	0.09
	<i>Macoma crussula</i>	0	0	0	0	0.01	0
	<i>Macoma identata</i>	0.02	0	0	0	0	0
	<i>Macoma elimata</i>	0.06	0.05	0.01	0.05	0	0.05
	<i>Macoma nasuta</i>	0.01	0	0.01	0	0	0.05
F. Thraciidae	<i>Thracia (Ixartia) curta</i>	0.01	0	0	0	0	0
F. Thyasiridae	<i>Thyasira flexuosa</i>	0	0	0.01	0	0	0
	<i>Axinopsida serricata</i>	0	0	0	0	0.01	0

¹ Too young to positively identify

TABLE 6: (continued)

Taxonomic Division	Genus/Species	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02
		TBBT--1 POST-A	TBBT-1 POST-B	TBBT-2 POST-A	TBBT-2 POST-B	TBBT-3 POST-A	TBBT-3 POST-B
	<i>Adontorhina cyclicia</i>	0.01	0	0	0	0	0
	<i>Mendicula ferruginosa</i>	0	0	0.01	0	0	0
F. Veneridae	<i>Nutricola lordi</i>	0.49	0.66	0.09	0	0.25	0.29
	<i>Compsomyax subdiaphana</i>	0.01	0.01	0.01	0	0	0.01
OTHER ¹		0.41	0.1	0.11	0.24	0.74	0.15
Cl. Scaphopoda		0.3	0.32	0	0	0.1	0.09
P. Nematoda		0.38	0.54	0.15	1.2	0.19	0.36
P. Arthropoda							
Cl. Copepoda							
O. Harpacticoida		0.05	0.06	0.01	0.02	0	0.01
O. Calanoida		0.01 ²	0.02	0.03	0.01	0.31 ³	0.09
O. Poecilostomatoida		0	0	0	0.02	0	0
Cl. Ostracoda		0.15	0.19	0.3	0.3	0.19	0.17
Cl. Malacostraca							
O. Amphipoda	S.O. Gammeridea	1.00	0.94	0.63 ⁴	1.38	0.38	0.55
O. Cumacea		0.1	0.22	0.07	0.09	0.03	0.05
O. Decapoda	<i>unknown crabs</i>	0.01	0	0	0	0	0
	<i>Penixa eberna</i>	0	0	0	0	0	0
O. Isopoda							
F. Munnidae		0.07	0.06	0.07	0.08	0.01	0.05
F. Asellota		0	0	0	0	0.01	0
other		0.01	0	0.14 ⁵	0.03	0.01	0.03
O. Tanaidacea		0.01	0.03	0.02	0.11	0.06	0.05 ⁶
Unknown arthropod		0	0	0	0.06	0	0
P. Nemertea		0.01	0.01	0.01	0.09	0	0.02

¹ Too small/damaged to positively identify² Molt³ Many quite damaged⁴ +1 embryonic⁵ +1 molt⁶ One with brood

TABLE 6: (continued)

Taxonomic Division	Genus/Species	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02
		TBBT-1	TBBT-1	TBBT-2	TBBT-2	TBBT-3	TBBT-3
		POST-A	POST-B	POST-A	POST-B	POST-A	POST-B
	parts	0	0.04	0.05	0.03	0	0.01
P. Sipuncula		0.01	0	0.02	0.23	0.01	0.01
P. Echinodermata		0	0	0	0	0	0
Cl. Ophiuroidea		0.01	0.02	0	0	0	0
Cl. Holothuroidea		0.01	0	0	0	0.01	0
P. Cnidaria		0	0	0	0	0	0
O. Pennatulacea	Juvenile	0.01	0.08	0.04	0.08	0.11	0.09 ¹
Totals		5.25	5.72	3.25	4.97	4.45	4.73

¹ *Haipteris willemosi*

TABLE 7: Meiofauna taxonomic abbreviations referred to in Tables 11, 12, and 13.

Abbreviated Name	Taxa	Stage
ACAR	Acari	adult
AMPH	Amphipoda	adult
BIVA	Bivalvia	larvae
CALN	Calanoida	adult
COPN	Copepoda	nauplii
CUMA	Cumacea	adult
EGG	Invertebrate	
FORA	Foraminifera	
GASA	Gastropoda	adult
HARP	Harpacticoida	adult
HYDR	Hydroida	sessile
ISOP	Isopoda	adult
KINO	Kinorhyncha	adult
NEMA	Nematoda	adult
NEMR	Nemertea	adult
OLIG	Oligochaeta	adult
OSTR	Ostracoda	adult
POLA	Polychaeta	adult
ROTI	Rotifera	adult
SCAP	Scaphopoda	adult
TURB	Turbellaria	adult

TABLE 8: Meiofaunal abundance (No. ml⁻¹) sampled from grab samples deployed before (pre) and after (post) beam trawling in Turnour Bay, January 17-19, 2002. Refer to Table 5 for taxa abbreviations. Each core-depth interval was 1-cm in depth, 5.5 ml in volume, and 5.5 cm² in surface area. TBBT= Turnour Bay beam trawl; 1,2,3 = replicate trawl transects; C,D = sample type (see Table 2).

Date	Location	Depth (cm)	Taxonomic Division																			
			FORA	HYDR	TURB	KINO	ROTI	NEMA	OLIG	POLA	BIVA	GASA	SCAP	NEMR	ACAR	HARP	COPN	OSTR	ISOP	AMPH	CUMA	EGG
17-Jan-02	TBBT-1 PRE-C	1	0.0	0.0	0.0	7.3	0.0	67.3	0.0	6.9	2.5	0.0	0.0	0.0	0.0	19.0	4.5	2.4	0.0	0.0	0.0	0.2
17-Jan-02	TBBT-1 PRE-C	2	0.2	0.0	0.0	0.2	0.0	25.2	0.0	1.3	0.0	0.0	0.0	0.0	0.0	0.2	0.2	0.0	0.0	0.0	0.0	0.4
17-Jan-02	TBBT-1 PRE-C	3	0.0	0.0	0.0	0.5	0.0	30.5	0.0	0.2	0.0	0.0	0.0	0.0	0.0	1.1	0.0	0.0	0.0	0.0	0.0	102.1
17-Jan-02	TBBT-1 PRE-C	4	0.2	0.0	0.0	0.0	0.0	19.9	0.0	0.4	0.0	0.0	0.0	0.0	0.0	0.0	0.2	0.0	0.0	0.0	0.0	102.1
17-Jan-02	TBBT-1 PRE-C	5	0.4	0.0	0.0	0.0	0.0	35.9	0.0	0.5	0.2	0.0	0.0	0.0	0.0	0.2	0.0	0.2	0.0	0.0	0.0	1.5
17-Jan-02	TBBT-1 PRE-C	6	0.0	0.0	0.0	0.0	0.0	32.5	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.2
17-Jan-02	TBBT-1 PRE-D	1	0.7	0.0	0.0	3.6	0.0	43.2	0.0	5.1	0.9	0.0	0.0	0.2	0.0	4.2	1.6	1.1	0.0	0.0	0.2	0.5
17-Jan-02	TBBT-1 PRE-D	2	0.0	0.0	0.0	0.4	0.0	20.1	0.0	0.5	0.0	0.0	0.0	0.0	0.0	0.4	0.0	0.0	0.0	0.0	0.0	0.2
17-Jan-02	TBBT-1 PRE-D	3	1.3	0.0	0.0	0.0	0.0	25.9	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.4	0.2	0.0	0.0	0.0	0.0	0.2
17-Jan-02	TBBT-1 PRE-D	4	0.2	0.0	0.0	0.0	0.0	4.5	0.0	0.2	0.0	0.0	0.0	0.0	0.0	1.8	0.2	0.0	0.0	0.0	0.0	0.0
17-Jan-02	TBBT-1 PRE-D	5	0.0	0.0	0.0	0.0	0.0	5.8	0.0	0.4	0.0	0.0	0.0	0.0	0.0	0.5	0.0	0.0	0.0	0.0	0.0	0.0
17-Jan-02	TBBT-1 PRE-D	6	0.2	0.0	0.0	0.2	0.0	6.7	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
19-Jan-02	TBBT-1 POST-C	1	0.0	0.0	0.0	0.5	0.0	24.1	0.0	3.6	1.1	0.0	0.0	0.0	0.0	3.4	1.1	0.4	0.0	0.0	0.0	0.0
19-Jan-02	TBBT-1 POST-C	2	0.0	0.0	0.0	0.4	0.0	13.4	0.0	0.0	0.4	0.0	0.0	0.0	0.0	0.2	0.0	0.7	0.0	0.0	0.0	0.0
19-Jan-02	TBBT-1 POST-C	3	0.0	0.0	0.0	0.4	0.0	8.7	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.4	0.4	0.0	0.0	0.0	0.0	0.5
19-Jan-02	TBBT-1 POST-C	4	0.4	0.0	0.0	0.4	0.0	8.3	0.0	0.0	0.0	0.0	0.0	0.0	0.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0
19-Jan-02	TBBT-1 POST-C	5	0.5	0.2	0.0	0.0	0.0	8.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.4	0.0	0.0	0.0	0.0	0.0	0.5
19-Jan-02	TBBT-1 POST-D	1	0.9	0.0	0.0	2.0	0.0	41.5	0.0	2.2	1.1	0.0	0.2	0.0	0.2	5.6	1.6	0.5	0.0	0.0	0.2	0.4
19-Jan-02	TBBT-1 POST-D	2	0.5	0.0	0.0	0.4	0.0	43.3	0.0	0.4	1.1	0.0	0.0	0.0	0.0	0.2	0.0	0.4	0.0	0.0	0.0	0.4
19-Jan-02	TBBT-1 POST-D	3	0.2	0.0	0.0	0.0	0.0	22.5	0.0	0.0	0.5	0.2	0.0	0.0	0.0	0.2	0.0	0.0	0.0	0.2	0.0	0.5
19-Jan-02	TBBT-1 POST-D	4	0.4	0.0	0.0	0.0	0.0	20.5	0.0	0.2	0.4	0.0	0.0	0.0	0.0	2.0	0.0	0.0	0.0	0.2	0.0	0.4
19-Jan-02	TBBT-1 POST-D	5	0.2	0.0	0.0	0.0	0.0	16.7	0.0	0.4	0.0	0.0	0.0	0.0	0.0	0.0	0.2	0.0	0.0	0.0	0.0	0.2
19-Jan-02	TBBT-1 POST-D	6	0.4	0.0	0.0	0.0	0.0	14.5	0.0	0.2	0.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
18-Jan-02	TBBT-2 PRE-C	1	14.3	0.0	0.0	3.8	0.0	103.0	0.0	7.6	1.3	0.2	0.0	0.2	0.0	19.0	8.9	2.7	0.0	0.4	0.0	0.7
18-Jan-02	TBBT-2 PRE-C	2	6.9	0.0	0.0	0.9	0.0	85.6	0.0	6.3	0.4	0.0	0.0	0.2	0.0	4.5	1.6	0.9	0.0	0.0	0.0	2.2
18-Jan-02	TBBT-2 PRE-D	1	9.2	0.0	0.0	2.4	0.0	108.6	0.0	9.6	2.0	0.4	0.0	0.4	0.0	15.4	5.3	2.2	0.0	0.0	0.0	1.6
18-Jan-02	TBBT-2 PRE-D	2	10.5	0.0	0.0	0.2	0.0	50.6	0.0	3.4	0.4	0.2	0.2	0.0	0.0	1.1	0.5	0.5	0.0	0.0	0.0	1.3
18-Jan-02	TBBT-2 PRE-D	3	2.4	0.0	0.0	0.2	0.0	25.4	0.0	1.5	0.5	0.0	0.0	0.0	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.2
19-Jan-02	TBBT-2 POST-C	1	2.5	0.4	0.2	3.8	0.2	62.9	0.0	10.0	2.9	0.0	0.0	0.0	0.0	21.8	6.3	2.2	0.4	0.2	0.2	2.2
19-Jan-02	TBBT-2 POST-C	2	5.3	0.0	0.2	0.5	0.0	67.1	0.7	7.4	0.9	0.0	0.0	0.0	0.0	7.4	0.2	2.5	0.0	0.0	0.0	1.1
19-Jan-02	TBBT-2 POST-C	3	3.1	0.0	0.0	0.0	0.0	36.6	0.0	2.9	0.5	0.0	0.0	0.0	0.0	4.4	1.8	0.0	0.0	0.0	0.0	0.4
19-Jan-02	TBBT-2 POST-C	4	3.3	0.0	0.0	0.0	0.0	31.0	0.0	1.5	0.9	0.0	0.0	0.0	0.0	2.0	0.0	1.1	0.0	0.0	0.0	0.4
19-Jan-02	TBBT-2 POST-D	1	0.5	0.0	0.0	3.4	0.0	61.3	0.0	6.5	1.1	0.0	0.0	0.0	0.0	17.2	4.5	3.1	0.2	0.0	0.2	1.3
19-Jan-02	TBBT-2 POST-D	2	11.2	0.0	0.0	1.5	0.0	74.3	0.0	4.9	0.0	0.0	0.0	0.0	0.2	4.5	0.5	3.8	0.4	0.0	0.2	0.5
19-Jan-02	TBBT-2 POST-D	3	9.1	0.0	0.0	0.0	0.0	50.8	0.0	1.5	0.2	0.0	0.0	0.0	0.0	2.2	0.2	0.5	0.0	0.0	0.0	0.5

TABLE 8: (continued)

Taxonomic Division

Date	Location	Depth (cm)	FORA	HYDR	TURB	KINO	ROTI	NEMA	OLIG	POLA	BIVA	GASA	SCAP	NEMR	ACAR	HARP	COPN	OSTR	ISOP	AMPH	CUMA	EGG
18-Jan-02	TBBT-3 PRE-C	1	0.2	0.0	0.0	2.5	0.0	40.1	0.0	1.5	0.9	0.0	0.0	0.0	0.0	2.7	0.0	1.8	0.0	0.4	0.0	0.5
18-Jan-02	TBBT-3 PRE-C	2	1.6	0.0	0.0	0.2	0.0	56.8	0.0	0.9	0.4	0.0	0.0	0.0	0.0	0.5	0.4	1.3	0.0	0.2	0.0	0.2
18-Jan-02	TBBT-3 PREC	3	0.5	0.0	0.0	0.9	0.0	31.0	0.0	0.4	0.2	0.0	0.0	0.0	0.0	0.4	0.0	0.5	0.0	0.0	0.0	0.0
18-Jan-02	TBBT-3 PREC	4	0.5	0.0	0.0	0.4	0.0	18.9	0.0	0.5	0.0	0.0	0.0	0.0	0.0	0.4	0.0	2.0	0.0	0.0	0.0	0.9
18-Jan-02	TBBT-3 PREC	5	0.9	0.0	0.0	0.0	0.0	22.8	0.0	0.5	0.0	0.0	0.0	0.0	0.0	0.5	0.4	0.7	0.0	0.0	0.0	0.2
18-Jan-02	TBBT-3 PRED	1	2.5	0.0	0.0	2.9	0.0	63.1	0.0	10.2	3.4	0.4	0.0	0.0	0.0	18.1	4.5	2.9	0.0	0.0	0.0	0.7
18-Jan-02	TBBT-3 PRED	2	0.9	0.0	0.2	1.8	0.0	43.5	0.2	3.4	0.7	0.0	0.0	0.0	0.0	2.2	0.0	1.6	0.0	0.0	0.2	0.5
18-Jan-02	TBBT-3 PRED	3	3.4	0.0	0.0	0.2	0.0	26.7	0.0	1.6	0.2	0.0	0.0	0.0	0.0	1.5	0.5	1.6	0.0	0.0	0.0	2.2
18-Jan-02	TBBT-3 PRED	4	0.5	0.0	0.0	0.0	0.0	8.2	0.0	0.4	0.0	0.2	0.0	0.0	0.0	0.2	0.0	0.0	0.0	0.0	0.0	38.4
19-Jan-02	TBBT-3 POSTC	1	2.0	0.0	0.0	0.2	0.0	11.4	0.0	2.0	0.0	0.0	0.0	0.0	0.0	1.5	0.2	2.5	0.0	0.0	0.0	1.8
19-Jan-02	TBBT-3 POSTC	2	3.3	16.5	0.0	0.4	0.0	6.5	0.0	0.5	0.0	0.2	0.0	0.0	0.0	0.4	0.2	0.4	0.0	0.2	0.0	0.2
19-Jan-02	TBBT-3 POSTC	3	2.0	0.0	0.0	0.0	0.0	8.0	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.5	0.2	0.2	0.0	0.0	0.0	0.4
19-Jan-02	TBBT-3 POSTC	4	0.2	0.0	0.0	0.0	0.0	17.6	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.0
19-Jan-02	TBBT-3 POSTC	5	0.2	0.0	0.2	0.2	0.0	11.1	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
19-Jan-02	TBBT-3 POSTD	1	1.5	0.0	0.2	1.1	0.0	62.2	0.0	0.9	0.2	0.0	0.0	0.0	0.0	0.5	0.4	0.9	0.0	0.0	0.0	0.4
19-Jan-02	TBBT-3 POST-D	2	1.1	0.0	0.7	4.2	0.0	103.7	0.0	8.7	1.5	0.0	0.0	0.0	0.0	12.9	2.9	4.7	0.0	0.5	0.4	9.2
19-Jan-02	TBBT-3 POST-D	3	2.0	0.0	0.0	0.2	0.0	59.5	0.0	2.4	0.7	0.0	0.0	0.0	0.0	5.6	0.9	0.9	0.0	0.0	0.0	0.7

TABLE 9: Carbon (C) and Nitrogen (N) content of sediment samples collected from Turnour Bay collected before (pre) and after (post) beam trawling, January 17-19, 2002. TBBT = Turnour Bay Beam Trawl; 1,2,3 = replicate trawl transects; C,D = sample type (see Table 2).

Date	Location	%Total N	%Total C	% Org C
17-Jan-02	TBBT-1 PRE C	0.06	0.52	0.46
17-Jan-02	TBBT-1 PRE D	0.07	0.75	0.67
18-Jan-02	TTBT-2 PRE C	0.04	0.37	0.35
18-Jan-02	TTBT-2 PRE D	0.04	0.33	0.31
18-Jan-02	TTBT-3 PRE C	0.04	0.36	0.35
18-Jan-02	TTBT-3 PRE D	0.05	0.41	0.39
19-Jan-02	TBBT-1 POST C	0.03	0.28	0.26
19-Jan-02	TBBT-1 POST D	0.06	0.72	0.65
19-Jan-02	TBBT-2 POST C	0.04	0.41	0.38
19-Jan-02	TBBT-2 POST D	0.04	0.33	0.31
19-Jan-02	TTBT-3 POST C	0.03	0.20	0.19
19-Jan-02	TTBT-3 POST D	0.04	0.28	0.26

TABLE 10: Sediment chlorophyll concentration ($\mu\text{g mL}^{-1}$ sediment) of grab samples collected from Turnour Bay collected before (pre) and after (post) beam trawling, January 17-19, 2002. Depth refers to core depth-interval. TBBT = Turnour Bay Beam Trawl; 1,2,3 = replicate trawl transects; C,D = sample type (see Table 2).

Date	Location	Depth (mm)	[chl-a] ($\mu\text{g mL}^{-1}$)	[pheo] ($\mu\text{g mL}^{-1}$)
17-Jan-02	TBBT-1 PRE C	0-5	0.346	2.890
17-Jan-02	TBBT-1 PRE C	5-10	0.354	2.708
17-Jan-02	TBBT-1 PRE C	10-15	0.404	2.519
17-Jan-02	TBBT-1 PRE C	15-20	0.375	2.613
17-Jan-02	TBBT-1 PRE C	20-25	0.368	2.438
17-Jan-02	TBBT-1 PRE D	0-5	0.361	2.635
17-Jan-02	TBBT-1 PRE D	5-10	0.382	2.387
17-Jan-02	TBBT-1 PRE D	10-15	0.281	2.466
17-Jan-02	TBBT-1 PRE D	15-20	0.318	2.598
17-Jan-02	TBBT-1 PRE D	20-25	0.354	2.489
18-Jan-02	TBBT-2 PRE C	0-5	0.960	6.438
18-Jan-02	TBBT-2 PRE C	5-10	1.313	7.513
18-Jan-02	TBBT-2 PRE C	10-15	0.411	2.818
18-Jan-02	TBBT-2 PRE C	15-20	0.447	2.723
18-Jan-02	TBBT-2 PRE C	20-25	0.433	2.636
18-Jan-02	TBBT-2 PRE D	0-5	0.390	2.919
18-Jan-02	TBBT-2 PRE D	5-10	0.404	2.919
18-Jan-02	TBBT-2 PRE D	10-15	0.411	2.694
18-Jan-02	TBBT-2 PRE D	15-20	0.397	2.599
18-Jan-02	TBBT-2 PRE D	20-25	0.339	2.591
18-Jan-02	TBBT-3 PRE C	0-5	0.404	3.007
18-Jan-02	TBBT-3 PRE C	5-10	0.303	2.634
18-Jan-02	TBBT-3 PRE C	10-15	0.245	2.123
18-Jan-02	TBBT-3 PRE C	15-20	0.231	1.978
18-Jan-02	TBBT-3 PRE C	20-25	0.260	1.912
18-Jan-02	TBBT-3 PRE D	0-5	0.568	2.686
18-Jan-02	TBBT-3 PRE D	5-10	0.114	0.808
18-Jan-02	TBBT-3 PRE D	10-15	0.118	0.749

TABLE 10: (continued)

Date	Location	Depth (mm)	[chl-a] ($\mu\text{g mL}^{-1}$)	[pheo] ($\mu\text{g mL}^{-1}$)
18-Jan-02	TBBT-3 PRE D	15-20	0.132	0.736
18-Jan-02	TBBT-3 PRE D	20-25	0.111	0.681
19-Jan-02	TBBT-1 POST C	0-5	0.361	3.007
19-Jan-02	TBBT-1 POST C	5--10	0.253	2.430
19-Jan-02	TBBT-1 POST C	10--15	0.339	2.489
19-Jan-02	TBBT-1 POST C	15--20	0.296	2.379
19-Jan-02	TBBT-1 POST C	20-25	0.274	2.510
19-Jan-02	TBBT-1 POST D	0-5	0.361	2.861
19-Jan-02	TBBT-1 POST D	5-10	0.354	3.036
19-Jan-02	TBBT-1 POST D	10-15	0.390	2.810
19-Jan-02	TBBT-1 POST D	15-20	0.332	2.722
19-Jan-02	TBBT-1 POST D	20-25	0.354	2.729
19-Jan-02	TBBT-2 POST C	0-5	0.361	3.174
19-Jan-02	TBBT-2 POST C	5-10	0.375	2.912
19-Jan-02	TBBT-2 POST C	10-15	0.267	2.707
19-Jan-02	TBBT-2 POST C	15-20	0.375	2.504
19-Jan-02	TBBT-2 POST C	20-25	0.375	2.438
19-Jan-02	TBBT-2 POST D	0-5	0.390	2.970
19-Jan-02	TBBT-2 POST D	5-10	0.346	3.174
19-Jan-02	TBBT-2 POST D	10-15	0.354	3.057
19-Jan-02	TBBT-2 POST D	15-20	0.426	2.759
19-Jan-02	TBBT-2 POST D	20-25	0.318	2.707
19-Jan-02	TBBT-3 POST C	0-5	0.303	3.013
19-Jan-02	TBBT-3 POST C	5-10	0.260	2.167
19-Jan-02	TBBT-3 POST C	10-15	0.267	2.182
19-Jan-02	TBBT-3 POST-C	15-20	0.267	2.109
19-Jan-02	TBBT-3 POST C	20-25	0.318	2.496
19-Jan-02	TBBT-3 POST D	0-5	0.397	3.065
19-Jan-02	TBBT-3 POST D	5-10	0.411	2.992
19-Jan-02	TBBT-3 POST D	10-15	0.339	3.057
19-Jan-02	TBBT-3 POST D	15-20	0.361	2.657

TABLE 11: Concentrations of trace-elements ($\mu\text{g g}^{-1}$ dry weight sediment) for sediments at Turnour Bay collected before (pre) and after (post) beam trawling January 17-19, 2002. TBBT = Turnour Bay Beam Trawl; 1,2,3 = replicate trawl transects.

	17-Jan-02	18-Jan-02	18-Jan-02	19-Jan-02	19-Jan-02	19-Jan-02
	TBBT-1	TBBT-2	TBBT-3	TBBT-1	TBBT-2	TBBT-3
	PRE	PRE	PRE	POST	POST	POST
Aluminium	4800	4230	4780	4970	4540	4740
Antimony	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0
Arsenic	<1.0	3.5	1	5	1	1.5
Barium	14.4	11.9	13.5	11.7	13.7	12.3
Beryllium	<0.02	0.035	0.02	0.03	<0.02	0.04
Bismuth	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0
Cadmium	0.427	0.434	0.463	0.469	0.414	0.48
Calcium	8750	4020	4330	4500	4460	4620
Chromium	4.22	4.04	4.28	4.94	4.79	4.5
Cobalt	1.59	1.65	1.89	1.99	1.79	1.95
Copper	5.47	3.74	4.18	4.19	4.14	3.9
Iron	6460	4990	4930	5730	5330	4950
Lead	<0.2	0.85	<0.2	0.65	0.9	1.1
Lithium	2.4	2.4	2.2	2.3	2.54	2.1
Magnesium	2050	1890	2030	2000	2070	2040
Manganese	55.2	54.3	61.3	58.8	66.3	60.9
Molybdenum	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5
Nickel	2.63	2.59	2.79	3.14	2.79	2.8
Phosphorus	390	449	447	381	465	504
Potassium	890	753	772	798	877	719
Selenium	<2	<2	<2	<2	<2	<2
Silicon	39.8	19	22	20	21	33.5
Silver	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
Sodium	7160	4180	3850	3690	3950	3620
Strontium	27.1	21.1	21.9	23.3	23	23.8
Sulphur	850	1050	618	628	623	579
Thorium	<0.2	<0.2	<0.2	<0.2	<0.2	1.7
Tin	<0.2	0.45	<0.2	0.3	0.6	0.45
Titanium	302	268	321	336	296	336
Uranium	<3	<3	<3	<3	<3	<3
Vanadium	11.7	11.6	11.7	13.2	12.3	12.3
Zinc	10.1	9.62	9.91	9.77	10.4	9.69
Zirconium	0.84	0.95	1.1	1.2	1	1.2

TABLE 12: Position data and duration for trapping at Turnour Bay, March 11-13, 2002. Start and end locations of traps for each trap line presented. TTR = Turnour Bay trap lines; 1,2,3 = replicate trap-line transects.

Date	Time (PST)	Location	Lat. (°N)	Long. (°W)	Depth Range (m)	Comments	Line Retrieval
11-Mar-02	17:30	TTR-1	50 35.332	126 26.946		start trap	
12-Mar-02	8:45	TTR-1	50 34.530	126 28.867	67.6-78.6	end trap	12-Mar-02 07:53
12-Mar-02	9:04	TTR-3	50 34.908	126 27.845		start trap	
12-Mar-02	9:09	TTR-3	50 34.575	126 28.138	38.0	end trap	12-Mar-02 09:55
12-Mar-02	10:05	TTR-2	50 35.038	126 27.799		start trap	
12-Mar-02	10:12	TTR-2	50 35.039	126 27.331	34.0	end trap	12-Mar-02 10:52
12-Mar-02	11:14	TTR-1	50 34.562	126 28.733		start trap	
12-Mar-02	11:19	TTR-1	50 34.399	126 29.305	41.3	end trap	12-Mar-02 12:28
12-Mar-02	12:42	TTR-3	50 34.709	126 28.163		start trap	
12-Mar-02	12:48	TTR-3	50 34.937	126 27.765	37.0	end trap	12-Mar-02 14:45
12-Mar-02	14:51	TTR-2	50 35.072	126 27.814		start trap	
12-Mar-02	14:57	TTR-2	50 35.014	126 27.242	31.0	end trap	12-Mar-02 15:58
12-Mar-02	16:14	TTR-1	50 34.554	126 28.827		start trap	
12-Mar-02	16:16	TTR-1	50 34.371	126 29.432	46.0-58.0	end trap	12-Mar-02 17:12
12-Mar-02	17:23	TTR-3	50 34.692	126 28.110		start trap	
12-Mar-02	17:28	TTR-3	50 35.045	126 27.787	37.5	end trap	13-Mar-02 08:50
13-Mar-02	9:01	TTR-2	50 35.076	126 27.768		start trap	
13-Mar-02	9:06	TTR-2	50 35.008	126 27.089	39.5	end trap	13-Mar-02 11:31
13-Mar-02	11:40	TTR-1	50 34.558	126 28.815		start trap	
13-Mar-02	11:46	TTR-1	50 34.374	126 29.359		end trap	13-Mar-02 14:13
13-Mar-02	14:20	TTR-3	50 34.986	126 27.813		start trap	
13-Mar-02	14:26	TTR-3	50 34.650	126 28.114		end trap	13-Mar-02 15:20
13-Mar-02	15:31	TTR-2	50 35.083	126 27.821		start trap	
13-Mar-02	15:36	TTR-2	50 35.034	126 27.244		end trap	13-Mar-02 16:22
13-Mar-02	16:37	TTR-1	50 34.565	126 28.752		start trap	
13-Mar-02	16:44	TTR-1	50 34.398	126 29.296		end trap	13-Mar-02 17:33
13-Mar-02	17:41	TTR-3	50 34.644	126 28.085	33.2	start trap	
13-Mar-02	17:45	TTR-3	50 35.018	126 27.775		end trap	13-Mar-02 18:42
13-Mar-02	18:47	TTR-2	50 35.089	126 27.743		start trap	
13-Mar-02	18:51	TTR-2	50 35.000	126 27.103		end trap	14-Mar-02 08:49

TABLE 13: Position data for sediment chemistry grab locations in Turnour Bay, March 12-13, 2002. T = grabs.

Date	Time (PST)	Location	Sample Type	Lat. (°N)	Long. (°W)	Depth (m)	% grab fullness	Comments
13-Mar-02	9:43	T1	A/chemistry	50 34.478	126 29.055	37.5	30	brown / olive
13-Mar-02	9:56	T1	B/chemistry	50 34.521	126 28.909	37.8	30	brown / olive . Sea whip 1.77m
13-Mar-02	10:07	T1	C/chemistry	50 34.470	126 29.087	43.3	20	brown / olive
13-Mar-02	15:50	T1	D/chemistry	50 34.480	126 29.056		60	brown olive
12-Mar-02	10:22	T2	A/chemistry	50 35.053	126 27.515	34	10	sandy olive green
13-Mar-02	10:45	T2	B/chemistry	50 35.048	126 27.496	33.1	30	brown / olive , seawhip attached
13-Mar-02	11:02	T2	C/chemistry	50 35.043	126 27.455	31.8	35	brown / olive
13-Mar-02	14:51	T2	D/chemistry	50 35.046	126 27.454	32.2	20	olive brown
13-Mar-02	9:29	T3	A/chemistry	50 34.807	126 27.951	39.5	50	
13-Mar-02	11:40	T3	B/chemistry	50 34.742	126 28.004	43.1	20	brown / olive
13-Mar-02	11:45	T3	C/chemistry			43.1	10	brown / olive
13-Mar-02	12:08	T3	D/chemistry	50 34.860	126 27.914	42.6	15	brown / olive

TABLE 14: Percent proportions of grain size categories from sediment samples collected during trapping (T) in Turnour Bay, between March 12-13, 2002. 1,2,3 = Trap line replicate transects.

Date	Location	Sample Type	>2mm	<2mm	<1mm	<.5mm	<.25mm	<.10mm	<63µm	<4µm	<2 µm
13-Mar-02	T1	A/chemistry		0.1	0.6	8.1	71.5	10.6	4.8	0.9	3.4
13-Mar-02	T1	B/chemistry		0.1	0.3	5.5	76.1	9.8	4.5	1.4	2.3
13-Mar-02	T1	C/chemistry			0.4	14.7	75.1	6.6	1.6	0.2	1.4
13-Mar-02	T1	D/chemistry			0.5	11	72.3	10.4	2.8	0.5	2.5
12-Mar-02	T2	A/chemistry	0.4	0.6	2.7	13.6	70.8	7	2.6	0.7	1.6
13-Mar-02	T2	B/chemistry	1.2	1.6	5.6	20.8	60.9	5	2.6	0.2	2.1
13-Mar-02	T2	C/chemistry	2.7	2.9	8.4	21.4	53.2	6.7	2.6	0.2	1.9
13-Mar-02	T2	D/chemistry	4.1	2.3	6.5	21.8	55.2	5.4	2.5	0.5	1.7
13-Mar-02	T3	A/chemistry		0.5	2.9	12.1	75.2	6.1	1.6	0.2	1.4
13-Mar-02	T3	B/chemistry			0.8	6.7	81.6	5.8	2.8	0.2	2.1
13-Mar-02	T3	C/chemistry		0.2	1	7	80.9	5.7	2.6	0.5	2.1
13-Mar-02	T3	D/chemistry	0.3	1	2.9	8.3	77.4	5.6	2.6	0.5	1.4

TABLE 15: Position data for macrofauna and meiofauna/sediment grab locations at Bones Bay on, 17 January, 2002. BBBT = Bones Bay Beam Trawl. 1,2,3 = replicate transects.

Date	Time (PST)	Location	Sample type	Lat. (°N)	Long. (°W)	Depth (m)	% grab fullness	Temp (°C)	Comments
17-Jan-02	08:53	BBBT-1	A/macrofauna	50 35.363	126 22.532	74	70	-	Olive green unconsolidated mud
17-Jan-02	09:02	BBBT-1	B/macrofauna	50 35.368	126 22.600	74.2	65	-	Olive green unconsolidated mud
17-Jan-02	09:13	BBBT-1	C/meiofauna/sed	50 35.375	126 22.597	73.6	30	7.5	Olive green unconsolidated mud
17-Jan-02	09:30	BBBT-1	D/meiofauna/sed	50 35.359	126 22.610	74.6	30	7.5	Olive green unconsolidated mud
17-Jan-02	09:41	BBBT-2	A/macrofauna	50 35.354	126 22.431	71.4	80	7.5	Olive green lumpy mud
17-Jan-02	09:54	BBBT-2	B/macrofauna	50 35.361	126 22.432	71.3	60	-	Olive green unconsolidated mud
17-Jan-02	10:15	BBBT-2	C/meiofauna/sed	50 35.346	126 22.448	70.9	80	7.5	Olive green mud
17-Jan-02	10:42	BBBT-2	D/meiofauna/sed	50 35.348	126 22.464	71.4	78	7	Olive green mud
17-Jan-02	11:01	BBBT-3	A/macrofauna	50 35.296	126 22.421	71.8	55	-	Olive green mud
17-Jan-02	11:10	BBBT-3	B/macrofauna	50 35.262	126 22.430	71.5	55	-	green gray mud
17-Jan-02	12:05	BBBT-3	C/meiofauna/sed	50 35.298	126 22.411	71	30	8	grey-green mud with some slumping
17-Jan-02	12:25	BBBT-3	D/meiofauna/sed	50 35 293	126 22.439	73.4	68	7	Olive green with black streaks

TABLE 16: Percent proportion of grain size categories from sediment samples collected from Bones Bay, January 17, 2002. BBBT = Bones Bay Beam Trawl; 1,2,3 = replicate beam trawl transects; C,D = sediment sample type, see Table 2).

Date	Station	>2mm	<2mm	<1mm	<.5mm	<.25mm	<.10mm	<63 μm	<4 μm	<2 μm
17-Jan-02	BBBT-1 C	4.1	3	4.8	7.9	33	28	9	1.4	8.8
17-Jan-02	BBBT-1 D	-	1	3.2	6.9	49.1	20.4	9.5	1.6	8.3
17-Jan-02	BBBT-2 C	0.5	0	0.6	2	31.8	41.8	11.8	1.2	10.3
17-Jan-02	BBBT-2 D	-	0.1	0.5	1.9	50.6	24.5	13	2.8	6.6
17-Jan-02	BBBT-3 C	5.5	0.6	1.4	3.4	27.5	38.6	9.8	2.4	10.8
17-Jan-02	BBBT-3 D	0.7	0.1	1.1	3.5	51	23.6	10.3	2.5	7.2

TABLE 17: Sediment chlorophyll concentration ($\mu\text{g mL}^{-1}$ sediment) of grab samples from Bones Bay (BB) collected in January 17, 2002.

Date	Location	Core depth (mm)	[chl-a] ($\mu\text{g mL}^{-1}$)	[pheo] ($\mu\text{g mL}^{-1}$)
17-Jan-02	BBBT-1 C	0-5	0.202	1.241
17-Jan-02	BBBT-1 C	5-10	0.281	2.824
17-Jan-02	BBBT-1 C	10-15	0.318	2.904
17-Jan-02	BBBT-1 C	15-20	0.325	2.736
17-Jan-02	BBBT-1 C	20-25	0.281	2.168
17-Jan-02	BBBT-1 D	0-5	0.382	2.985
17-Jan-02	BBBT-1 D	5-10	0.361	2.730
17-Jan-02	BBBT-1 D	10-15	0.346	2.372
17-Jan-02	BBBT-1 D	15-20	0.382	2.118
17-Jan-02	BBBT-1 D	20-25	0.354	2.314
17-Jan-02	BBBT-2 C	0-5	0.397	2.854
17-Jan-02	BBBT-2 C	5-10	0.397	2.220
17-Jan-02	BBBT-2 C	10-15	0.332	2.474
17-Jan-02	BBBT-2 C	15-20	0.325	2.255
17-Jan-02	BBBT-2 C	20-25	0.318	2.343
17-Jan-02	BBBT-2 D	0-5	1.263	8.635
17-Jan-02	BBBT-2 D	5-10	0.390	2.416
17-Jan-02	BBBT-2 D	10-15	0.332	2.277
17-Jan-02	BBBT-2 D	15-20	0.318	2.146
17-Jan-02	BBBT-2 D	20-25	0.274	2.058

TABLE 17: (continued)

Date	Location	Core-depth (mm)	[chl-a] ($\mu\text{g mL}^{-1}$)	[pheo] ($\mu\text{g mL}^{-1}$)
17-Jan-02	BBBT-3 C	0-5	0.411	2.723
17-Jan-02	BBBT-3 C	5-10	0.303	2.197
17-Jan-02	BBBT-3 C	10-15	0.354	2.270
17-Jan-02	BBBT-3 C	15-20	0.368	2.183
17-Jan-02	BBBT-3 C	20-25	0.354	2.044
17-Jan-02	BBBT-3 D	0-5	0.361	2.853
17-Jan-02	BBBT-3 D	5-10	0.267	2.415
17-Jan-02	BBBT-3 D	10-15	0.404	2.380
17-Jan-02	BBBT-3 D	15-20	0.339	2.183
17-Jan-02	BBBT-3 D	20-25	0.354	2.248

TABLE 18: Bottom water characteristics at oceanographic station near Turnour Bay and Bones Bay in 2001 and 2002. NA = not available (Troffe et al. 2005)

Variable	Sept. 27, 2001	Jan 25, 2002	May 21, 2002	Dec 14, 2002
Temperature ($^{\circ}\text{C}$)	8.4 / 8.4	7.5 / 7.7	7.7 / 7.7	7.9 / 7.9
Salinity (psu)	31.31 / 31.49	30.63 / 30.74	31.04 / 31.16	31.28 / 31.39
Dissolved oxygen (mg L^{-1})	2.76 / 2.85	5.43 / 4.87	NA / 5.97	3.61 / 3.56

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6.0 LITERATURE CITED

Arar, E.J., and G.B. Collins. 1997. U.S. Environmental Protection Agency Method 445.0. *In vitro* determination of chlorophyll-a and pheophytin in marine and freshwater algae by fluorescence, revision 1.2: Cincinnati, Ohio, U.S. Environmental Protection Agency National Exposure Research Laboratory, Office of Research and Development.

McKeague, J.A. 1978. Manual of Soil Sampling and Methods of Analysis. Canadian Society of Soil Science.

Ong, S., C.D. Levings, T.F. Sutherland, G.E. Piercey, V. Keong, and R. Davis. 2002. Data record on trawling and trapping effects on humpback shrimp bycatch organisms in Simoom Sound and Northumberland Channel. Can. Data Rep. Fish. Aquat. Sci. 1084: 47 p.

Sutherland, T.F., Levings, C.D., Petersen, S.A., Poon, P., McDermid, M., Byers, S.C., and Piercey, G.E. 2023. Sediment attributes and infaunal abundance in Simoom Sound, British Columbia, Canada. Can. Data Rep. Fish. Aquat. Sci. 1366: viii + 26 p.

Troffe, P.M., C.D. Levings, G. (Beth) E. Piercey and V. Keong. 2005. Fishing gear effects and ecology of the sea whip (*Halipteris willemoesi* (Cnidaria: Octocorallia: Pennatulacea)) in British Columbia, Canada: preliminary observations. Aquatic Conserv. Mar. Freshwat. Ecosyst. 15: 523-533.

Troffe, P.M., C.D. Levings, T.F. Sutherland, V. Keong, and G.E. Piercey. 2003. Data report on beam trawl and prawn trap catches in Clio Channel, British Columbia, 2002. Can. Data Rep. Fish. Aquat. Sci. 1115: 21 p.